UNDERSTANDING NEURODEGENERATION AND RESCUING PATHOLOGY ASSOCIATED WITH PARKINSON'S DISEASE IN *DROSOPHILA* MODEL

Thesis

Submitted to

NAGALAND UNIVERSITY

(A Central University)

In fulfillment of requirements for the Degree

of

DOCTOR OF PHILOSOPHY IN ZOOLOGY

By

Mr. LIMAMANEN PHOM Registration No: 565/2014



DEPARTMENT OF ZOOLOGY NAGALAND UNIVERSITY LUMAMI-798627 NAGALAND, INDIA

2018



(A Central University) OFFICE OF THE HEAD OF THE DEPARTMENT DEPARTMENT OF ZOOLOGY

Headquaters: LUMAMI, ZUNHEBOTO DISTRICT-798627 NAGALAND, INDIA Email: yschandrays@gmail.com Tel: +91-9402908988 (mobile)

Prof. Sarat Chandra Yenisetti, Ph.D., PDF (NINDS/NIH, USA and Univ. Regensberg, Germany)

CERTIFICATE

This is to certify that the Thesis entitled "Understanding Neurodegeneration and Rescuing Pathology Associated with Parkinson's Disease in *Drosophila* Model" is a record of original research work done by Mr. Limamanen Phom under my supervision. He is a registered research scholar (Regd. No.565/2014) of the Department and has fulfilled all the requirements of Ph.D. regulations of Nagaland University for the submission of the Thesis. The work is original and neither the Thesis nor any part of it has been submitted elsewhere for the award of any degree or distinctions. The Thesis is therefore, forwarded for adjudication and consideration for the award of degree of Doctor of Philosophy in Zoology under Nagaland University.

Date: -11-2018

Place: Lumami

Supervisor

inh Yenisetti (Dr.Sarat C.

Professor and Head of the Department

বিদ্যাশ্যায্যস্থা / Head সাणি বিজ্ঞান বিদ্যাশ / Department of Zoology নাশার্লিण্ড বিষরবিদ্যালয় / Nageland University ন্ত্রদানা / Lumami- 798 627



DECLARATION

I, Mr. Limamanen Phom hereby declare that the subject matter of this Thesis is the record of work done by me, that the contents of this Thesis did not form basis of the award of any previous degree to me or to the best of my knowledge to anybody else, and that the Thesis has not been submitted by me for any research degree in any other University.

This is being submitted to Nagaland University for the degree of Doctor of Philosophy in Zoology.

Lingmanen Phon (Candidate)

Y, Sanat Chy Supervisor) 11-2018

Dr. Sarat Chandra Yenisetti, Ph.D Professor Department of Zoology Nagaland University Lumami - 798627 Nagaland, India.

ACKNOWLEDGEMENTS

All glory to God. I express immense joy and gratitude to my supervisor Dr. Sarat Chandra Yenisetti, Professor and Head, Department of Zoology, Nagaland University, under whose inspiring guidance this research work has been successfully carried out. Without his encouragement, constructive criticism and careful supervision, the completion of this Thesis work would not have been possible. As my supervisor, his concern has always been for my welfare and I am fortunate to have worked under his direction. Today as I look back to where I began, I must mention that learning under his guidance, I move ahead with confidence in the field of science.

I am extremely thankful to all the teaching faculties of the Department of Zoology for their valuable suggestions and encouragement. I also express my gratitude to Ms. Zevelou, Ms. Priyanka, Mr. Abhik, Ms. Nukshimenla, Ms. Abuno, Mr. Ayajuddin, Mr. Rahul and Sir. Rajesh, for their cooperation. I am thankful to all my friends for their moral support. I express my gratefulness to my parents, uncle and aunt, brothers and sister for constant encouragement and support in prayer.

Sincere THANKS to-

Department of Biotechnology (DBT), New Delhi Government of India, for all the financial support as Research Fellowship.

Indian Council of Medical Research (ICMR), New Delhi Government of India, for Senior Research Fellowship.

Indian Council of Medical Research (ICMR), New Delhi Government of India, for the financial support as Young Scientist International Travel Grant Award to attend Conference in Frankfurt, Germany during 11-13 August, 2014.

Department of Biotechnology (DBT), New Delhi Government of India, for the opportunity to attend the Japan-Asia Youth Exchange Program in Science at National Institute of Advanced Industrial Science and Technology Biomedical Research Institute, Tsukuba, Japan during 14-22 October, 2018.

All the authors for accepting my request to reproduce the figures from their manuscripts.

(Limamanen Phom)

DEDICATED TO

My Supervisor, with whom this work was successfully carried out.

My family members, who supported and prayed for me throughout the years of the study.

CONTENTS

List of Contents	Page No.
List of Figures and Tables	i-ii
List of Abbreviations	iii-iv
Preface	v-vii
Chapter I	1-37
Introduction and Review of Literature	
Chapter II	38-61
Developing Adult Stage Specific <i>Drosophila</i> Model of Sporadic Parkinson's	
Disease	00.51
Introduction	38-51
Materials and Methods	52-53
Results	54-59
Discussion	60-61
Chapter III	62-78
Assessing the Toxicity of Curcumin in <i>Drosophila</i> Model	<o. 70<="" td=""></o.>
Introduction	62-72
Materials and Methods	72-73
Results	74-76
Discussion	77-78
Chapter IV	79-108
Deciphering the Neuroprotective Efficacy of Curcumin in <i>Drosophila</i> Model of Parkinson's Disease	
Introduction	79-84
Materials and Methods	84-86
Results	87-104
Discussion	105-108
Chapter V	109-153
Mechanistic Insights into the Therapeutic Propensity of Curcumin in	
Drosophila Model of Parkinson's Disease	
Introduction	109-124
Materials and Methods	125-131
Results	132-146
Discussion	147-153
Summary	154-158
References	159-181
Conference/Seminar/Workshop attended	182
Honors and Awards	183
List of Publications	184

LIST OF FIGURES AND TABLES

	Chapter 1	Page No.
Figure 1	Suggested physiological processes related to pathogenesis of Parkinson's disease	10
Figure 2 Table 1	Potential etiological factors of Sporadic Parkinson's disease Monogenetic forms of PD and its fly homolog(s)	21 22-23
	Chapter 2	
Figure 1	Schematic representation of an adult fly brain with the distribution of DA neuron clusters	42
Figure 2	Molecular events related to PQ induced damage	51
Figure 3	Set up for Negative geotaxis assay	53
Figure 4	Survival curve of Oregon K male flies in standard culture media	55
Figure 5	The mortality curve of <i>Drosophila melanogaster</i> in various concentrations of PQ	57
Figure 6	Negative geotaxis assay performed on (a) Health stage (4-5 days old) and (b) Transition stage (55 days old) of male fly	59
Table 1	Parkinsonian genes and their fly homologues	43
	Chapter 3	
Figure 1	The curcumin toxicity in health stage (4-5 days old) of fly	75
Figure 2	The curcumin toxicity curve in transition stage (55 days old) of fly	76
Table 1	Commercial formulation and supplements of Curcumim	66
Table 2	Successful trials completed in Curcumin	69
	Chapter 4	
Figure 1	The survival curve of co-treatment in 4 days old flies (health stage)	88
Figure 2	Negative geotaxis assay for co-treatment regime in 4 days old flies (health stage)	89
Figure 3	The survival curve of pre-treatment in 4 days old flies (health stage)	90
Figure 4	Negative geotaxis assay for pre-treatment regime in 4 days old flies (health stage)	91
Figure 5	The survival curve of co-treatment in 30 days old flies (health stage)	93
Figure 6	Negative geotaxis assay for co-treatment regime in 30 days old flies (health stage)	94
Figure 7	The survival curve of pre-treatment in 30 days old flies (health stage)	95
Figure 8	Negative geotaxis assay for pre-treatment regime in 30 day old flies (health stage)	96
Figure 9	The survival curve of co-treatment in transition stage (55 days old flies)	98
Figure 10	Negative geotaxis assay for co-treatment regime in transition stage (55 days old flies)	99
Figure 11	The survival curve of pre-treatment in transition stage (55 days old flies)	100
Figure 12	Negative geotaxis assay for pre-treatment regime in transition phase (55 days old flies)	101
Figure13	Quantification of brain dopamine level with high-performance liquid chromatography	103
Figure 14	Relative dopamine levels in 5 days (health stage) and 55 days (transition phase) old fly brains exposed to 10mM PQ for 24hrs	104

Chapter 5

Figure 1	The structure of carbonyl derivatives produced by direct oxidation of amino acid side chains	113
Figure 2	A summary of the reactions of protein hydroperoxides and their potential consequences in vivo	114
Figure 3	Mechanisms leading to oxidative stress in PD and the role of PD-related gene products in this process	120
Figure 4	Measurement of ROS levels in (a) Health stage and (b) Transition stage	137
Figure 5	Measurement of MDA levels in (a) Health stage and (b) Transition stage	138
Figure 6	Measure of Protein Carbonyl levels in (a) Health stage and (b) Transition stage	139
Figure 7	Measurement of HP levels in (a) Health stage and (b) Transition stage	140
Figure 8	Measurement of SOD activity in (a) Health stage and (b) Transition stage	141
Figure 9	Measurement of Catalase activity in (a) Health stage and (b) Transition stage	142
Figure 10	Measurement of GST level in (a) Health stage and (b) Transition stage	143
Figure 11	Measurement of GSH levels in (a) Health stage and (b) Transition stage	144
Figure 12	Measurement of total Thiols level in (a) Health stage and (b) Transition stage	145
Figure 13	Measurement of AChE activity in (a) Health stage and (b) Transition stage	146
Table 1	Listing of pathways through which neuroprotective compounds confer neuroprotection: Lessons from <i>Drosophila</i> model of PD	122

LIST OF ABBREVIATIONS

AADC	:	Aromatic amino decarboxylase
AchE	:	Acetylcholinesterase
Acp		Acyl carrier protein
Ad		Alzheimer's disease
AD	•	Autosomal dominant
ALS	•	Amyotrophic Lateral Sclerosis
AR	•	Autosomal recessive
AR-JP	•	Autosomal recessive-juvenile Parkinsonism
ATP132A	•	ATPase cation transporting 132A
ATXN2	•	Ataxin-2
CAT	•	Catalase
CNS	•	Central nervous system
DA	•	Dopamine
DAergic	•	Dopaminergic
DAMB	•	D1-like dopamine receptor
DAND DJ-1	•	Daisuke-Junko-1
DJ-1 EIF4G1	•	Eukaryotic translation initiation factor 4 gamma 1
GSH	•	Glutathione
GST	•	Glutathion S-transferase
HD	•	Huntington's disease
	•	-
HP <i>HtrA2</i>	•	Hydroperoxides
HITAZ HTT	•	HtrA serine peptidase 2
	•	Huntingtin
K	:	Curcumin
LB	:	Lewy bodies
L-dopa	:	Levodopa
LN	:	Lewy neurites
LP	:	Lipid peroxides
LRRK2	:	Leucine-rich repeat kinase 2
MDA	:	Malondialdehyde
Mn	:	Manganese
MPTP	:	1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine
NDD	:	Neurodegeneration disease
NMS	:	Non-motor symptoms
OS	:	Oxidative Stress
PARK16	:	Parkinson disease 16 (susceptibility)
PARKIN	:	Parkin RBR E3 ubiquitin protein ligase
PC	:	Protein carbonyls
PD	:	Parkinson's disease
PINK1	:	Pten induced putative kinase 1
PLA2G6	:	Phospholipase A2 group VI
PQ	:	Paraquat, 1,1'-dimethyl-4,4'-pyridinium

PTEN	:	Phosphatase and tensin homolog
REM	:	Rapid eye movement
ROS	:	Reactive oxygen species
SN	:	Substantia nigra
SNCA	:	α-synuclein
SN <i>pc</i>	:	Substantia nigra pars compacta
TH	:	Tyrosine hydroxylase
UCH-L1	:	Ubiquitin carboxyl-terminal hydroxylase L1
VPS	:	Vacuolar protein sorting
VPS35	:	Vacuolar protein sorting-associated protein 35
α-synuclein	:	Alpha-synuclein

PREFACE

Parkinson's disease (PD) is the second most common neurodegenerative disease and it is characterized by degeneration of dopaminergic neurons in the substantia nigra of mid brain that lead to impairments of motor functions (Cacabelos *et al*, 2017). The loss of dopaminergic neurons give rise to motor symptoms like bradykinesia, rest tremor, postural instability, and shuffling gait; non-motor symptoms like impaired olfaction, constipation, depression, increased daytime sleep, rapid eye movement sleep disorder, and behavioral deficits are also commonly observed (Saleem *et al*, 2013).

Presently, the therapeutic strategies for PD include Deep Brain Stimulation and Levo-dopa supplementation. However the medication is limited only to improve the progressing symptoms and that too with more of side effects (Zrinzo *et al*, 2012; Fahn *et al*, 2000) and there is no therapy available that will cure the disease. Developing a therapeutic strategy for neurodegenerative disease such as PD remains a challenge till date. While attempting to understand the PD progression researchers have developed several animal models including *Drosophila* model. A suitable model for PD should show histopathologically characterizable progressive loss of dopamine neurons together with other neurons and significant reduction in dopamine level. Since PD is a late onset neurodegenerative disease, the symptoms depiction in the model organism should be in a stage of adulthood equivalent to the age where PD sets in. The model animal should also manifest disease in such a way that it would mimic the PD affected human motor symptoms.

Numerous case studies have reported that the subjects having exposure to pesticides, herbicides showed symptoms similar to Parkinsonism. Laboratory exposure of model organisms to environmental toxins like paraquat (PQ) is therefore productively employed to study the disease progressions. Studies on post-mortem brains from PD patients have implicated the role of oxidative damage in the pathogenesis of PD (Yuan *et al*, 2016; Zeevalk *et al*, 2008; Bosco *et al*, 2006). Accumulation of free radicals and subsequent neurodegeneration in specific brain regions have been proposed as the underlying factors in neurodegenerative diseases such as Alzheimer's and Parkinson's disease (Halliwell, 2006) suggesting that oxidative modifications of enzymes and proteins play a significant role in their pathogenesis.

Several researchers employ natural compounds with neuroprotective properties; try to explore ways for their therapeutic application. Curcumin, a natural active compound present in *Curcuma Longa* L. (Turmeric) has been shown to possess potent neuroprotective properties. It is largely used in food as spices, coloring agent, and traditional medicines in India, South Korea, China (Aggarwal *et al*, 2007) and properties of curcumin performing neuroprotective effect, anti-oxidant, anti-inflammatory and anti-cancer are well known. It crosses the blood-brain barrier and exerts protective action on neurons in central neurological disease (Hagl *et al*, 2015; Lee *et al*, 2013). In *Drosophila* model, curcumin has been shown to extend life span, sequester oxidative stress mediated free radicals, enhance locomotor ability and show chemo preventive property, improves characteristic symptoms associated with PD (Nguyen *et al*, 2018; Liu *et al*, 2013; Lee *et al*, 2010) suggesting its potential use in treatment applicability in higher organisms.

However, available investigations were performed in young model organisms. It is reported that there exists significant change of about 23% in genome-wide transcript profiles with age in *Drosophila* (Pletcher *et al*, 2002) and genotropic drugs would be effective only during those life cycle stages wherein target molecules are available (Soh *et al*, 2013), suggesting that targets of genotropic compounds under study may well not be present in all life stages. In spite of these important studies, no reports are available

regarding the efficacy of curcumin in PD models during later phases of adult life. Therefore, it is necessary to understand the neuroprotective efficacy of compounds at the adult phases like the transition phase in *Drosophila*, where the disease such as idiopathic PD sets in. Further, it is equally important to decipher mechanistic insights of these natural products in adult life in a stage specific manner.

A hard effort is made in this direction using Drosophila model of PD.

CHAPTER I

INTRODUCTION AND REVIEW OF LITERATURE

1. Introduction

Neurodegeneration refers to the progressive loss of structure or function of neurons that lead to severe neurological conditions. Neurodegenerative diseases (NDD) are a varied assortment of central nervous system (CNS) disorders accompanied by gradual deterioration in a person's cognitive abilities such as memory, characterized by the progressive loss of neural tissues (Ramanan and Saykin, 2013). This loss is due to either structural change that prevents neurons of brain cells from functioning normally, or to cell death. The most notable NDD are Alzheimer's disease (Ad), Parkinson's disease (PD), Huntington's disease (HD), and amyotrophic lateral sclerosis (ALS). NDD pose an important threat to human health, as these age-dependent disorders are becoming more and more prevalent partly due to the increasing number of elderly populations in recent years (Heemels, 2016). Genetics and environmental factors and the aging process are believed to play important role, although neurodegeneration is frequently multifactorial in origin and the involvement of genes and environment remains debatable. For neurodegenerative disorders such as Ad and PD, both familial and sporadic forms exist whereas in HD, it is solely genetic in nature (Tan *et al*, 2014).

The main pathological hallmarks of neurodegenerative diseases consist of oxidative stress (OS), proteasomal impairment, mitochondrial dysfunction, and accumulation of abnormal protein aggregates. The cause and effect of inflammation in neurodegeneration are still unknown. There is no complete cure for this disorder since neurons of the CNS cannot regenerate on their own after cell death or damage. Thus, neurodegenerative diseases carry a risk of reduction in lifespan (L'opez-Ot'ın *et al*, 2013). Generally aging is a comprehensive phenomenon resulting from constant physiological degeneration over the lifetime of almost all organisms (Kirkwood and Austad, 2000). However, many studies have reported that the lifespan of patients with neurodegenerative diseases such as Ad, PD,

and HD is decreased (Steenland *et al*, 2010). For example, people with dementia are two to four times more likely to die at a given age than individuals without dementia of the same age (Guehne *et al*, 2005).

The mammalian nervous system is highly complex and a major challenge in understanding and designing therapeutic interventions in NDD is to reduce or eliminate the early symptoms and also to improve the inflammatory responses inside the CNS once the disease sets in. In spite of several years of investigations and scientific advancements in basic and clinical research, etiology and pathogenesis of these diseases still remain vague and neurodegenerative diseases represent a scientific challenge. Developing efficient treatments for such conditions where there are no concrete findings about its causes is a challenging and essential task for the scientific community.

2. Neurodegenerative Diseases

2.1. Alzheimer's Disease

Alzheimer's disease (Ad) is a most common devastating brain disease that generally affects people above the age of 65 years. It is characterized by progressive loss of neurons in the hippocampus and cortex that consequently led to reduction of brain mass. Essentially, the cells in the brain that is necessary for processing, storing and retrieving the neuronal information are destroyed which often is the case in neurodegenerative disorders (Singh, 2012). The outcome of which is manifested by loss of cognitive abilities and behavioral functions including memory, thinking and language skills (O'Brien and Wong, 2011). Ad is correlated with synaptic degeneration and neuronal death in limbic structures, such as the hippocampus and the amygdala, and related area of the cerebral cortex. Sporadic form of Ad wherein the cause is unknown make up majority of the cases and a complex interaction of environmental and genetic factors are believed to be associated in the

development of the disorder. With autosomal dominant (AD) inheritance, about 2-3% of Ad cases are early onset and familial in nature whereas bulk of them are late-onset with unknown cause.

One prominent neuropathological hallmark of Ad is an extensive buildup of extracellular plaques consisting of amyloid- β in the brain cerebral cortex. This accumulation has been reported to commence at 10-20 years prior to start of dementia (Bateman *et al*, 2012; Buchhave *et al*, 2012). It is suggested that at the initial extended period of disease progression, soluble amyloid- β oligomers and amyloid plaques change the nature and role of restricted neuronal circuits and large-scale networks via unsettling the stability of synaptic excitation and inhibition (*E/I* balance) in the brain (Busche *et al*, 2016).

Familial Ad can be caused due to mutations in amyloid precursor protein (APP), which is cleaved successively by α - and β -secretases, and presenilins 1 and 2 (PS1 and PS2), one or other of which is a part of each γ -secretases complex (Walsh and Selkoe, 2007; Selkoe, 1998). Presenilin is involved in regulation of Notch and Wnt signaling system and is responsible for progressive maturation of glia and neurons (Gaiano and Fishell, 2002). β -secretases consecutively cleave the Notch receptor to produce a Notch intracellular domain (Kojro and Fahrenholz, 2005), that in turn activate nuclear genes like hairy and enhancer of split 1 (HES1) and hairy and enhancer of split 5 (HES5) to assist in neurogenesis throughout development process and repairing of damage (Kopan *et al*, 2009). Most forms of sporadic late-onset Ad have complex etiology because environmental or genetic factors alone are not adequate enough for the disease development. Currently the main risk factor in sporadic Ad is acknowledged in the allele e4 of apolipoprotein E (ApoE4) (Holtzman *et al*, 2000). But in most of the late-onset Ad cases, the factors that are responsible for the disease are still unidentified and genetics factor most likely interact with environmental factors or with additional physiologic setting to exert the pathogenic outcome.

3

2.2. Huntington's Disease

Huntington's Disease (HD), another heritable devastating neurodegenerative disorder is caused due to an AD mutation in the Huntingtin (HTT) gene (Pavese et al, 2006, Gusella et al, 1995). At the molecular level, HD is caused by a CAG trinucleotide repeat expansion within exon 1 of the HTT gene. In patients the amount of CAG repeats expands from the normal population range of about 16 and 20 repeats to >35 (Munoz-Sanjuan et al, 2011; Warby et al, 2011), which results in extended polyglutamine region at amino terminal side of translated HTT protein which is linked to protein aggregation and toxicity associated with gain-of-function (Williams et al, 2008). Aggregation of mutated gene leads to neuronal injuries in the medium spiny neurons of the neostriatum and other neurons such as the cortex (Li and Li, 2004). Individuals usually experience progressive loss of muscular functions and decline in cognitive ability, decline in self and spatial alertness, depression, dementia, and high anxiety during the period of about 20 years prior to death. Impaired olfactory function was noticed in patients and pre-symptomatic gene carriers (Mochel et al, 2007). Among the number of mechanisms, a supposed toxic mechanism is because of the presence of toxic oligomers (Sathasivam et al, 2010). HD pathogenesis is very complex although it is monogenic in nature. The HTT interactome consists of proteins that are involved in transcription, preserving DNA integrity, regulation of cellular cycle, signaling and organization, transport and homeostasis of proteins, energy metabolism (Shirasaki et al, 2012). Long-term memory is often unaffected but it usually impairs decision-making ability such as organizing or adapting alternatives, and delay in the acquisition of new motor skills (Craufurd et al, 2002; Folstein, 1990), which becomes worse over time and communication deteriorates more rapidly than understanding. Manic and psychotic symptoms also develop in some HD patients (Folstein, 1990). Prevalence of HD is 4-10 per 100000 in the western world, with many more people at risk of the

disease. Mean age of onset of the disease is 40 years, with death occurring 15-20 years after the diagnosis of the disease (Ross *et al*, 2011). At present, management is limited to controlling chorea, which is the unintentional and uneven mobility of the arms and legs, and controlling mood changing aspects; there is no therapy to completely cure the disease (Munoz-Sanjuan *et al*, 2011).

2.3. Amyotrophic Lateral Sclerosis

ALS is the most common adult-onset motor neuron degenerative disease. It is identified by a progressive loss of motor neurons located in the spinal cord, brain stem and motor cortex which leads to reduced muscular function and ultimately respiratory failure. Normally the disease starts at about 55 years of age and patients on average dies within 3-5 years of diagnosis. About 90% of the reported disorders are sporadic (sALS) with unknown cause and 10% are reported to be familial (fALS) having a genetic component. The understanding of etiology and pathogenesis of the sALS are inadequate and there are no successful treatments accessible at present; as a result there is a high unmet need for novel therapeutic approach (Goyal and Mozaffar, 2014). Patients show the symptoms of progressive muscle degeneration and fatique, increased exhaustion and troubles with swallowing, which normally lead to respiratory malfunction and death (Sejvar *et al*, 2005; Rowland, 1994). Progressive functional deficits lead to a complete loss of independence (Ng et al, 2011), and individual become solely dependent on care and support. About 20% of familial cases happen due to mutations in the gene that code for the Cu/Zn-superoxide dismutase (SOD1) (Rosen et al, 1993). Despite of the difference in causal factor, approximately 95% of affected individuals are reported to follow similar pattern of molecular pathology, cytoplasmic buildup ubiquitinated, that includes of hyperphosphorylated and non-soluble TAR DNA-binding Protein 43 kDa (TDP-43

protein) aggregates and reduced levels of nuclear TDP-43 (Brettschneider *et al*, 2013; Geser *et al*, 2008; Mackenzie *et al*, 2007; Neumann *et al*, 2006).

2.4. Parkinson's Disease

Parkinson's disease (PD) is the second most common NDD and it is characterized by degeneration of dopaminergic (DAergic) neurons in the substantia nigra (SN) of mid brain that lead to impairments of motor functions (Cacabelos 2017; De Silva *et al*, 2000). Progressive loss of DAergic neurons and intraneuronal protein inclusions, called Lewy bodies (LB) are characteristic of PD (Olanow and Tatton, 1999; Lang and Lozano, 1998). The protein α -synuclein normally found in presynaptic terminals and nuclei are the most abundant in LBs. The deposition of LB and neurites has been demonstrated to occur years before the degeneration of the substantia nigra *pars compacta* (SN*pc*) and appearance of Parkinsonism (Braak *et al*, 2000).

The neuronal loss in this region gives rise to motor symptoms like bradykinesia, resting tremor, postural instability, and gait impairment where non-motor symptoms (NMS) like impaired olfaction, constipation, depression, increased daytime sleep, rapid eye movement sleep disorder, and behavioral deficits are commonly observed (Saleem *et al*, 2013). During the time of diagnosis, about 50% of the DAergic neurons have already been degenerated. Consequently, the main focus for effective therapies would be to sustain the remaining neurons. The majority of PD cases are sporadic with unknown cause but few of the cases are familial, inherited as a Mendelian trait. Medication of L-dopa, which is a precursor of dopamine (DA), is the existing treatment for PD. However, as the therapy progress, the effectiveness of L-dopa decline over the years following which the patient is required to consume DA agonists and activate the DA receptor. Compounds that slow down DA degrading enzymes are being tested in PD. Most of the therapeutic approach

for PD is aimed to increase or sustain the existing DA pool instead of finding means to inhibit the continuous neuronal loss. There is a need for additional approach that will sustain the neurons coupled with DA replacement therapy.

Current research focuses on developing model that reproduces the α -synucleinopathy of nigrostriatal pathway (Maries *et al*, 2003). However, it is vastly dependent on the capability to either overexpress different α -synuclein protein species locally in the SN or create relevant transgenic animal models (Niu *et al*, 2015). The whole picture of PD pathology is far more complicated than a DA depletion associated motor defect, because, the neurodegenerative progression is not limited to effect on the nigrostraital DAergic pathway but it is also involved in alteration of glutamatergic, serotonergic, noradrenergic, GABAergic and cholinergic systems, along with the deterioration of other DAergic structures in addition to SN*pc*, such as ventral tegmental area (Brichta *et al*, 2013).

2.4.1. Brief History of PD

PD is a disorder which is recognized since the ancient times in Indian and Chinese civilization (Manyam, 1990). In the Ayurveda, an ancient Indian medicine, the condition is termed as *Kampavata* ("*kampa*" means tremor in Sanskrit). An Egyptian papyrus during the 12th century BC recorded about a king drooling with age (Gracia Ruiz, 2004). In Western health and clinical text, the shaky sign was elaborated by a doctor named Galen in 175 AD. In 1817, James Parkinson elaborated in his description of the disorder considering the six cases as the basis which he had noticed during his own practice and observation around his neighborhood. The description about the condition was to propose and persuade others to study the disease in more detail for better understanding. This made to establish the disorder as an accepted medical condition. He named it as "*shaking palsy* or *paralysis agitans*" and wrote thorough descriptions in "An Essay on the Shaking Palsy".

controls over muscular activity at rest and including during on a support; with an inclination to turn the body trunk frontwards, and to go by a walking to a running speed: the mind and cognitive ability remain unharmed'' (Parkinson, 1817).

Later in 1865, William Rutherford Sanders coined the term "Parkinson's disease" and it was then popularized through the works of French neurologist Jean Martin Charcot with his colleagues (Lees, 2007). They narrated symptoms of the disorder considering two criterias: tremor and rigidity. They explained in detail the arthritic changes, dysautonomia, and pain that can come along with the disease but documented that individuals having PD may not be noticeably weak and tremor is not an essential symptom (Charcot, 1872).

2.4.2. Pathophysiology of PD

PD is the second most prevalent neurodegenerative disorder that affects about 1% of the population over age 50 (Modi *et al*, 2016). An essential pathological characteristic of PD is the selective degeneration of DA neurons in the SN*pc*. The DAergic neurons are necessary for accurate functioning of muscle and coordination; their loss is associated with tremor, rigidity, bradykinesia, and postural instability. A second neuropathological hallmark of PD is the formation of proteinaceous LB inclusion in the SN of mid brain which is reported to be full of a protein combination such as *a-synuclein*, ubiquitin, *parkin*, and neurofilaments (Jellinger, 2009). The mode of progression where *a-synuclein* and additional proteins combine for generating Lewy pathology is not well understood, however it possibly involve oxidative alteration and/or cross-linking. Though, degeneration of neurons appears first in the DAergic cell of SN*pc*, degeneration of non-DAergic neurons also take place in later stage of PD (Hornykiewicz and Kish, 1987). The cholinergic nucleus basalis of Meynert, the raphe nucleus serotoninergic neurons, and the hypotralamus undergo neuronal loss as the disease advance.

The DAergic neurons are necessary to perform right motor function and its deficiency or loss led to manifestation of popular condition called bradykinesia, tremors and rigidity which remain the characteristic features of PD. Also, hippocampal and cortical neurodegeneration can add up to dementia which is frequently linked with PD.

Several non-motor symptoms associated with PD are sleep instability, constipation, turning off cognitive ability, depression, fright, nervousness, bladder troubles, weight fluctuation, weariness, hypotension and sexual dysfunction. Such weakening conditions greatly affect the patient's quality of life (Pfeiffer, 2016) and with the passage of time as the disease and motor symptoms advance, the patient lose independence and cannot carry out basic function, thus they become fully dependent on care and support. The present available treatment methods deal with controlling the symptoms but by and large it cannot halt the disease development and incidence of increased mortality among PD patients still persist (Levy *et al*, 2002).

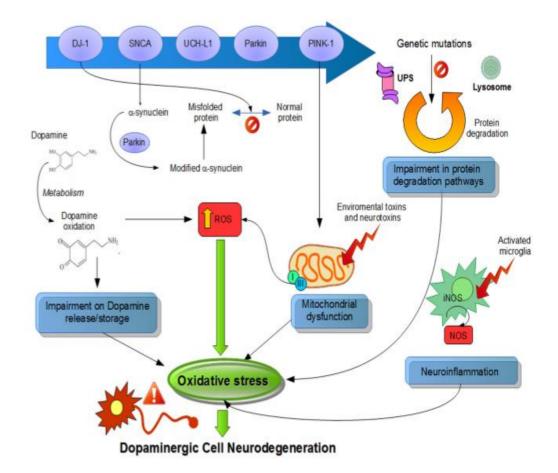


Figure.1. Suggested physiological processes related to pathogenesis of Parkinson's disease

Genetic alteration through various external influences affects pathways and lead to abnormal functions in genes that are associated with PD resulting in augmented OS, mitochondrial dysfunction and protein misfolding. Further neurotransmitter DA metabolism can be oxidized to reactive DA quinines that will contribute to high ROS. Conformational change in α -synuclein protein and its aggregation takes place. ROS affect the normal functioning of UPS resulting in decreased degradation of damaged and misfolded proteins consequently impairing the cell homeostasis and inducing cell death. Environmental stressors affect usual functioning of mitochondria, amplify the reactive free radicals and lead to accumulation of proteins like α -synuclein. Inhibition of mitochondrial complex I lead to decreased ATP synthesis which results in injury to intracellular mechanism and cell death. Furthermore, neuroinflammatory mechanisms might add up to process that cause the cells more susceptible to degenerate. All these numerous cellular mechanisms recognized to induce OS are implicated in the selective degeneration of DAergic neurons (Adapted from Blesa *et al*, 2015).

3. Symptoms of PD

3.1. Motor symptoms in PD

PD is associated with the following motor symptoms like resting tremor, bradykinesia or sluggish movements, stiffness, shuffling gait, postural unsteadiness. The setting in of PD is insidious where affected persons may point the symptoms to aging processes. The symptoms progresses over time however the speed at which motor problem develop are extremely variable (Fritsch *et al*, 2012).

Apart from the "classic" movement disorders that are previously described, there are several additional motor symptoms observed among the patients. They are masked facial expression (hypomimia), reduced eye blinking, hazy vision, lame upward gaze, dystonia, bent position, trouble rotating in bed, kyphosis, scoliosis, freezing and difficulty in verbal communication, such as hypophonia (progressively softer voice), or palilalia (recurrence of word or phrase) (Chou, 2013). One of the worst experiences the PD patients come across in severe condition is they lose the ability to do even basic works and become fully dependent on someone for support.

3.2. Non-motor symptoms in PD

Non-motor symptoms (NMS) are also widespread among PD patients. Independent studies have shown no less than one NMS in nearly all of the affected individuals (Kim *et al*, 2013; Krishnan *et al*, 2011). Additionally, NMS were reported among all the PD patients who were also under unstable motor ability (Witjas *et al*, 2002). Comparatively, aged individuals will experience atleast one NMS and thus among individuals without PD, it occurs normally in the process of aging. Therefore, it may be irrelevant to attribute all NMS experienced by PD subjects as caused by the PD (Kim *et al*, 2013; Krishnan *et al*, 2011). However, it is also reported that, PD patients have higher tendency to develop

diverse form of NMS which is often accompanied with increased occurrence and severity (Khoo *et al*, 2013).

Abnormalities of sensation: Various sensation abnormalities are reported with the olfactory impairment the most commonly described. Several other issues also arise such as vision and patterns of pain. During the point of diagnosis olfactory impairment is already familiar in about 90% of the cases. But the impaired sense of smell may go unnoticed to over 70% of affected individuals and that the impaired olfaction is independent of disease stage and duration (Doty *et al*, 1988). There are other report suggesting that impairment of olfaction is related to increased severity of the disease; acute hyposmia among PD individual scan signal the progress of PD dementia (Cavaco *et al*, 2015). NMS like olfactory testing are recommended for screening of PD although at the clinical level such testing can take a long time before concluding. As such, researchers conducted a study for just three odors (coffee, peppermint, and anise) and found comparable outcome to test with a complete odor panel (Casjens *et al*, 2013). Positive results in odor detection have been reported through olfactory exercise, but substantiation with further studies is required (Knudsen *et al*, 2015).

Behavioral changes: Depression is a regular phenomenon seen in PD. Occurrence of major depressive disorder in 17%, minor depression in 22% and dysthymia in 13% of individuals with PD have been reported (Riejnders *et al*, 2008). This behavioral change may occur prior to motor impairment as an early indication of PD. Medications including antidepressant are found to be efficient for treatment of Parkinsonian depression and beneficial role of transcranial magnetic stimulation is being developed.

The occurrence of anxiety in PD is 25-40% (Simuni *et al*, 2013). Anxiety may also emerge at any stage of the disease which often develops before the motor dysfunction and

persons with persistent anxiety are at higher risk group to develop PD. Among the PD patients generalized anxiety disorder, panic and one of the phobic disorders are common which may turn into noticeable wearing off occurrence. Recent meta-analysis indicates that increased incidence of apathy among patients is around 40% (den Brock *et al*, 2015) and a connection of apathy with impaired executive function particularly complexity with initiation is clear in PD.

Autonomic dysfunction: One of the common non-motor symptoms in PD is autonomic dysfunction that can be manifested at any point of time of disease and some facets like constipation may be developed with higher incidence earlier than the appearance of motor problem. Orthostatic hypotension which may be present in almost 60% of PD case is largely recognized feature of cardiovascular dysfunction in PD. Gastrointestinal dysfunction in PD may appear as dysphagia, impaired peristalsis, bacterial overgrowth in small intestine, and bowel dysfunction (Fasano *et al*, 2015). PD patients with gastroparesis may experience satiety, low appetite, bloating, abdominal stiffness, nausea, vomiting, and weight loss. Small intestinal bacterial overgrowth is described among PD patients and may be responsible for progression of motor fluctuations in certain case (Fasano *et al*, 2015).

Sleep disturbances: Sleep disturbance or impaired sleep is a common situation that occurs among the PD patients with its prevalence rate reaching about 90% (Kurtis *et al*, 2013). Sleep disturbance with recurrent nocturnal awakenings is one of the most regular forms of insomnia in PD. The basis is multifactorial that is associated with stiffness in sleep rendering difficulty in turning sides in bed. Identification and targeting the source of the problem will result in an effective treatment of this symptom. Substantial interest has generated in rapid eye movement (REM) sleep behavior disorder (RBD) distinguished by constant capacity to move about in REM sleep, both as a symptom and as susceptible factor to develop PD. The occurrence of RBD in PD is not clearly identified and it may possibly be present years before classic motor features of PD appear.

Fatigue: Although very little is known about the mechanism involved and efficient treatment of fatigue is limited, it has been gradually accepted to be involved in PD progression. Fatigue is commonly recognized by individuals having PD as one of the most disabling symptoms that has negative impact on their daily life (Dogan *et al*, 2015).

4. Synthesis and Role of DA

Dr. Carlsson and colleagues showed that DA was a neurotransmitter in the brain and not just a precursor of norepinephrine (Carlsson *et al*, 1957). It is involved in a range of important CNS functions, including voluntary movement, feedback mechanism, sleep and awareness, effective memory and learning. In the periphery, DA act on essential physiological function in the regulation of olfaction, retinal processes, hormonal and sympathetic regulation, cardiovascular functions, immune system and renal functions, among others (McHugh and Buckley, 2015; Iversen and Iversen, 2007; Carlsson, 2001; Sibley, 1999; Missale *et al*, 1998).

DA synthesis in DAergic terminals requires tyrosine hydroxylase (TH) which, in the presence of iron and tetrahydropteridine, oxidizes tyrosine to 3,4-dihydroxyphenylalanine (Levodopa or L-dopa). L-dopa is decarboxylated to DA by aromatic amino acid decarboxylase (AADC), an enzyme which requires pyridoxyl phosphate as a coenzyme. Once released from presynaptic terminals, DA mediates its action by activating members of a family of G protein-coupled DA receptors named D1 to D5. The actions of DA are terminated through presynaptic reuptake. Some of the DA is then re-incorporated into vesicles, while the rest is metabolized.

DA receptors were classified on the basis of physiological (Cools *et al*, 1976), pharmacological and biochemical role they play (Trabucchi *et al*, 1975); alternatively, a dynamic equilibrium of a single DA receptor fluctuating between two state of configuration has been postulated (Creese *et al*, 1975). The demonstration of a DA sensitive adenylyl cyclase not only provided an in vitro model to study the properties of DA receptors but also suggested a mechanism for the generation of the physiological response to the amine (Kebabian *et al*, 1971). However not all the DA receptors have properties similar to those of the receptor linked to the adenylyl cyclase.

In a study employing rat model, fetal DA neurons could survive transplantation from the ventral midbrain to the 6-hydroxy DA lesioned dorsal striatum and could also perform motor functions (Bjorklund *et al*, 1980). This finding resulted in the experimental use of DA grafts in PD patients (Dunnett *et al*, 1999). Additionally, grafting studies in rats established the topographical cortico-striatal makeup of the dorsal striatum and the significance of this region of the DA system for understanding of complex motor behaviors (Dunnett *et al*, 1999; Dobrossy *et al*, 1996).

In *Drosophila*, DA is both a vital neuromodulator and a precursor of molecules that are necessary for hardening and pigmentation of the external cuticle (Riemensperger *et al*, 2011; Friggi-Grelin *et al*, 2003). DA released from fly neurons interacts with specific G protein-coupled DA receptors, either of the D1 or D2 subtypes (Hearn *et al*, 2002; Blenau *et al*, 2001), including dDA1 and D1-like DA receptor (DAMB), which play important function in arousal and memory (Berry *et al*, 2012; Lebestky *et al*, 2009). A long-term over expression of the DA-synthesizing enzyme TH in the nervous system prolongs the survival of adult PQ-intoxicated flies and a down-regulation of the DAMB receptor results in an enhanced PQ resistance in fly. DA signaling modulates OS resistance in the

Drosophila nervous system since an expansion of DAMB expression appears to trigger the age-related increase in PQ susceptibility of young adult flies.

5. Genes involved in DA metabolism and in metabolism of xenobiotics

A number of gene variants which code for proteins that are associated with DA transport and metabolism, metabolism of xenobiotics and in OS response, have mainly been under investigation as a probable cause for PD susceptibility. But results are conflicting or inconclusive for vast majority of them. Several authors support the opinion that there is no concrete result with regard to risk factor of PD because quite a few investigations have suggested relationship between gene polymorphisms and PD, just a handful have studied the gene-environment connection (Mellick, 2006). A large case control study across 5 European centers was conducted with the sole intention to understand the geneenvironment interaction and the risk of developing PD (Dick et al, 2007). Exposure to solvents, pesticides and metals like iron, copper and manganese (Mn) was analyzed and linked to polymorphisms in a number of PD putative risk genes Cytochrome P450 2D6 (CYP2D6), PON1 (Serum paraoxonase/arylesterase 1), GSTM1&3 (Glutathione Stransferase Mu 1 & 3), GSTT1 (Glutathione S-Transferase Theta 1), GSTP1(Glutathione s-transferase pi 1), NQO1 (NAD(P)H Quinone Dehydrogenase 1), CYP1B1 (Cytochrome P450 Family 1 Subfamily B Member 1), MAO-A&B (Monoamine oxidase A&B), SOD2 (Superoxide dismutase 2), EPHX (Epoxide Hydrolase), DAT1 (DA Transporter 1), DRD2 (DA Receptor D2)and NAT2 (N-Acetyltransferase 2). In male subjects, a modest association was found between MAO-A polymorphism and PD susceptibility and a greater part of gene-environment analysis did not show noticeable interaction effects. But, an increased risk of PD was seen among GSTM1 null subjects heavily exposed to solvents. In a pesticide exposed environment GSTP1 polymorphisms have been associated with PD among the population (Menegon et al, 1998), and there are evidence suggesting that herbicide exposure modify the relation between *GSTP1* polymorphisms and PD age at onset (Wilk *et al*, 2006). There is no decisive confirmation found for an overall relationship of N-acetyl transferase 2 slow acetylator genotypes to PD in a meta-analysis of published papers. Also, the *CYP2D6* gene variants have also been broadly measured as genetic risk factors for PD with no concrete outcome. But some data suggests that the PD susceptibility might be modulated by exchange involving the *CYP2D6* genotype and environmental factors including contact with pesticides and cigarette smoking (Elbaz *et al*, 2004).

6. Epidemiological Studies on PD

A case study involving drug user patients was performed in 1980s. Patients showed symptoms of visual hallucinations, limb jerking and stiffness. At the initial use of the synthetic heroin, within 4 to 14 days they experienced a generalized slowness and difficulty in moving. Examination of each subject showed almost complete loss of mobility, noticeable tone increase, failure to converse clearly, fixed glancing, reduced eye blinking, facial seborrhea, regular drooling, responsive to glabellar tap test and cogwheel inflexibility in the upper extremities and one subject exhibited a pill rolling tremor at rest on the right hand. One could perform walking and showed a diminutive stepping, sluggish, shuffling gait and bradykinesia. All the subjects showed a flexed position distinctive of an advanced stage of PD. It was described to be linked with the chemical substance 1methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) and suggested that a product of Meperidine-analog synthesis lead to chronic Parkinsonism in humans (Langston et al, Almost immediately after the discovery of MPTP neurotoxicity, potential 1983). pesticides and herbicides involvement in development of PD was studied since there is close structural resemblance between MPP⁺ (an active toxin, a product of MPTP metabolism) and herbicide 1,1'-dimethyl-4,4'-pyridinium (paraquat). Interestingly, in the

1960s, MPP⁺ itself was experimented as an herbicide under the marketable name of cyperquat (Di Monte, 2002).

Since the Second World War, several pesticides, insecticides, herbicides, fungicides, have been extensivelyin use. Farmers were exposed to pesticides because of direct handling and contact skin, consumption of contaminated food and water or aerial inhaling of sprayed pesticides or fumigants. Later on, these farmers developed Parkinsonian symptom and in the recent decades, relationship between individuals contact with pesticide and frequency of PD occurrence has been well documented and recognized. A meta-analysis of epidemiological studies conducted around the world during 1989-1999 showed the relationship between exposures of pesticides to the incidence of PD (Priyadarshi *et al*, 2000).

A study in Taiwan investigated the role of environmental factors for PD. They considered 120 PD patients and 240 hospital control subjects having age and sex matched. Researchers undertook a standardized interview to gather the subjects' history of exposure to environmental factors. On analysis, the use of pesticides and herbicides in their rural occupational farming were found to be associated with an increased PD risk with highest among those subjects who had used paraquat (PQ) and other herbicide/pesticides. Therefore, concluding that exposure to environmental factors like PQ and other herbicides/pesticides, may take significant part in the developing PD (Liou *et al*, 1997).

Another study from southwestern region of Taiwan investigated whether functional variants of genes interacted with occupational pesticide exposure to increase PD risk. A total of 153 patients with sporadic PD and 155 healthy subjects were genotyped for genetic variants. This study found significant differences in genotypes between PD patients and the control subjects. Besides this the combined Mn dependent superoxide dismutase

18

(MnSOD) and NAD(P)H Quinone Dehydrogenase 1 (NQO1) variant genotype was notably linked to a 4.09-fold increased risk of PD in pesticide exposed subjects (Fong *et al*, 2007), suggesting the role of environmental influence on susceptible genes. Another study conducted in Val Camonica (Italy), a geographic area with higher prevalence of individuals affected by Parkinsonism, probably related to increased exposure to Mn. On investigating whether polymorphism in genes regulating Mn metabolism and toxicity could modify neurophysiological effects of Mn exposure, it was found that Mn exposure significantly impaired motor coordination in the elderly people (Rentschler *et al*, 2012).

PQ exposure in *Drosophila* model shows OS induced neurodegenerative phenotype like motor dysfunction as seen among human PD subjects. It has been observed that *Drosophila* mutant for DA regulating genes show an uneven susceptibility to PQ. Exposure of PQ in *Drosophila DJ-1* mutants lead to impairment of motor ability but found resistant to its toxicity in loss of function of $DJ-1\beta$ mutants and overexpression of $DJ-1\alpha$ in DA neurons confers protection. PQ induced DAergic neuronal loss are protected and confers delay in symptoms when there is mutation in down regulator of DA production Catecholamines-up (Catsup) (Chaudhuri *et al*, 2007). Combined exposure of PQ and Maneb on E1 ligase knockdown *Drosophila* resulted in considerable loss of DA neuron (Martin *et al*, 2014) suggesting that environmental agents like pesticides may act in synergy and develop as risk factor for PD.

7. Sporadic Parkinson's Disease

In sporadic PD there is no known cause and studies implicate the role of environmental factors with susceptible genetic background as one of the main causes of the disease. Sporadic PD pathogenesis is thus believed to have multiple factors which involve interaction between the environment and gene. Environmental factors like neurotoxicants are thought to be involved in the pathogenesis of nigrostriatal degeneration, which support idea that there is relationship between environment agents and PD (Di Monte *et al*, 2002).

Furthermore, the report that mutation in α -synuclein gene causes familial PD (Polymeropoulos *et al*, 1997) and its recognition as a possible risk factor for sporadic form of PD (Simon-Sanchez *et al*, 2009) provides an association of genetic background that causes the disease. Several studies have suggested that among the rural people, regularly consuming well water, exposure to farm pesticides and agricultural farm occupation, mining and welding are more prone to gradually develop PD (Ritz *et al*, 2009; Elbaz *et al*, 2009; Dhillon *et al*, 2008; Kamel *et al*, 2007). Epidemiological studies indicate the link between PD and environmental toxic factors. Further several other results propose that contact with environmental pesticide like bipyridyl, organochlorine, PQ and carbamate derivatives could add to developing PD (Liou *et al*, 1997; Seidler *et al*, 1996).

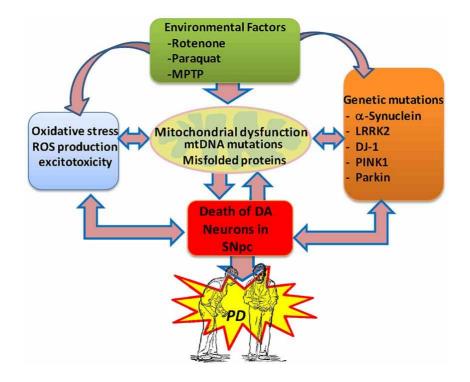


Figure.2. Potential etiological factors of Sporadic Parkinson's disease. Current evidence suggests that environmental factors and genetic factors provoking OS, excitotoxicity and mitochondrial dysfunction in the brain which lead to the degeneration of the midbrain DAergic system, resulting in PD (Adapted from George *et al*, 2014).

ŝ
Gen
letics
of]
Par]
kins
ion'
s Dis
ease

There are about 15 genes identified till now and 21 loci that are associated with genetic form of PD. Human PD genes and their

Drosophila homolog are depicted in Table 1. Some of them are related below.

Confirmed	Kufor-Rakeb syndrome, a form of juvenile-onset atypical Parkinsonism with dementia, spasticity and supranuclear gaze palsy	AR	CG32000	ATP13A2 (Ramirez <i>et al</i> , 2006)	1p36	PARK9
Confirmed	Classical Parkinsonism	AD	Lırık	LRRK2 (Paisán-Ruíz et al, 2004)	12q12	PARK8
Confirmed	Early onset Parkinsonism	AR	Dj-1 α and dj-1 β	PARK7 encoding DJ-1 (Bonifati et al, 2003)	1p36	PARK7
Confirmed	Early onset Parkinsonism	AR	pink1	<i>PINKI</i> (Valente <i>et al</i> , 2004)	1p35-p36	PARK6
Un-confirmed	Classical Parkinsonism	AD	Uch	UCHLI	4p13	PARK5
Erroneous locus (identical to PARK1)	Early-onset Parkinsonism	AD	no homolog	SNCA	4q21-q23	PARK4
Unconfirmed	Classical Parkinsonism	AD	I	Unknown	2p13	PARK3
Confirmed	Early onset Parkinsonism	AR	parkin	PARK2 encoding Parkin (Kitada et al, 1998)	6q25.2-q27	PARK2
Confirmed	Early-onset Parkinsonism	AD	no homolog	SNCA (Polymeropoulos et al, 1997)	4q21-22	PARK1
Status and remarks	Disorder	Inheritance	<i>Drosophila</i> homolog	Gene	Gene locus	Symbol

22

Unknown-Risk factorClassical ParkinsonismHTRA2HtrA2AD or riskClassical ParkinsonismHTRA2HtrA2AD or riskClassical ParkinsonismPLA2G6 (Paisan-Ruiz et al, 2009)iPLA2-VIAAREarly-onset dystonia-PLA2G6 (Paisan-Ruiz et al, 2008)no homologAREarly-onset parkinsoniamPLA2G6 (Paisan-Ruiz et al, 2008)no homologAREarly-onset parkinsonian-PLA2G6 (Paisan-Ruiz et al, 2008)no homologAREarly-onset parkinsonian-PLA2G6 (Paisan-Ruiz et al, 2008)no homologAREarly-onset parkinsoniamFBXO7 (Shojaee et al, 2008)no homologAREarly-onset parkinsoniamFBXO7 (Shojaee et al, 2008)no homologAREarly-onset parkinsonismFBXO7 (Shojaee at al, 2012)vps35ADClassical ParkinsonismEIF4G1eIF4GADLate onset ParkinsonismDNAJC6 (Edvardson et al, 2012)auxillinARParkinsonismEVVIT (Krehk et al 2013: On advisor al 2013)SoniAREarly-onset Parkinsonism
- Risk factor HtrA2 AD or risk factor iPLA2-VIA AR no homolog AR - Risk factor Vps35 AD eIF4G AD AR
-Risk factor-Risk factorHtrA2AD or risk factoriPLA2-VIAARno homologAR-Risk factor-Risk factorVps35ADADAD
-Risk factor-Risk factorHtrA2AD or risk factoriPLA2-VIAARiPLA2-VIAAR-AR-Risk factor-Risk factorVps35AD
- Risk factor AD or risk factor iPLA2-VIA no homolog - Risk factor
- Risk factor HtrA2 AD or risk factor iPLA2-VIA AR no homolog AR
- Risk factor d HtrA2 AD or risk factor factor A iPLA2-VIA AR
ItrA2 AD or risk factor
Risk factor
- AD Late onset Parkinsonism
- Risk factor Classical Parkinsonism

Table: 1. Monogenetic forms of PD and its fly homolog(s). (Adapted from Modi *et al*, 2016).

23

8.1. α-Synuclein (SNCA)

The *SNCA* gene provides instructions for making a small protein called α -synuclein. A missense mutation in the *SNCA* gene was identified in a large Italian family by Polymeropoulos and colleagues. Their study involved a traditional linkage approach through which they could track the underlying genetic damage in the long arm of human chromosome number 4 (Polymeropoulos *et al*, 1996) which brought about a FRAME SHIFT in the PD genetic study. Another study showed that the main composition of LB, pathological hallmark of PD was the α -synuclein protein (Spillantini *et al*, 1998). Thus, these two important scientific results suggested an association of sporadic form with familial forms of PD.

Patients with *SNCA* mutation have early age onset of PD with initially good response to L-dopa treatment however dementia develop in increased pace at regular feature as the disease advance. Then case such as cognitive decline, hallucinations and alteration in self-awareness becomes apparent and upon histopathological investigation reveals a rich LB pathology (Sheerin *et al*, 2014). Of the two-point mutations, H50Q was identified with late onset PD showing a L-dopa responsiveness and cognitive impairment (Proukakis *et al*, 2013; Appel-Cresswell *et al*, 2013). The G51D point mutation which was initially described in French kin having Parkinsonian-pyramidal syndrome was accompanied by an early-onset Parkinsonism with mild to modest response to L-dopa coupled with and fast disease development (Lesage *et al*, 2013). Two additional copies of the genomic region consisting the *SNCA* gene as a basis of PD have been reported. Thus, duplications and triplications of *SNCA* locus serve as a ground of causing PD as well which is more common. In case of triplication, carriers have early-onset disease development whereas *SNCA* duplication patients have later onset of symptoms (Nishioka *et al*, 2006).

8.2. Leucine-rich repeat kinase 2 (*LRRK2*)

The LRRK2 gene provides information for making a protein called dardarin that are involved in kinase and GTPase activity. A study involving a large Japanese family reported link between autosomal-dominant Parkinsonism and chromosome number 12 (Funayama *et al*, 2002). It was then recognized that mutations in *LRRK2* gene results in fundamental genetic cause of chromosome number 12 associated with PD (Paisan-ruiz *et al*, 2004; Zimprich *et al*, 2004). Although numerous *LRRK2* mutations have been reportedly suggested as a cause of PD, only few mutations are found to have high degree of proof. G2019S is the most common mutation of *LRRK2* which is identified in about 1% and 6% case of the sporadic and familial PD respectively (Healy *et al*, 2008). The second frequent *LRRK2* mutation is the R1441G which is very normally found among the Basques (Di Fonzo *et al*, 2006).

8.3. Parkin RBR E3 ubiquitin protein ligase (*PARKIN*)

PARKIN was the second identified PD gene. *PARKIN* plays a role in the cell machinery that degrades unneeded proteins by tagging damaged and excess proteins with molecules called ubiquitin. Initially, a disorder which is denoted by early dystonia and L-dopa medication induced complication, osteotendinous hyper-reflexia with relatively sluggish motor development was reported in Japan in 1973, later it was called the Autosomal-recessive juvenile Parkinsonism (AR-JP) (Khan *et al*, 2002). The condition was recognized as due to mutations in *PARKIN* gene (Kitada *et al*, 1998). It was then suggested that AR-JP locate to long arm of chromosome number 6; associated with pointer *D6S305* and *D6S253* and the former is reported to be deleted in an AR-JP Japanese patient (Matsumine *et al*, 1998). Mutations in *PARKIN* are the main reason of autosomal-recessive (AR) early-onset Parkinsonism. In most of the cases, the disorder is associated with heterozygous *PARKIN* mutations where the

transfer is through dominant mode, thereby indicating that even carting a single *PARKIN* mutation may serve as possible factor for a person and ultimately develop PD (Lohmann *et al*, 2003; Lincoln *et al*, 2003). Clinically, slow disease progression coupled with better response to L-dopa treatment and early dyskinesia has been reported in the profiling of patients carrying the mutated *PARKIN* gene. Uncharacteristic symptoms like key psychiatric manifestations, cerebellar, neuropathy, hyper-reflexia and dystonia signs were also observed (Lohmann *et al*, 2003).

8.4. Daisuke-Junko-1 (*DJ-1*)

DJ1 is involved in cell protection, particularly brain cells, from OS; serve as a chaperone molecule that helps fold newly produced proteins into the proper 3-dimensional shape and helps refold damaged proteins and involved in delivering selected proteins to proteasomes. *DJ1* gene mutation was identified as a basis of AR early onset PD (Bonifati *et al*, 2003) with the locus for this form of PD localized to 1p36 in the Netherlands. This type of mutations is rare and only about 1% *DJ-1* mutation has been identified in early-onset PD individuals (Abou-Sleiman *et al*, 2003). Clinically, the symptoms seen among of *DJ-1* subject are similar to *PARKIN* and *PINK1* associated Parkinsonism (Massano *et al*, 2012).

8.5. PTEN induced putative kinase 1 (*PINK1*)

The *PINK1* gene provides instructions for making a protein called Phosphatase and tensin homolog (PTEN) induced putative kinase 1, which help protect mitochondria from malfunctioning during periods of cellular stress, such as unusually high energy demands. In a study involving Sicilian family with AR Parkinsonism, mutations in the *PINK1* was first recognized (Valente *et al*, 2004). Most of the cases observed were of

missense mutation but copy number mutations, genetic and exonic shifting was also reported (Samaranch *et al*, 2013). Homozygous and compound heterozygous form of *PINK1* mutations have been identified in sporadic and genetic case of PD. This suggests possible part of lone *PINK1* heterozygous mutation as causal factor for PD (Nuytemans *et al*, 2010). *PINK1* mutation is related to early-onset PD in most cases and its connection in sporadic cases is about 2%–4%. Clinically, the phenotype is similar with those patients of *PARKIN* and *DJ-1* mutations. The affected individuals show L-dopa-responsive pattern steadily (Samaranch *et al*, 2013). At neuropathology, neuronal loss in the SN*pc*, LB and aberrant neuritis in the reticular nuclei of the brainstem, SN*pc* and nucleus basalis of Meynert are reported in the *PINK1*-linked PD (Valente *et al*, 2004).

8.6. Vacuolar protein sorting-associated protein 35 (VPS35)

This gene belongs to a group of vacuolar protein sorting (VPS) genes. The encoded protein is a component of a large multimeric complex, termed the retromer complex, involved in retrograde transport of proteins from endosomes to the trans-Golgi network. Two separate studies using exome sequencing described that mutation in *VPS35* causes monogenic form of PD. They detected p.D620N mutation in *VPS35* in Swiss family members with a late-onset, AD PD (Vilarino *et al*, 2011). Later study in a large multigeneration and two Austrian PD families reported the identification of the p.D620N mutation and two other families screened for *VPS35* mutations was published (Zimprich *et al*, 2011) but details of this case are inadequate.

8.7. ATPase cation transporting 13A2 (*ATP13A2*)

This gene encodes a member of the P5 subfamily of ATPases which transports inorganic cations as well as other substrates. Study in consanguineous Jordanian

family initially reported loss of function in a neuronal P-type ATPase gene, *ATP13A2* (Najim al-Din *et al*, 1994). At clinical level, the patients showed an early disease ons*et al*ong with signs of rigid-akinetic phenotype and less trembling at rest, pyramidal syndrome, gradual cognitive dysfunction, vertical gaze palsy, small myoclonus, sleeplessness and L-dopa receptive features (Najim al-Din *et al*, 1994). Loss of function in this gene mapping on chromosome 1p36 lead to an uncommon form of PD recognized as Kufor-Rakeb syndrome (Hampshire *et al*, 2001). Using homozygosity mapping and positional cloning, compound heterozygous mutations in *ATP13A2* have been identified in juvenile Chilean kin (Ramirez *et al*, 2006). Clinical features of such early-onset pallidopyramidal syndrome show varying effect of severity and barely few cases have been identified (Schneider *et al*, 2010; Ning *et al*, 2008).

8.8. Ubiquitin carboxyl-terminal hydrolase L1 (*UCH-L1*)

The UCHL1 gene provides information for making an enzyme called ubiquitin carboxyl-terminal esterase L1. Ubiquitin carboxyl-terminal esterase L1 is possibly involved in the cell machinery that breaks down (degrades) unneeded proteins. In a German family with AD transmission PD, reported a missense mutations of gene that code for the ubiquitin carboxyl-terminal hydrolase L1 (UCH-L1), a ubiquitin recovery enzyme found on chromosome 4p14 (Leroy *et al*, 1998). The subjects show phenotype similar to those of idiopathic PD patients. Later a connection between the UCH-L1 gene S18Y variant and PD (Ragland *et al*, 2009) was established through an epidemiological study. Overexpression of UCH-L1 gene causes upregulation in formation of aggresomes by impairment of proteasome system (Ardley *et al*, 2004).

8.9. *HtrA* serine peptidase 2(*HtrA2*)

This gene encodes a serine protease, a protein that is found in the endoplasmic reticulum and interacts with an alternatively spliced form of mitogen-activated protein kinase 14. The protein has also been localized to the mitochondria with release to the cytosol following apoptotic stimulus. The protein is thought to induce apoptosis by binding the apoptosis inhibitory protein. Loss of function of the gene coding for *Omi/HtrA2* (high temperature requiring A2 mitochondrial protein) have been suggested to be one of the risk factors for PD through several studies employing German (Strauss *et al*, 2005) and Belgian PD patients (Bogaerts *et al*, 2008). At molecular level, mitochondria release the *Omi/HtrA2* and promote apoptosis and mutations in *Omi/HtrA2* gene interrupt its protease action associated to mitochondrial impairment (Bogaerts *et al*, 2008; Strauss *et al*, 2005). *Omi/HtrA2* functions on *PINK1/PARKIN* pathway downstream of *PINK1* and perform function free from *Parkin* influence (Whitworth *et al*, 2008). However, apart from *PINK1* or *PARKIN* null mutants, this is not established in *Omi/HtrA2* knock-out mutants (Yun *et al*, 2008).

8.10. Eukaryotic translation initiation factor 4 gamma 1 (*EIF4G1*)

The protein encoded by this gene is a component of the multi-subunit protein complex *EIF4F*. This complex facilitates the recruitment of mRNA to the ribosome, which is a rate-limiting step during the initiation phase of protein synthesis. In chromosome 3q26-q28 of a northern French family with AD late-onset Parkinsonism, a mutations in the eukaryotic translation initiation factor 4-gamma (*EIF4G1*) as a causal factor was reported. Analysis recognized a heterozygous mutation in *EIF4G1* that was established afterwards in LB disease patients and several other PD subjects (Chartier-Harlin *et al*, 2009). Additionally, in a study on a different multiplex separate family,

a pathogenic mutation was detected among all the affected members along with an unaffected 86-year-old kin indicative of partial penetrance (Nuytemans *et al*, 2013).

8.11. Phospholipase A2 group VI (*PLA2G6*)

The *PLA2G6* gene provides instructions for making a type of enzyme called an A2 phospholipase that is involved in breaking down (metabolizing) fats called phospholipids. In an investigation employing three patients of two inbred Pakistani families, a homozygous mutation was reported in phospholipase A2 gene (*PLA2G6*) found on chromosome 22q13.1. Their phenotype was related to cognition and psychiatric difficulty and dystonic features, pyramidal syndrome (Paisan Ruiz *et al*, 2009). Frontotemporal lobar atrophy and dementia were prominent in compound heterozygous mutations in *PLA2G6* induced early-onset recessive Parkinsonism in a study involving an Asian group (Yoshino *et al*, 2010).

8.12. Parkinson disease 16 (susceptibility) (PARK16)

Following genome wide association and a double replication studies, this locus was identified in chromosome 1q32 in Japanese patients (Satake *et al*, 2009). It was replicated in Caucasians and Han population (Chang *et al*, 2013; Simon-Sanchez *et al*, 2009). Evidence of association with PD was found in a case control Scandinavian study for a coding variant that is located around 5' region of RAB7L1 (Pihlstrom *et al*, 2015). In PD patients of United Kingdom other variants were also identified within the *SLC41A1* gene (Tucci *et al*, 2010) although it was not replicated in another study of PD patients from Spain (Mata *et al*, 2011).

8.13. Spinocerebellar ataxias type 2 (*ATXN2*)

ATXN2 gene encodes Ataxin-2 protein involved in epidermal growth factor receptor (EGFR) trafficking, acting as negative regulator of endocytic EGFR internalization at the plasma membrane. L-dopa responsive Parkinsonism identified in a large Asian-American family showed that the cause of disease was due to an expansion mutation in *ATXN2*, the cause of spinocerebellar ataxias type 2 (SCA2) (Gwinn-Hardy *et al*, 2000). Two out of the three affected family members met the PD Society Brain Bank and NINDS (National Institute of Neurological Disorders and Stroke) norms for diagnosis of PD. The other member was also diagnosed under PD by their physician, although the patient did not satisfy the norms. This study was briefly followed by a number of independent results with parallel conclusion (Furtado *et al*, 2004). Patients may present both L-dopa responsive Parkinsonism and also show features like ataxia, disease reminiscent of advancing supranuclear palsy, trembling and dementia. A comprehensive neuropathologic description on patients with *ATXN2* expansion mutation and L-dopa-responsive Parkinsonism is lacking.

9. Animal Models of PD

Animal models allow researchers to recapitulate some of the clinical manifestation of the diseases of humans hence giving an optimal opportunity to better understand the mechanism involve in the disease progression. Though model animal shares several molecular and cellular processes common to humans there exist limitations too. Complex human behaviors such as aggression, circadian rhythms, sleep, learning, memory and mating are observed in other animal as well such as *Drosophila*. Therefore, by investigating these processes and behaviors in animal models, one can gain an understanding of the basic biology underlying them and apply this knowledge to figure out how diseases occur and find the corrective measures.

9.1. Non-Human Primate Models

Non-Human Primates has been successfully modeled to study PD. The clinical and behavioral assessments like abnormal in voluntary movement scale for dyskinesia can be practically applied to the marmoset monkeys free from any adaption (Van Vliet et al, 2006). MPTP induced marmoset monkeys show clear and lasting behavioral features of PD and many quantitative motor behavior tests such as full body motor behavior such as locomotor activity, rotational task; specific motor performance such as 'arm reaching' motor behavior is very well described in such model (Marshall and Ridley, 1996). These studies are extensively used in PD diagnosis and moreover treatment in non-human primate model can directly be used for PD patients at the clinical level (Philippens, 2008). However, high cost of production and maintenance limit the wider use of non-human primate model; there is very less genetic data to work with considering the fact that there is a low efficiency of assisted reproductive technology for producing genetically engineered oocytes and embryos necessary for genetics base study for PD (Oliveira et al, 2017). Also, there are several technical issues for deriving and cloning embryonic stem cells and long period of gestation of embryo and fewer off spring per year makes it very extended process to make any result conclusive.

9.2. Rodent Models

Mice and rats represent one of the most commonly used animal models in PD. Most of these models have been developed with use of neurotoxins and also genetic models through genomic manipulations. α -synuclein aggregation have been implicated to be main reason for development of PD and mouse prion promoter (mPrP) A53T α synuclein transgenic mice display complete range of α -synuclein pathology that is seen in humans including α -synuclein aggregation, fibrils and truncation, α -synuclein phosphorylation and ubiquitination and gradual age-dependent neurodegeneration (Chesselet, 2008; Dawson *et al*, 2003). Several approach to create α -synuclein transgenic mice with progressive loss of DA neurons such as crossing α -synuclein transgenics to *parkin* knockouts (von Coelln *et al*, 2006), conditional expression of α synuclein (Lin et al, 2009), and DJ-1 knockouts (Ramsey et al, 2010) and overexpressing α -synuclein with DA specific promoters have not been very productive (Daher et al, 2009; Thiruchelvam et al, 2004). Rodent's resistance to neurotoxin like MPTP varies from species to species and even strain to strain (Hamre et al, 1999). Even though there is a complete penetrance of α -synuclein in presence of mutant PARKI, PINK1 or DJ1, there is no meaningful DA neuron degeneration in mouse model thus leaving a wide gap on the accuracy of this model. Besides, the open field test and swim tests which is employed to study DA deficiency induced motor deficit in mouse models require the animal to learn first. Not all the animals learn these skills even before receiving DA lesions so when failed it is difficult to deduce if it is learning deficit or PD motor symptom, even the specimen which show symptoms are typical of bradykinesia and akinesia (Potashkin et al, 2010).

9.3. Zebra Fish Model

The Zebra fish model is efficient because of its higher offspring production rate and possibility of screening toxin induced or genetically engineered traits very rapidly. The transparent embryo develops outside the mother so the neural developmental pattern under induced PD condition is easily traceable. Its high similarity in catecholaminergic system with human makes them an excellent model for PD (Pienaar *et al*, 2010). Moreover, the TH containing neurons (the DA neurons) in the ventral diencephalon of zebrafish brains seems to be homologous to mammalian midbrain SN and ventral tegmental neurons (Son *et al*, 2003). PD-related genes, such as *DJ-1* (Bai

et al, 2006; Bretaud *et al*, 2006), *UCH-L1* (Son *et al*, 2003), *SNCA*, *PINK1*, *PARK2* and *LRRK2* (Flinn *et al*, 2008) are evolutionarily preserved between humans and zebrafish, and their protein products are expressed in zebrafish ventral diencephalic DA neurons and the PD symptom in this region affects the spontaneous swimming phenotype hence the genotype to phenotype is highly relatable.

The similarities are limited to lower motor neurons of the fish while, upper motor neurons are partially resembling with the human as there is absence of the corticospinal and rubrospinal tracts (Babin *et al*, 2014). The progression in knockout lines used as most common way of silencing a gene through Antisense Morpholino Oligonucleotides (AMO) works only during few days of development, therefore generating a late disease specimen like PD is difficult (Babin *et al*, 2014). Moreover, AMOs tend to go off target and induce apoptosis in motor neurons (Bill *et al*, 2009; Eisen and Smith, 2008), thus assessing the phenotype with regard to gene activity becomes challenging. Further, loss or gain of function of a gene produces delayed development and body malformation, so when there is a motor neuron deficiency or malfunction it might be secondary effect rather being primary effect of the gene in question (Babin *et al*, 2014).

9.4. Drosophila Model

In the Order Insecta, the family Drosophilidiae comprises of more than 3,500 described species including the genus *Drosophila* (Bachli, 1998). It is estimated that there are more than 2240 biologically valid species of *Drosophila* (Wheeler, 1986). In India, reports from the review of literature show more than 200 *Drosophila* species (Hegde *et al*, 2000) some of which are endemic to certain regions of the country and a few are cosmopolitan in distribution.

Drosophila has been an essential model organism to study human diseases. It was initially used in the laboratory by William Castle at Harvard University as a minute, fast generating organism for embryological studies. It was not until 1909 that the fruit fly made its initial application in the field of genetics. A natural transformation in eye color from brick red wild type to white caught the attention of Columbia University professor Thomas Hunt Morgan. He investigated if this mutation would go after the hereditary patterns as predicted earlier by Gregor Mendel. After mating the white-eyed male (w) with a wild type female (w+), he found that the resulting ratios of progenies followed the predicted patterns. This demonstration brought to light the importance of *Drosophila* to the research community. It has been reported that about 75% of identified human disease genes have orthologs in fly genome (Llyod *et al*, 2010; Reiter *et al*, 2001). Many investigations employ environmental toxins in *Drosophila* models and try to understand the disease pathology for exploring possible therapeutic intervention.

9.4.1. Environmental Toxin Models of PD using Drosophila

A number of environmental neurotoxins including herbicides and pesticides are used in *Drosophila* to mimic PD-like symptoms and understand the disease mechanism (Bonini and Fortini, 2003). The fly has well developed CNS and complex functions like walking, climbing, and flying makes *Drosophila* more appropriate model for understanding PD. Such intricate actions are alike among different strains which makes easier to characterize a toxin induced PD model (Feany and Bender, 2000). Widely employed environment neurotoxin PD models are discussed below.

9.4.2. Rotenone Model of PD in Drosophila

Rotenone induced slowing down of mitochondrial respiratory complex has been extensively employed for studying the function of mitochondria in apoptosis (Chauvin *et al*, 2001; Barrientos and Moraes, 1999). Mitochondria are not only involved in ATP generation in eukaryotes but also play significant role in apoptosis (Wang, 2001; Kroemer and Reed, 2000; Green and Reed, 1998). Rotenone is lipophilic and therefore can simply cross the blood-brain barrier (BBB) (Coulom and Birman, 2004). In one such model, rotenone exposure led to neuromotor defect and significant inhibition of neurotransmitter enzyme AchE activity. They reported induction of OS among rotenone exposed flies as evident by sharp increase in ROS and malondialdehyde (MDA) levels. Another study subjected fly to rotenone treatment and reported decreased in DA content and impairment in motor activity, increased mRNA expression of antioxidant enzymes (Rao *et al*, 2016; Sudati *et al*, 2013).

9.4.3. PQ Model of PD in Drosophila

PQ is commonly employed in the *Drosophila* model to induce phenotype associated with PD system and other animal models to study PD. It has been reported that D1-like DA receptor (DAMB), strongly add to the rapid CNS breakdown as a result of PQ exposure in the fly (Casser, 2015). They have shown that DAMB expression is reduced upon continuous neuronal DA synthesis and confers resistance to the PQ toxicity while age-related diminution of resistance to PQ was assisted by a considerable enhancement in DAMB receptor. The findings suggest that several other aspects remain to study and understand DA pathogenesis in *Drosophila*. The fly display motor impairment including tremor at rest, slowness, restless and unsteadiness which is similar to PD symptoms. When allowed to climb over the vials in testing motor functions, the flies freeze and frequently lose the grip and fall to the bottom (Chaudhuri *et al*, 2007). Degeneration of DAergic neuron upon PQ exposure is time

dependent where it has been shown that PPL1 (protocerebral posterior lateral) cluster is affected by 6 hours, 12 hours for posterior inferiormedial protocerebrum (PPM) 2 and 3 cluster but 20-24 hours for PPM1 and PPL2 clusters. This process is accompanied by alteration in neuronal appearance by aggregation of cell bodies to circular shape, fragmentation and then loss (Chaudhuri *et al*, 2007). Recent study employing PQ toxin model of PD in fly have also reported increased mortality, locomotor impairment, decreased DA levels (Soares *et al*, 2017) suggesting the toxicity associated with PQ exposure and relevance of the model to induce Parkinsonism.

10. Conclusion

Many workers have made extensive effort to understand the pathophysiology associated with PD and there are numerous data that are available on fly PD models. They treat the organism with environmental toxins and study the modulatory effect of prospective therapeutic compounds. However, till date their study is limited to only in young animals model. PD is a late onset disease; it is important to understand the disease progression and therapeutic effectiveness at later stage of the organism, the age equivalent to disease onset in human. Therefore, I decided to develop stage specific *Drosophila* model of sporadic PD and screen the protective efficacy of K at different stages of *Drosophila* adult life span.

CHAPTER II

DEVELOPING ADULT STAGE SPECIFIC *DROSOPHILA* MODEL OF SPORADIC PARKINSON'S DISEASE

1. Introduction

A suitable model for PD must recreate the condition showing progressive loss of DA neurons, accompanied by degeneration of other neurons and considerable diminution in DA level. In addition, since the onset of the PD is late onset, the model organism should also show the phenotype during adulthood. The model animal must also imitate the PD pathology and neuronal dysfunction by showing phenotype such as bradykinesia, rigidity, postural instability and resting tremor, with motor symptoms having positively reactive to L-DOPA (L-dopa, the precursor of DA) or any anti PD drug therapy.

2. Drosophila as a Model of PD

2.1. Advantages

The advantage of using *Drosophila* to study human NDD and their essential molecular and cellular pathology are numerous. One of the main factors is its short life cycle which takes about 10-14 days to develop from egg to fully matured adult fly; single female can produce many offspring and there are several established methods to maneuver its genetic expression (Bilen and Bonini, 2005). Besides, as compared to other models, *Drosophila* is commonly employed to study disease for decades; making its anatomy and phenotypes relatively more comprehensive to experimental biologists (Venken, 2007; Matthews *et al*, 2005). With a well developed CNS and prominent number of DA neurons (White *et al*, 2010, Botella *et al*, 2009), the fly has properly distinguished behaviors that are conserved with different strains in 90% of the cases (Lessing and Bonini, 2009) which makes the *Drosophila* an efficient and less expensive model.

Genetically, about 75% of human disease related genes have functional orthologs in the fly (Lloyd and Taylor, 2010, Reiter *et al*, 2001). The whole resemblance at nucleotide or protein level is 40% while in terms of the conserved domain it is 80%-90% or even higher (Pandey and Nichols, 2011).

Although the invertebrate fly have major differences in physiology when compared to human, yet through genetic manipulations it is possible to induce and mimic human PD pathology in the fly. Over the years, researchers using fly model have developed notable range of genetic tools. The binary system expression like UAS/GAL4 system that allows inserting a specific gene of interest in selected set of cells, activate and deactivate them has made the fly more convenient and cost effective model for research. Using such approach researchers can examine role of gene in the region of interest while keeping the other regions intact and also maintaining the fly health, which is significant for replicating human disease conditions targeting particular cell-types. Such approach using transgenic fly can be achieved with precision including observation of age dependent DA neuronal degeneration and convenient visible of LB and Lewy neuritis (LN) which make up the clinical hallmark of PD (Feany and Bender, 2000). Degeneration of DAergic neurons with aging and motor dysfunction remains the two main phenotypes in Drosophila PD model. There are discrete clusters of DA neurons distributed all over the fly brain that are functional for its diverse behaviors. Thus Drosophila shows very complex motor behavior like climbing, flying, locomotion, conditioning to fear etc. Hence age dependent loss of DA neurons in PD conditions (Feany and Bender 2000) and sporadically induced PD conditions showing DA neuron dysfunction and subsequent phenotypes are easily characterizable and can be related to humans. Thus apart from its use in advancing the understanding of basic biology, Drosophila has been effectively employed to investigate a number of human diseases, including PD.

2.2. Disadvantages

Drosophila being an invertebrate animal shares some important dissimilarity in physiology as compared to human such as the brain anatomy, cardio vascular and respiratory system; Less complex and adaptive immune system as in vertebrates, effects

of some drugs on the organism might differ strongly (e.g. conversion of pro-toxins to toxins in liver) (Prubing *et al*, 2013). Discrepancy related to blood-brain permeability has been a matter of concern when it comes to studing disease of CNS (Stork *et al*, 2008). The model cannot accurately predict the effects of drugs on human and the metabolic distinction is another matter to consider in study of drug efficiency and toxin induced disease phenotype.

3. Drosophila Mimicking Human PD Symptoms

Although the physiological dissimilarity of fly when compared to human are very significant, the central phenotype associated with PD pathology experienced in human are reproduced in a commendable fashion through the method of disease induction using toxins or by employing transgenic modification. The accuracy is so close that brain area specific and age dependent DA neuronal degeneration as commonly observed in PD patients through post mortem studies and hallmark PD biomarker i.e., the LB and LN inclusions are visible in transgenic model system (Feany and Bender, 2000). Behaviors like mating, aggression, conditioning to fear; learning and motor qualities like flying, climbing and walking which are attributed to human are also seen in *Drosophila* (Lessing and Bonini, 2009; Sokolowski, 2001). These multitude triats are affected by the PD onset and progression in human that are reproduced and characterized in the fly model.

In *Drosophila*, the DA neurons are spread as a group of clusters all over the brain. Specific clusters derive specific functions for various behavioral patterns of the fly. The two areas generally targetted are (i) the mushroom bodies: Associated to memory formation and motivation, (ii) the Central Complex: Centre for motor behavior control. Region specific controlling of diverse behavioral patterns in fly is similar to those seen among the mammalian brain (White *et al*, 2010). Schematic representation of an adult fly brain with the distribution of DA neuron clusters are given in Figure 1.

4. Drosophila Genetic Similarities with Human

Genetically, *Drosophila* shares 61% homology with human. All the genes till date reported in human associated with Parkinsonism, be it familial or sporadic are available in *Drosophila* as a homolog except for the α -synuclein gene which produces LB and LN at the extracellular brain matrix which is the distinctive hallmark biomarker under PD condition in mammalian brain. This adds up to the advantage for genetic manipulation employing *Drosophila* as a model for PD.

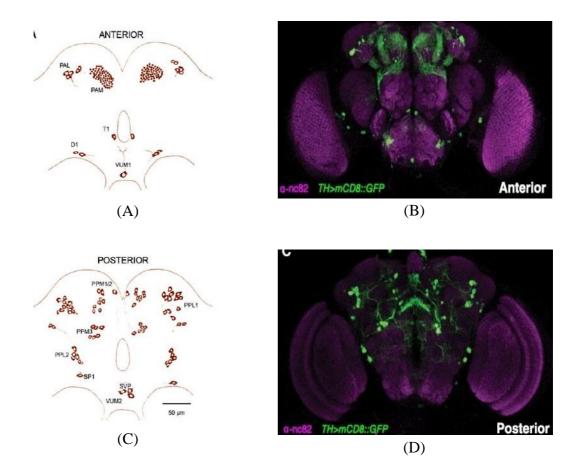


Figure.1(A-D) : A and C Schematic representation of an adult fly brain with the distribution of DA neurons grouped in clusters and arranged with bilateral symmetry (modified after Nassel and Elekes, 1992) [Image adapted from Botella *et al*, 2009]. B and D Confocal Z-stack of TH > mCD8::GFP brain; anti-nc82 immuno-reactivity together with GFP labeling reveals DAergic neurons in the anterior and posterior brain (White *et al*, 2010).

List of Parkinsonian genes and their homologues in flies is given in Table 1.

Table 1: Showing Parkinsonian genes and their fly homologues (adapted from				
Ayajuddin <i>et al</i> , 2018)				
Gene/Protein	Inheritance	Fly Homolog	Protein Function	
A-synuclein	AD	None	Pre-synaptic protein	
Parkin	AR	Parkin/CG10523	E3 ubiquitin ligase	
UCH-L1	unclear	Uch/CG4265	E3 ubiquitin hydrolase/ligase	
PINK1	AR	Pink1/CG4523	Mitochondrial kinase	
DJ-1	AR	DJ-1a/CG6646	Redox sensor/Chaperone	
		DJ-1b/CG1349		
LRRK2	AD	lrrk2/CG5483	Kinase/GTPase	
HtrA2	AD	HtrA2/CG8486	Mitochondrial pro-apoptotic protease	
GBA	unclear	CG33090	Lysosomal enzyme	
POLG	unclear	tamas/CG8987	Mitochondrial DNA polymerase	
Tau	unclear	tau/CG31057	Microtubule stabilization	

<u>Legends</u>: UCH-L1 = ubiquitin carboxyl-terminal esterase L1; PINK1 = PTEN induced putative kinase 1; LRRK2 = leucine-rich repeat kinase 2; HtrA2 = high temperature requirement protein A2; glucocerebrosidase = GBA; POLG = polymerase gamma; AD = autosomal dominant; AR = autosomal recessive.

5. Scope for Stage Specific studies in Drosophila Model of PD

Several laboratory studies have reportedly used Drosophila to develop model of PD using young flies of 5-10 days old (Jahromi et al, 2015; Cassar et al, 2015). Studies regarding susceptibility genes and pathways on PD under PQ treatment have been conducted mostly using young flies (Wang et al 2014; 2003). However, it is important to note that NDD such as PD is late onset disease. The adult life span of fly has been reported to comprise of three phases namely health, transition and senescent span (Arking *et al*, 2002). These stages are governed by varied assortment of gene expression and such pattern is comparable to corresponding life stages of human. There is a marked variation of about 23% in genome-wide transcription profiles as the Drosophila cross through the adult stages and age. Genes including those associated with stress response and oogenesis showed age dependent transcript representation and this aging process is characterized by changes in expression of many genes depending upon life stage specific factor (Pletcher *et al*, 2002). Since the animal models are indispensable for disease progression studies and therapeutic approach, it is equally important to follow the age specific study while developing animal model for NDD such as PD because the onset of this disease in human is clinically manifested during the later stage of life. Therefore, research comprising the age specific stages will greatly enable the early identification of markers, providing timely interventions in the initial course of disease progression.

Hence, I developed a *Drosophila* model of PD by considering all the different time points in the fly life span i.e., health span, transition span and senescence span.

All these life stages of *Drosophila* are believed to have both common and unique complex process and can be influenced independently by a relatively large number of stages associated pathways. Therefore, adult specific model of PD is important because a particular target may be active at one stages of life while inactive in another stage and therefore genetic targets of compounds such as curcumin (K) may be present only at one stage but not in all stages of life. As such the compound will be effective in the organism only during those phases when its target molecules are present (Soh *et al*, 2013) which is an important and interesting paradigm.

Until now, there are no reports relating to the effectiveness of K neuroprotection using PD models at later stage of adult life. It is an indispensable aspect while developing animal model for studying the disorder since it is during the later phase where late-onset NDD like the idiopathic PD sets in.

Drosophila has been an excellent organism to study different stages of life span and specific gene expression. Similar to the different human life stages like childhood, adulthood and old age. Various phases of life stages have a distinctive pattern of gene expression (Pletcher *et al*, 2002). There exists considerable variation of about 23% in genome-wide transcript profile according to different age group in *Drosophila*, demonstrating that genetic targets of prospective polyphenol compounds may be absent during some phase in the entire stages of life. Therefore, this aspect has to be considered while developing any disease model for therapeutic approach. Also implicating that polyphenol compound may have distinctive beneficial effect during certain stage of life and later turn out to have neutral or harmful action in another stage in the adult life period of *Drosophila* (Phom *et al*, 2014).

Major changes in gene expression with age occur in the genes related to energy metabolism such as proton transport, energy pathways, oxidative phosphorylation and neuronal function, especially responses to light. Genes involved in protein catabolism and several other metabolic processes also showed age dependent changes. Interestingly, biological processes namely, the caloric restriction and the light reduction are two known environmental changes that increase life span in *Drosophila*. Decline in reproductive capacity is an age-related phenotype, and the reproductive system seems to play an important role in longevity (Finch *et al*, 1990). There are decreased RNA levels for several genes that are involved in reproduction including two genes that encode for members of the Acyl carrier protein (Acp) family. The Acp from male flies stimulates female egg-laying and facilitates storage of sperm in the female genital tract. The transition span and the senescence span of *Drosophila melanogaster* are accompanied by the reduction of transcript levels for genes involve in reproduction, metabolism, and protein turnover stemming from age regulated transcript level changed in response to OS and accumulation of free radicals.

6. Oxidative Stress in PD

OS is a condition of cellular discrepancy in the levels of reactive oxygen species (ROS) generated and the capacity of a biological system to detoxify the reactive intermediates, which creates an imbalance and lead to harmful state that contributes to cellular injury. OS is classically defined as a redox imbalance with an excess formation of oxidants or a defect in antioxidants (Sies, 2015). In physiological conditions, tissues have a functional anti oxidative system glutathione (GSH) that is depleted due to OS. Excessive generation of free radicals causes alteration in GSH level thus decrease in GSH concentration contributes in OS and consequent brain damage (Shungua *et al*, 2017; Jain *et al*, 1991). Alterations in the physiologic maintenance of neuronal redox balance interfere with several biological processes that eventually lead to cell death. Several machineries are suggested to be responsible for ROS influx that include the normal metabolism of DA itself, injury to mitochondria, iron, neuroinflammatory cells, calcium, and aging (Dias, 2013).

The body in general has developed several defense mechanisms to counteract OS, however, the brain which comprises only 2% of the total body weight, appears to be more susceptible to oxidative damage than any other organ as it consumes about 20% of the resting total body oxygen processing a lot of oxygen per unit tissue mass. Human brain has relatively low levels of antioxidants, high polyunsaturated fatty acids and large amount of iron in regions like the globus pallidus and the SN, thus susceptible to OS. Humans need oxygen to survive, but hyperoxia produces toxicity, including neurotoxicity (Chavko *et al*, 2003). Moreover, being post mitotic, when the neurons in the brain are injured, their functional ability becomes permanently impaired (Calabrese *et al*, 2005).

OS is suggested to be involved in the degeneration of DAergic neurons in PD. Abnormal acculmulation of reactive oxygen and nitrogen species in PD lead to injury in important cellular components such as lipids, proteins, and DNA. Elevated levels of MDA which is an indicator for lipid peroxidation and 4-hydroxynonenal; high protein oxidation as indicated by protein cross-linking, carbonylation and breakdown; high levels of 8-hydroxy-2'-deoxyguanosine, a product of DNA oxidation, causes oxidative injury to brain. Due to all these modifications which are experimentally demonstrated, the "free radical hypothesis" has become a major factor for understanding the etiology and therapeutic approach to PD (Jenner, 2003).

7. Environmental Exposure to PQ

PD progressions have been associated with exposure to environmental stressors like PQ an herbicide widely used in agricultural farming. Ingestion of PQ by accident or for suicidal purposes causes severe pulmonary lesions, which are often fatal. PQ has two positive charges and it undergoes single electron reduction chemistry in those cells that can sequester it, which result in a widespread oxidative and nitrosative stress. The underlying mechanisms are directly or indirectly related to reactive oxygen species. PQ can induce PD-like lesions in certain mouse strains and rats and number of evidences suggests that the exposure of PQ results in neurotoxicity. Therefore, PQ is generally used in animal models including the fruit fly *Drosophila* to study and understand PD and the degeneration of DAergic neurons which characterizes the disease.

Since PQ is used extensively in agriculture, there has been a concern that gradual exposure to herbicide may increase the risk of developing PD in the human population. Considerable attention was gained when cases of human exposure resulting in extreme toxicity were reported and its use in eradicating marijuana crops. Additionally, the structure of PQ is closely comparable to that of MPP⁺, which led to assumption that PQ also might be a potential DA neurotoxin as well (Dawson *et al*, 2003). PQ is accumulated into the brain through the BBB, possibly via a carrier mediated mechanism (Shimizu *et al*, 2001).

8. Epidemiological studies on link between PD and PQ

A number of epidemiological studies have reported a connection between PQ exposure and the development of late onset of PD suggesting the possible role of environmental factors in developing sporadic PD, which make up about 90% of the PD cases. A study conducted in Taiwan between 1993 and 1995 on 120 PD patients and 240 hospital controls was reported (Liou *et al*, 1997), generating more attention on possible association between PQ and PD. In their analysis, they showed that the account of living in rural surroundings, regular exposure to herbicides/pesticides and farming and use of PQ were linked with an increased possibility to PD risk. Reports explaining the interaction between PQ and PD have been published (Hertzman *et al*, 1990). A study conducted on disease risk linked with occupational exposure to environmental factors like pesticides support a significant association of PD with toxin exposure (Tanner *et al*, 2011).

Laboratory studies have provided contradictory outcome. Early studies suggested that the pharmacokinetic properties of PQ made it an improbable candidate since it has low partition coefficient, restricted absorption and poor accessibility to CNS (Koller, 1986). Besides that, PQ also showed modest penetrative ability in the brain structures of rats with an intact BBB (Widdowson *et al*, 1996). However, some researchers have reported the capacity of PQ to pass through the BBB, probably via the neutral amino acid transporter (Shimizu *et al*, 2001), and to build up in certain brain regions of the mouse (Prasad *et al*, 2014).

9. PQ induced Parkinsonism in Drosophila Model

Exposure to environmental toxins like the herbicide PQ over a period of time has been linked to the development of PD. PQ is a nonselective pesticide that was first produced in 1961; a member of a chemical class known as bipyridyl derivatives, which includes diquat (1,10-ethylene-2,20-bipyridylium dibromide) and cyperquat that has the same structure as the MPTP metabolite MPP⁺. The main cause of death associated with PQ consumption and poisoning is due to pulmonary accumulation of bipyridyl compound causing respiratory failure due to oxidative induced damage to the alveolar epithelium with subsequent pulmonary fibrosis. PQ exposure also triggers OS by disturbing the redox homeostasis in the body and generates abnormal amount of ROS which can neutralize cellular defense mechanisms (Fig 2).

Laboratory exposure of model animals to PQ leads to reduction in action of neurotransmitters in brain (Mehdi and Qamar, 2013). Also, several other studies have reported neuronal damage and death induced by PQ; cerebellar granule neurons in rats

(Stelmashook *et al*, 2007), spinal motor neurons (Kriscenski-Perry *et al*, 2002). Using rat brain neuroblast cell model it is shown that PQ also triggers rapid activation of intracellular signaling cascades leading to neural cell death (Niso-Santano *et al*, 2006).

To carry out initial discovery and characterization of neurodegenerative genes and the molecular pathways involved, Drosophila model offers an optimal research tool. More importantly, the fly model is extensively used to study gene-environment interactions with influence on phenotypic outcomes (Okray and Hassan, 2013). Study of survivability of Drosophila exposed to PQ reveals that the fly is vulnerable to PQ in a concentration dependent way. Varying degrees of toxicity has been shown upon PQ exposure. Exposing PQ systemically cause about 30% selective DA neuronal loss in the SNpc and augmented expression and aggregation of α -synuclein (Manning-Bog *et al*, 2002). PQ also is reported to cause loss of DAergic neurons from SN and disintegration of striatal terminals accompanied by a reduced locomotor action (Ossowska et al, 2005; Brooks et al, 1999). These studies support the understanding that loss of DA neurons upon PQ exposure can be experimentally reproduced. A recent study in Drosophila has suggested that DA receptor contributes to PQ induced neurotoxicity. Targeted inactivation of D1-like DA receptor, DAMB in glutamatergic neurons (GNs) noticeably improved the survival of Drosophila exposed to either PQ or neurotoxic levels of DA, and DAMB overexpression in these cells made the flies more susceptible to both compounds (Cassar et al, 2015).

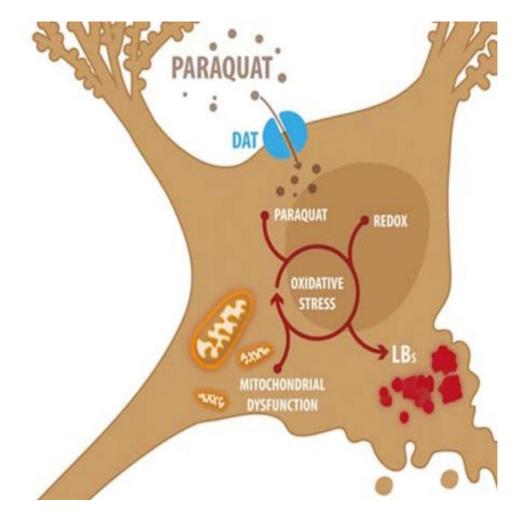


Figure.2. Molecular events related to PQ-induced damage. PQ crosses the BBB, entering DAergic neurons via the DA transporter (DAT). In neurons, PQ triggers a vicious cycle related to OS that involves an imbalance in the redox status, resulting in mitochondrial dysfunction, protein aggregation (the formation of LBs) and further production of pro-oxidant mediators (Adapted from Bastías-Candia *et al*, 2018).

- **10. Materials and Methods**
- 10.1. Fly Stock

Drosophila melanogaster Oregon K were obtained from National *Drosophila* Stock Centre of University of Mysore, Karnataka, India. The flies were raised at 22°C-24°C in a *Drosophila* environmental chamber (Percival, DR41VL). The flies were fed with regular culture media prepared with sucrose, yeast, agar agar, and propionic acid. Male flies of were used in the present study.

10.2. Chemicals

Methyl viologen dichloride hydrate or Paraquat was purchased from Sigma Aldrich (St.Louis, MO). Sucrose was procured from Sisco Research Laboratory (India). Whatman filter paper no.1 disc was used as a feeding medium in the experiment in 30x100mm glass vial.

10.3. Treatment Protocol

Life span curves were obtained from independent trials with a minimum of 100 flies per experiment and the experimental flies were shifted to freshly prepared culture media every third day until all the flies were recorded dead. In filter disc treatment method volume of 250µL of 5% sucrose was feed to the control flies and 250µL of different concentrations of PQ solution prepared in 5% sucrose were pipette on a filter disc placed in 100x30mm glass vial. Twenty-five flies were transferred on each vial for the treatment and total of 100 flies were used for each concentration in the experiment. At every 24 hours (hrs) duration fresh media solution was prepared and the flies were transferred on it. Mortality was recorded till all flies were dead.

10.4. Negative Geotaxis Assay

Taking advantage of the fly natural tendency to climb towards the light the negative geotaxis assay also called as climbing assay was performed. The assay was employed using a plastic climbing tube where each experimental fly was dropped in the tube using an aspirator and initially left it undisturbed to become accustomed in the new environment for 2 minutes. Each tube was fixed on a sponge and the experimental flies were gently tapped to the base of plastic climbing tube. The distance (measured in centimeter) each fly could climb up in 12 seconds was noted. Each fly was subjected to the experiment three times and data were collected from at least 10 flies in all the experimental set (Botella *et al*, 2004).

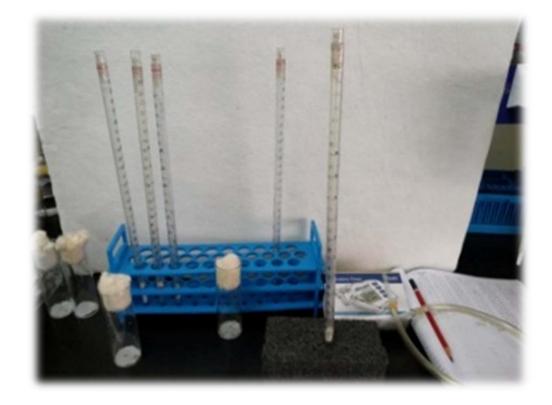


Figure.3. Set up for Negative geotaxis assay. Individual flies were made to climb up the cylindrical tube. The distance travelled by the fly in 12 seconds was recorded.

11. Results

11.1. Characterization of adult Health, Transition and Senescence Life Stage in *Drosophila*

The adult life period of *Drosophila* has three stages namely; health span, transition span and senescent span (Arking et al, 2002). The health phase is indicated by a period of life span where no natural deaths occur. The adult transition period is recognized when there is visible but less significant decrease in the survival of adults and described by instance accompanied by about 10% mortality and 90% survival. The adult senescent phase is defined by the slow and stable decrease in the number of live flies as evident by decline in the survival curve due to gradual increase in mortality rate. It is illustrated with gap in between last part of the transition stage and the greatest extent of fly prolonged existence. In animal studies the upper limit of longevity is normally considered as the mean life stages of the longest lived 10% of a particular group. Flies were cultured on the standard culture medium by transferring to freshly prepared culture media on 3rd days. Number of death in each groups were recorded till every flies were dead. Basing on the experiments of longevity assessment using Oregon K strain of Drosophila, it was found that the health span extended up to 30 days; the transition stage was recorded from 31-60 days of adult period and the senescent period is from 61-120 days. The greatest extend of the fly longevity was recorded as 121 days, while the median survival duration was 95 days as shown (Fig.4) (Phom *et al*, 2014).

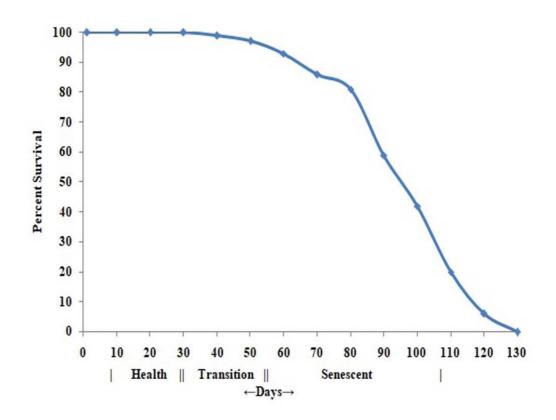
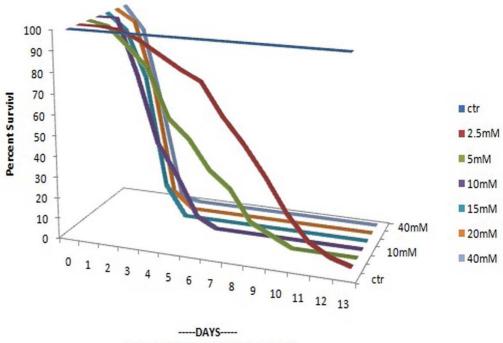


Figure.4. Survival curve of Oregon K male flies in standard culture media. The flies were transferred to fresh media on every 3rd day. The numbers of mortality were recorded until all of the flies died. Health span extends from day 4/5 to 30 days; the transition phase is 31–60 days of adult span, and the senescent span is 61–120 days. Maximum life span is 121 days and median life span is 95 days.

11.2. Drosophila is susceptible to PQ in a concentration dependent manner

For exploring the effect of OS in Drosophila, the flies on eclosion were collected and fed on normal culture medium till they were age 4-5 days old. Then they were transferred to a glass vials containing several range of PQ concentrations (2.5, 5, 10, 15, 20, and 40 mM). Total of 100 flies with 25 flies per vial were used for each concentration. In a glass vial single filter paper disc was soaked with 250µL of 5% sucrose solution as control and with various concentrations of PQ prepared in 5% sucrose solution and experimental flies were transferred on it. The survival rate of the flies were noted at each 24 hour time point till all the experimental flies were recorded dead in 2.5mM PQ solution which is the most diluted concentration used (Fig.5). After 24 hrs of exposure the mortality in the highest concentration was about 15%, while in 20mM and 15mM, it was about 8% and 5% respectively. In the concentrations of 10mM and below, there was no mortality recorded at 24 hrs of exposure. At 72 hrs of exposure, the survival rates were 95%, 79%, 48%, 15%, 8% and 5% from lowest to highest PQ concentrations respectively. The importance to assess the toxicity is evident from the experimental results in the highest concentration i.e., 40mM, where the toxicity of PQ was so high that, 98% of the flies were dead by 72 hrs of exposure; whereas in the lowest concentration i.e., 2.5mM all the flies were dead by 13th days of exposure. Therefore, it is necessary to circumvent the toxin concentration while developing a model so as to avoid wrong interpretation of the results from false phenotype that may be visible only due to the high concentration of the toxin and possible organismal failure, but not the actual phenotype required from the model organism.



4-5 days old flies susceptibility to PQ

Figure.5. The mortality curves of *Drosophila melanogaster* in various concentrations of PQ. Different concentrations (2.5, 5, 10, 15, 20, 40mM) of PQ were fed to adult 4-5 days old flies. The flies were susceptible to PQ dose and time dependent manner. Comparison of survival curves reveals that the response difference among different tested concentrations was significant (CTR=Control) (log-rank [Mantel–Cox test, p < 0.0001]).

11.3. Negative Geotaxis Assay to characterize mobility defects

PQ induced motor dysfunction was assessed by exposing the flies to 10mM PQ and subjecting them to Negative geotaxis assay. After 24 hrs of feeding, the flies were allowed to climb up on the tube to perform climbing assay also called the negative geotaxis assay. The climbing speed of PQ treated flies were decreased by 30% in 4-5 days and 56% among 55 days old flies (Fig.6a,b) when compared to their respective controls. PQ treated flies mimic the symptoms of resting tremor and bradykinesia distinguishing clinical symptoms associated to human PD patients. Some of the flies showed agitation and restlessness as seen from their continuous flipping of wings and some tried to climb up but lose their grip and fall back to the bottom of the climbing tube. This shows a typical motor dysfunction which is caused due to the effect of PQ treatment on DAergic neurons. Thus using the *Drosophila* as the model organism, PQ treatment could replicate the conditions related to motor symptoms that are seen among the human PD patients.

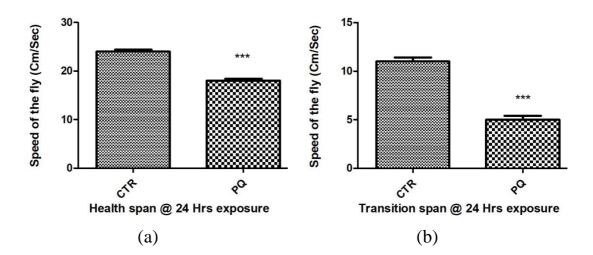


Figure.6a,b. Negative geotaxis assay performed on (a) Health stage (4-5 days) and (b) Transition stage (55 days) old male fly. Compared to the control flies, the PQ treated flies showed reduced mobility in both the age groups. As compared to controls there was 30% and 56% reduction in climbing ability of PQ treated flies in Health and Transition stages respectively. Suggesting the toxicity of PQ on DAergic neurons affects the motor ability of the flies (***) p<0.0001.

12. Discussion

12.1. Developing Fly Model of PD

Epidiomological studies have shown a link between exposure to environmental toxins like PQ and development of PD during later phase of life, which could be an additive factor of sporadic PD that comprise of more than 90% of the PD cases. *Drosophila* shows susceptibility to PQ exposure in a dose dependent manner.

12.1.1. Health Phase Specific Fly Model of PD

The health stage begins when the developmental period ends. It is defined by a low age specific mortality rate and a high survival rate resulting in a significant increase in both mean and maximum life span of Drosophila. The adult cohort starts with 100% survival and this high level is maintained for some time in most health cohorts. The HS ends when the survival rate declines to below 90%, or when there is a negative sign in the survival curve whichever comes first (Arking et al, 2005; 2002). Survival and longevity studies of Drosophila Oregon K strain was done on standard culture medium and on its basis, the survival of flies up to 30 days (adult fly eclosion from pupa to 30 days of culturing) is considered as the phase termed as health life stage of fly. Effect of OS on the health stage of Drosophila was studied using varied concentration of PQ. Upon 24 hrs exposure, no mortality was observed at 10mM and below whereas significant death was seen for flies exposed to the highest concentration of PQ. One of the hallmarks of PD is the onset of motor symptoms. Exposure of health stage flies 10mM PQ showed a 30% decline in climbing speed as seen by performing the negative geotaxis assay. The exposed flies also showed PD like symptoms of resting tremor and bradykinesia. This motor symptom is atypical of effect of PQ on DAergic neurons as loss of DAergic neurons in the SN along with associated reduction in motor ability upon PQ administration have been reported (Bastias-Candia et al, 2018).

12.1.2. Transition Phase Specific Fly Model of PD

The transition stage is defined as a period of the Life Span that encompassed between the adult survival rates of 89% and 81%. The transition stage shows an increased early survival that leads to a significant increase in mean, but not in maximum life span. The transition stage is variable. Some strains of *Drosophila* display an almost continuous decrease in the survival curve, suggesting that these animals are somewhat weaken such that they have a relative high age specific mortality rate from eclosion onwards (Arking *et al*, 2005; 2002). The transition life stage of *Drosophila* is considered from 31 days to 60 days as seen from the experiments done on survival and longevity. Exposure to PQ induces OS and increases the mortality of flies is a dose-dependent manner. Transition flies when exposed to 10mM PQ showed 36% decrease in climbing ability as performed using the negative geotaxis assay. Some of the flies showed agitation and restlessness and loss of grip which are typical motor dysfunction symptoms.

13. Conclusion

To develop an animal model including *Drosophila* model for disease such as PD and study of its pathophysiology, the need to figure out the adult life stages and prospective model toxin concentration is very essential. Because if the extent of its toxicity is not established, the particular model organism may die of organismal breakdown due to higher concentration of the toxin even before the DAergic neurons gets degenerated and the phenotype that the researchers see may not be relevent to the condition they are trying to understand. Therefore, after establishing the toxicity of PQ, I decided to expose the flies with 10mM PQ and differentiate the behavioral, biochemical, cytological, and molecular markers that are associated with PD at 24 hrs time point where there was no mortality.

CHAPTER III

ASSESSING THE TOXICITY OF CURCUMIN IN DROSOPHILA MODEL

1. Introduction

Several researchers have suggested the neuroprotective efficacy of natural products by demonstrating their modulating ability of oxidative stress markers and phenotype associated rescue upon exposure to neurotoxic agents in different disease models. However the pathway of activity of these compounds, in what concentration and biologically active form remains obscure. Sirtuins, an evolutionary conserved family of proteins has a modulatory function on age associated process among higher organisms. Their varied natural role in organisms overlay the basis in which more detailed studies were carried out for their therapeutic prospect in presently not treatable neurodegenerative disorders. Sirtuins inhibitors are found to show protective action on a-synuclein mediated toxicity (Outeiro et al, 2007). Resveratrol, (3,5,40-trihydroxystilbene) a natural polyphenol phytoalexin present in red wine and grapes exhibits strong antioxidant activity (Tadolini et al, 2000; Belguendouz et al, 1997). An aqueous extract of terrestrial herb Selaginella delicatula compensate abnormally altered oxidative function and neurotoxicity caused by rotenone in mice (Girish and Muralidhara et al, 2012). A shrub Buddleja cordata extracts led to considerable reduction of ipsilateral rotations in rat, which was coupled with significant protection of DA levels and high reduction of lepidic fluorescent products in the striatum (Pérez-Barrón et al, 2015). It has also been shown that the aqueous solution of Gastrodia elata improves the movement dysfunction and exerts anti-inflammatory action following traumatic brain damage in rats (Ng et al, 2016).

2. Turmeric

Naturally produced plant products have been used in various categories to meet different needs. The natural products in plants are produced as secondary metabolites which are found to be increased when they are attacked by herbivores and pathogens suggesting their protective role. Most of the natural products consist of active compound that have biological importance and can be explored to develop pharmaceutical drug for treating disease and infections. Both during ancient and in modern cultures plant products have been an indispensable in developing medicines and thus taking an essential role in health care (Newman and Cragg, 2007). In Indian system of holistic cure, Ayurvedic medicine is practiced and passed on from one generation to another for more than 5000 years. Ayurveda, known as the 'science of life' focus in prevention of disease and restoring body natural systems, prolonging life by means of intervening in the way of life and through application of natural therapies (Garodia *et al*, 2007). In the same way, there is a traditional Chinese medicine which is also popularly practiced all over the East Asian countries. In addition to these, traditional medicine of Unani, Japanese, Egyptian medicine, Korean and traditional Native American medicine have been in practice. In every systems of traditional medicine the ground rules are use of medicinal flora in the management and prevention of any ailment and safeguarding good health (Acharya *et al*, 2008).

Of all the molecules introduced for drug development, some are synthetically produced while most of them are based from natural products. Drug development for human diseases has been based on natural products derived from plants, animals and minerals. Plant based drugs have proven to be suitable for human use and medicines developed from plant and its products keep on holding a vital role disease treatment all over the world.

In pharmacological research, many medicines are directly use from natural products or its derivatives. The knowledge of traditional medicines has enabled to serve as a foundation for several early medicines such as aspirin, digitoxin, morphine, quinine, and pilocarpine which are derived predominantly from plants. These compounds are an important resource for therapeutic areas of drugs, particularly for treatment against disease like cancer,

hypertensive, immunosuppressant, neurological disorder and other infection (Butler, 2004).

2.1. Uses of Turmeric

Besides its use in kitchen as popular spices, turmeric is also an important component in commercially manufactured food products such as beverages, dairy products, cereals, sauces and curry powder. Apart from coloring and adding taste to the food, turmeric has long been considered to possess medicinal value and effectively used in the indigenous system of medicine.

Turmeric (the common name for *Curcuma longa* L. of the Zingiberaceae family) is a rhizomatous herbaceous perennial plant that has a long history of association for medicinal use as a treatment for several ailments. Its use dates back since the Vedic age in Ayurveda in India where it is commonly used as kitchen spice derived from the rhizomes of the plant. In the region of Southeast Asia, apart from its popular use as kitchen spices, turmeric is also used in religious functions. In most of the South Asian countries it is widely cultivated and popularly used as an antiseptic and antibacterial agent.

The initial processing of turmeric includes washing, drying and grinding then sieving the powder to produce a uniform colored product. It is practiced as storing the turmeric powder in shade protected from sunlight which retards its rate of deterioration. In India turmeric crop is produced in large scale as compared to other countries and also consumes about 80% of it. Indian turmeric contains high amount of its bioactive compound K and as such are regard as the best quality available in the world. The turmeric production and popular trade Centre is located at Erode in Tamil Nadu state of India.

The active component of turmeric includes tetrahydro curcuminoid, K, dimethoxy K and bis-methoxy K and nutritional analysis showed that turmeric is rich in omega-3 fatty acid and α -linolenic acid (Goud *et al*, 1993). Traditionally it has been used to treat abnormal health condition such as biliary disorders, anorexia, coughs, diabetic wounds, hepatic disorders, rheumatism and sinusitis. It has been attributed numerous pharmacological properties including treatment for chronic frontal uveitis, conjunctivitis, skin disease, chicken pox, injury curative, urinary tract infections, and liver ailments (Dixit et al, 1988). The rhizome of turmeric is used as a calming agent, a diuretic, a hepato protectant and in the treatment of skin diseases such as scabies, leech bites and bruises (Kirtikar and Basu, Further it is reported as anti-inflammatory, choleretic, antimicrobial, and 1993). carminative actions. Turmeric extract has been shown to be effective for improving the condition of type-2 diabetes by restoring increase in blood glucose level (Kuroda et al, 2005). The yellow pigments curcuminoids which is a component in turmeric powder has been found to exert protective action against aflatoxin B1 (Gowda et al, 2008). It has also been suggested that turmeric and its active compounds have distinct antioxidants, antimutagenic, anti-tumorigenic, anti-carcinogenic, anti-inflammatory, anti-arthritic, antimicrobial and hypocholesterolemic properties; including action against diabetic, bacterial and fungal infections, protozoal and viral infections (Abbas et al, 2010; Miquel et al, 2002). Thus with its effective remedial properties on several diseases as suggested, currently turmeric is investigated for possible benefits in human diseases like neurodegenerative Ad, Cancer, Arthritis and other disorders (Henrotin et al, 2010). It improved the pancreatitis condition linked with lung damage in mice (Seo et al, 2011) and consists of an active component that is useful to fuel bile secretion and bile surge that will ultimately support and maintain liver in proper conditions (Dono, 2013).

K and curcuminoids account for 2-6% of turmeric. Turmeric as a herb or spice has much more to offer than just K. But for certain health conditions, high concentrations of K are required which can't be provided by the amount of turmeric one consumes on a daily basis. That is one of the prime reasons why K supplements are prepared when considered it in a health perspective.

Also, we cannot discount the fact that it is much easier to consume K supplements rather than thinking of ways to include turmeric in diet and manage the taste.

Following table shows some of the popular K products and their formulation available commercially.

Table 1: Commercial formulation and supplements of Curcumin (Turmeric for Health, 2018).			
Commercial Curcumin	Products		
Formulation			
Curcumin C3 complex	Sports Research Turmeric K C3 Complex,		
	Viva Naturals Non-GMO Turmeric K C3,		
	Doctor's Best K C3 Complex.		
Curcumin Bioperine	Schwartz Bioresearch Turmeric K Bioperine,		
	Doctor Danielle Turmeric K with Bioperine,		
	Nutravita - Organic Turmeric.		
Turmeric Curcumin	Source Naturals Turmeric with Meriva,		
Meriva Supplement	Thorne Research – Meriva.		
Longvida	NOW CurcuBrain,		
	Longvida by Nutrivenee.		
Curcumin Theracurmin	Natural Factors – K Rich Double Strength Theracurmin		
BCM-95 Curcumin	Terry Naturally CuraMed,		
	Progressive Labs - BCM-95.		

2.2. Clinical Studies of Turmeric

Clinical studies of turmeric have been done against several diseases. A study on Antimutagenic property of turmeric involving 16 chronic smokers was performed (Polasa et al, 1992). In this study, dose of 1.5 g per day were given to the subjects for one month. This gives rise to considerable decrease in urinary flow of mutagens. Among six nonsmoker subjects studied, there was no alteration in urinary excretion of mutagens, suggesting that turmeric consumption act as a strong antimutagen that can be explored for chemo preventive studies. Ethanol extracted from turmeric produced significant respite in patients with outside cancerous lesions by reducing smell in 90% of the case in a number of 62 patients studied and itching was reduced in almost all cases and about 10% of the subjects attained reduced in pain and size of lesion (Kuttan et al, 1987). Turmeric offered counter action against benzo-a-pyrene induced increase in number of micronuclei (Hastak et al, 1997). In a subsequent study, turmeric extract was given to oral submucous fibrosis (OSF) patients at 3g per day for the period of 90 days and the micronuclei amount was found to be reduced in oral exfoliated cells to a large extent comparable to the level of normal healthy individuals. In peptic ulcer patients, when turmeric filled capsules was given orally at dose of 2 capsule of 300mg each five times daily. Following treatment for 4 weeks, the results was significant with the ulcers diminished in 48% of the cases and by 12 weeks of treatment the reduction in ulcers increased to 76% (Prucksunand *et al.* 2001). In another study among patients suffering from irritable bowel syndrome, the occurrence was significantly reduced upon daily consumption of standardized turmeric extract tablet for 8 weeks; including relief from the abdominal pain/discomfort score (Bundy et al, 2004). Further, it has been shown that turmeric consumption mixed in curry help to increase bowel movement and activate bacterial flora that generates colon hydrogen thereby raising the hydrogen concentration of breath (Shimouchi et al, 2008). A paste of turmeric is often applied for preventing infection and wound healing. Traditionally, turmeric is also used as healing agent and natural protectant by applying on the cut umbilical cord after delivery (Alam *et al*, 2008).

Now multiple successful clinical trials have been completed using K.

Following table shows the successful trials of K on various diseases and the outcome.

Disease	No of Patients	ted in Curcumin (Gupta <i>et a</i> Dosage, duration	Outcome
Cancer	no of Patients	Dosage, duration	Outcome
Cuileer	15	0.036-0.18 g/day, 4 months	Reduced glutathione S-transferase activity
Colorectal cancer	15	0.45-3.6 g/day, 4 months	Reduced PGE ₂ production
	12	0.45-3.6 g/day, 7 days	Reduced the levels of M_1G
	5	1.44 g/day; 6 months	Reduced the number and size of polyps without any appreciable toxicity
	44	2 and 4 g/day; 1 month	Reduced ACF formation in smokers Improved body weight, reduced serum TNF-α, and
	126	1.08 g/day; 10-30 days	induced p53 expression Reduced the lipid peroxidation and increased GSH
	20	1.5 g/day; 6 weeks	content in patients Well-tolerated, limited absorption, and showed
	25	8 g/day	activity in some patients
	17 21	8 g/day; 4 weeks 8 g/day	Not feasible for combination therapy Safe and well-tolerated in patients
Breast cancer	14	6 g/day; 7 day, every 3 weeks	Safe, well-tolerated, and efficacious
			Reduced the serum PSA content in combination
Prostate cancer	85	0.1 g/day; 6 months	with isoflavones Decreased paraprotein load and urinary N-
Multiple myeloma	26	4 g/day; 6 months	telopeptide of type I collagen Safe, bioavailable, and efficacious against multiple
Inyeloma	29	2-12 g/day; 12 weeks	myeloma Reduced the urinary excretion of mutagens in
Lung cancer	16	1.5 g/day; 30 days	smokers
Cancer lesions	62	Ointment	Produced remarkable symptomatic relief in patients with external cancerous lesions
			Reduced the number of micronuclei in mucosal
	58	3.6 g/day, 3 months	cells and in circulating lymphocytes
	25 100	8 g/day, 3 months 2 g/day; 7 weeks	Improved the precancerous lesions Well tolerated, but not efficacious
·	100	2 g/day, 7 weeks	Increased vitamins C and E levels, decreased MDA
YY 1 1 1	75	1 g/day, 7 day	and 8-OHdG contents in the serum and saliva
Head and neck cancer	39	2 tablets	Decreased IKK β kinase activity and IL-8 levels in the saliva
Inflammatory diseas			
Crohn disease		1.08 g/day, 1 month + 1.44	Significant reductions in CDAI and inflammatory
	5	g/day,	indices in patients
Ulcerative	2 months	1.1 g/day for 1 month + 1.65	Significant reduction in symptoms as well as
proctitis	5	g/day for 1 month	inflammatory indices in patients
	89	2 g/day; 6 months	Prevented relapse of disease
Ulcerative colitis	1	0.5 g/day; 2–10 months	Associated with clinical and endoscopic remission of the disease
Inflammatory bowel disease			Suppressed p38 MAPK activation, reduced IL-1β, and enhanced IL-10 levels in mucosal biopsies;
	ex vivo	5–20 μM; 0.5–24 h	suppressed MMP-3 in colonic myofibroblasts
Irritable bowel syndrome	207	0.072 and 0.144 g STE/day; 8 weeks	Produced significant reduction in the prevalence of symptoms
	0		Increased bowel motility and activated hydrogen
Rheumatoid arthritis	8	0.5 g in food	producing bacterial flora in the colon Improved joint swelling, morning stiffness, and
	18	1.2 g/day; 2 weeks	walking time Improved the RA symptoms in patients alone and
	45	0.5 g/day; 8 weeks	in combination with diclofenac sodium Efficacious in the management and treatment of
Osteoarthritis	50	0.2 g/day; 3 months	osteoarthritis
	100	1 g/day; 8 months	Efficacious in the long-term management of osteoarthritis
Characteria			Efficacy and recurrence of the disease comparable to that for corticosteroid therapy without any
Chronic anterior	53	1.125 g/day; 12 weeks	adverse effect Reduced the eye discomfort after a few weeks of
uveitis			treatment in more than 80% of patients
uveitis Recurrent anterior uveitis	106	1.2 g/day; 12-18 months	
uveitis Recurrent anterior uveitis Postoperative			Exhibited superior anti-inflammatory property
uveitis Recurrent anterior uveitis Postoperative inflammation	46	1.2 g/day; 6 day	Exhibited superior anti-inflammatory property compared with phenylbutazone
uveitis Recurrent anterior uveitis Postoperative inflammation Gastric ulcer	46 60	1.2 g/day; 6 day 1 g/day; 6–12 weeks	Exhibited superior anti-inflammatory property compared with phenylbutazone Reduced ulcer formation after 12 weeks
uveitis Recurrent anterior uveitis Postoperative inflammation Gastric ulcer Peptic ulcer	46 60 45	1.2 g/day; 6 day 1 g/day; 6–12 weeks 3 g/day; 4 weeks	Exhibited superior anti-inflammatory property compared with phenylbutazone Reduced ulcer formation after 12 weeks Reduced ulcer formation Improved dyspeptic symptoms and reduced
uveitis Recurrent anterior uveitis Postoperative inflammation Gastric ulcer	46 60	1.2 g/day; 6 day 1 g/day; 6–12 weeks	Exhibited superior anti-inflammatory property compared with phenylbutazone Reduced ulcer formation after 12 weeks Reduced ulcer formation

3. Curcumin

Curcumin (K), 1, 7-bis (4-hydroxy-3-methoxyphenyl)-1, 6-heptadiene-3, 5-dione, is the main, active, yellow colored natural polyphenolic compound. It is taken out from rhizomes of *Curcuma longa* L. (turmeric), a plant in the ginger family (Zingiberaceae). K is widely used all over the world and extensively used in Southeast Asia (Goel *et al*, 2008; Aggarwal *et al*, 2007). Turmeric contains curcuminoids in several forms (Prasad *et al*, 2014). Various researches on K have showed that it also exhibits anti-inflammatory and anti-oxidant properties. Thus K is one such potential candidate that can be explored for therapeutic approach to several human diseases including neurodegenerative disorder like PD.

3.1. Curcumin Toxicity Studies

Several studies have suggested the concern about K safety under various experimental conditions. Reports suggesting that K may cause toxicity under specific conditions are shown with the studies in mammalian cell lines on treatment with turmeric in which there is a dose and time-dependent induction of chromosome aberrations (Goodpasture and Arrighi, 1976).

NTP Toxicology and Carcinogenesis Studies of Turmeric Oleoresin in 2-year feed studies on rats found that turmeric oleoresin consumption was linked with higher incidences of ulcers, hyperplasia, swelling of the forestomach, cecum, and colon in male rats and of the cecum in female rats (NTP, 1993). Further studies on reports of K concentration having beneficial effect have also demonstrated that it induce DNA damage and chromosomal alterations (Urbina-Cano *et al*, 2006; Cao *et al*, 2006). An experiment testing if the chelator action of K is adequate to cause iron deficiency in vivo, it was shown that K suppressed the production of hepcidin, a peptide which plays an essential role in regulation and balancing of systemic iron. It causes iron insufficiency leading to anemia in mice that are fed with poor iron diets (Jiao *et al*, 2009). K therefore has ability to influence systemic iron metabolism among individuals with suboptimal iron status (Means, 2009).

K also confer negative action activity of enzymes such as glutathione-S-transferase and UDP-glucuronosyl transferase apart from its hampering the action of drug metabolizing enzyme such as cytochrome P450 (Appiah-Opong *et al*, 2007; Thapliyal and Maru, 2001; Oetari *et al*, 1996). Such action of K may result in abnormal plasma flow of certain drugs that may lead to toxic action among the individuals taking K (Mancuso and Barone, 2009). Contradicting to the reports that suggest beneficial properties of K, a study suggested both advantage and disadvantage of K in alcoholic liver injury and concentration-dependent K toxicity was reported in animal models in which K was found to accelerate liver injury and liver cellular edema (Zhao *et al*, 2012).

In a study on human oral administration of K among the individuals with chronic anterior uveitis, all the patients who received K alone improved but those who additionally received antitubercular therapy had developed complications in their eyes and lost vision over time (Lal *et al*, 1999) suggesting that K may react with certain compounds resulting in detrimental effect. In another randomly assigned 8 week crossover study consisting eleven healthy subjects in the age group of 21-38 years old, it was reported that consuming turmeric as additional drugs markedly enhanced the levels of urinary oxalate which among the vulnerable subjects increases the probability of developing kidney stone (Tang *et al*, 2008). In a clinical trial of K for the prevention of colorectal neoplasia it was found that, overall, 61% of participants showed toxicity, mainly the gastrointestinal disturbances, most prevalently diarrhea, as well as swollenness, gastroesophageal reflux (Caroll *et al*, 2011). Another experiment aimed to establish the linkage between exposures of mixed

metals with child health outcomes due to association of lead with environmental sources such as spices, it was found that most of the turmeric samples had high level of lead and upon further study showed its increased bio-accessibility. Suggesting that turmeric powder upon contamination through environmental lead exposure can have potential lead toxicity and poisoning among some population (Gleason *et al*, 2014).

Therefore while developing therapeutic approach for a disease, it is important to thoroughly validate the toxicity as well as beneficial activity of prospective compound in the model organism. Taking this into consideration, I used wide range of K concentrations and assess its toxicity. In order to understand this paradigm I made an effort to assess the toxicity of K in fly model at different age groups.

4. Materials and Methods

4.1. Fly Stock

Drosophila melanogaster Oregon K strain were obtained from National *Drosophila* Stock Centre of University of Mysore, Karnataka, India. The flies were raised at 22°C–24°C in a *Drosophila* environmental chamber (Percival, DR41VL) and fed on a regular culture media prepared with sucrose, yeast, agar agar, and propionic acid. Male flies of were used in the present study.

4.2. Chemicals

Curcumin was procured from Sigma Aldrich (St. Louis, MO). Sucrose and Dimethyl Sulfoxide, were procured from Sisco Research Laboratory (India). Whatman filter paper no.1 disc was used as a feeding medium in the experiment.

4.3. Treatment Protocol

In filter disc treatment method volume of 250μ L of 5% sucrose was fed to the control flies and 250μ L of different concentrations of K solution were pipetted on a filter disc placed in 100x30mm glass vial. Twenty-five flies were put on each vial for the treatment and total of 100 flies were used for each concentration in the experiment. At every 24 hrs duration fresh media solution was prepared and the flies were transferred on it. Longevity curves were obtained from independent trials with a minimum of 100 flies per experiment until all the flies were recorded dead.

5. Results

5.1. Assessing Curcumin toxicity during Health stage and Transition stage of adult *Drosophila*

It has been shown that K act against acute concentration (20mM) of PQ to improve the survival rate and climbing defects (Park *et al*, 2012). Also contrasting to some findings suggesting the effectiveness of K, it has been reported that K exhibit a concentration-dependent toxicity in animal models (Zhao *et al*, 2012). Hence, I initially tested a range of K concentrations from 25 μ M to 50mM using 4-5 days old male *Drosophila* to assess potential deleterious effects. It was found that K exerts concentration dependent effect on the fly. The toxicity of highest concentration (50mM) was so high that all the flies were dead by 3 days of feeding. At 5mM concentration the files lived for 25 days; 35 days at 500 μ M; 40 days at 100 μ M. The toxicity effect of lowest concentration (25 μ M) was negligible and the flies lived up to 45 days under the experimental conditions. While K concentration of 2.5mM and above show toxicity effect on flies, concentrations below 2.5mM reveal no visible toxicity and there was no toxicity related death up to 10 days of feeding (Fig.1). Data was collected every 24 hrs for each group.

K toxicity experiment was also performed on the Transition span (55 days old) male flies with selected concentrations of $100 \,\mu$ M, 1mM and 1.5mM. It was found that there was no toxicity associated mortality up to 10 days of K feeding with all the selected concentrations in this age group (Fig.2). Data were collected every 24 hrs for each group. To assess the effectiveness of therapeutic molecules, several laboratories employ young animal models. They treat the animals for few days and estimate if the therapeutic molecule is efficient to shield the DA neuronal degeneration through assessment of behavioral assay like motor defect and biochemical markers like estimating the brain DA levels or cytological markers like DAergic neuron degeneration. Subsequently, all the experiments employed performed in this study used K concentrations of 2mM or less to avoid drug related toxicity.

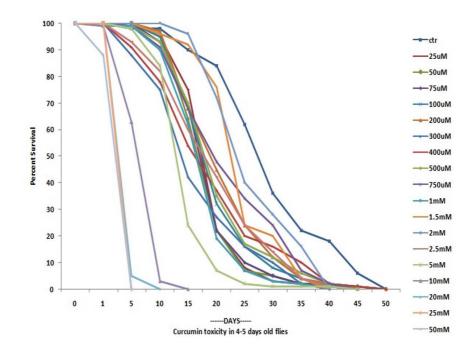


Figure.1. Curcumin toxicity in health stage (4-5 days old) of fly. Effect of increasing concentrations of K (25 μ M to 50mM) on survival of 4–5 days old male flies was concentration dependent toxicity. 4-5 days old flies were fed on fresh experimental medium of filter disc every 24 hrs and data were collected every 24 hrs for each group. Feeding of K concentration of 2.5mM or above has deleterious effect on viability of the fly, whereas concentrations lower to 2.5mM showed no observable toxicity (CTR=Control; K=Curcumin).

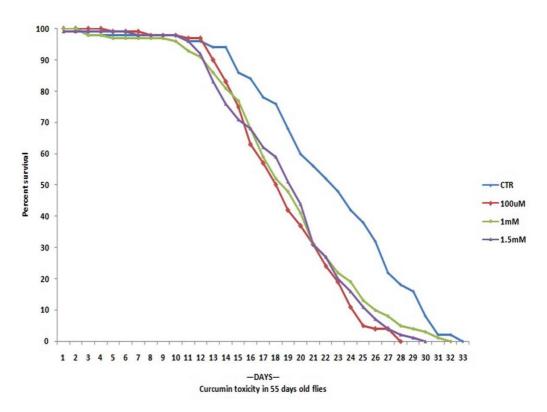


Figure.2. Curcumin toxicity curve in transition stage (55 days old) of fly. Effects of K concentrations (100 μ M, 1mM and 1.5mM) on survival of 55 days old male flies were investigated. The flies were aged in standard culture medium transferring to fresh media every 3rd day. At 55th day old, they were fed on fresh experimental medium of filter disc method every 24 hrs and data were collected every 24 hrs for each group (CTR=Control; K=Curcumin).

6. Discussion

Studies have suggested K toxicity under various conditions. K may react with certain compounds resulting in detrimental effect on certain individuals; enhance the levels of urinary oxalate which among the vulnerable subjects increases the probability of developing kidney stone (Tang *et al*, 2008). Turmeric powder may be environmentally contaminated by lead causing potential lead toxicity and poisoning among some population. K toxicity has also been studied suggesting that it primarily causes gastrointestinal disturbances, diarrhea, as well as distension and gastroesophageal reflux disease (Caroll *et al*, 2011).

In the present study, the first experimental approach employing K was to expose the model animal to a range of concentrations. I selected 18 concentrations ranging from 25μ M to 50mM of K in 4-5 days old male *Drosophila* and 100 μ M, 1mM and 1.5mM on 55 days old male *Drosophila* to assess possible detrimental effects of K.

It was found that K concentration of 2.5mM or above has deleterious effect on viability of the fly, whereas concentrations lower to 2.5mM showed no observable toxicity (Fig.1). In the highest concentration (50mM) all the flies employed in the experiment were dead by 5th day of K treatment, whereas 20% mortality was recorded in 2.5mM concentration on the 10th day of treatment. The concentration below 2.5mM did not show any mortality up to 10 days of exposure with K, indicating that there was no toxicity associated fly mortality in all the lower concentrations. K toxicity experiment was also performed on the 55 days old male flies (Fig.2). It was also found that there was no toxicity up to 10 days of K feeding on the fly which also indicates there was no toxicity associated mortality when treated with the selected concentrations of K.

Reports suggesting that K may cause toxicity under specific conditions are shown with the studies in mammalian cell lines on treatment with turmeric in which there is a dose and time-dependent induction of chromosome aberrations (Goodpasture and Arrighi, 1976). Further studies on reports of K concentration having beneficial effect have also demonstrated that it induce DNA damage and chromosomal alterations (Urbina-Cano *et al*, 2006; Cao *et al*, 2006). K has also been reported to cause anemia due to lack of iron content from low iron consumption in mice (Jiao *et al*, 2009) suggesting its ability to influence systemic iron metabolism among people with suboptimal iron status (Means, 2009). Further concentration-dependent K toxicity was reported in mice in which K was found to accelerate liver injury and liver cellular edema (Zhao *et al*, 2012).

7. Conclusion

From the present experimental results it is evident that increased in mortality rate among flies exposed to higher concentrations is due to K related toxicity. Therefore, after figuring out the concentration that is harmful to the fly, I wanted to use K as a neuroprotective agent in *Drosophila* model of sporadic PD.

This finding is important because for understanding any disease in animal model when the concentration of compound under investigation is not optimized, the phenotype or markers that the researchers find may be associated with its toxicity which can be wrongly interpreted. Therefore, in the subsequent experiments all the K concentrations that are used in rescue studies are sub lethal in all the age groups as shown in the result section.

CHAPTER IV

DECIPHERING THE NEUROPROTECTIVE EFFICACY OF CURCUMIN IN DROSOPHILA MODEL OF PARKINSON'S DISEASE

1. Introduction

A number of natural products with neuroprotective effects are assessed and reported by many researchers. Among them about 6000 research articles have suggested the multi therapeutic properties of K including potent antioxidant (Alhusaini *et al*, 2018; Trujillo *et al*, 2013; Dutta *et al*, 2005), anti inflammatory (Tizabi *et al*, 2014; Biswas *et al*, 2005), anticancer (Murray-Stewart *et al*, 2018; Singh and Khar, 2006), iron chelating (Yang *et al*, 2017; Du *et al*, 2012; Daniel *et al*, 2004) and neuroprotective activities (Huang *et al*, 2018; Ataie *et al*, 2010) in cell cultures and animal models (Yang *et al*, 2014). In addition, more than 100 clinical investigations have been done using K (Prasad *et al*, 2014). Apart from these properties, many other studies have reported the capability of K to slow down the key distinguishing features of PD like ROS accumulation, apoptosis, platelet assembly, cytokine production, cyclooxygenase lipoxygenase isoenzymes activities, repress oxidative injury, cognitive deficits in cell cultures and animal models (Yang *et al*, 2005). Therefore, it remains one of the natural products containing exceedingly promising curative properties that have been widely investigated by researchers for therapeutic approach to PD.

2. Antioxidant Properties of Curcumin

K exhibit potent antioxidant action that is similar to vitamins C and E (Toda *et al*, 1985). It has been reported to be a strong forager of a range of reactive oxygen species including superoxide anion radicals, hydroxyl radicals and protection of oxidative damage of kidney cells (LLC-PK1) by suppressing lipid degradation, lipid peroxidation and cellular breakdown (Cohly *et al*, 1998). K is also shown to inhibit the injury which is caused by hydrogen peroxide in human keratinocytes and fibroblasts (Phan *et al*, 2001) showing its antioxidant role in enhanced wound repair. Direct detoxification of reactive nitrogen species such as peroxynitrite by K has been demonstrated in vitro (Iwunze *et al*, 2004).

Treatment of DAergic neuronal cells and mice with K reinstate reduction of GSH levels, counter act protein oxidation and maintain mitochondrial complex I action that are generally affected because of low GSH activity (Jagatha et al, 2008). It is also suggested that iron induced primary cortical neurons toxicity was improved by K action through attenuating necroptosis (Dai et al, 2013). Out of 87 genes found to be differentially expressed in K treated Drosophila melanogaster, 50 genes were markedly up regulated and the rest 37 were down regulated (Zhang et al, 2015). They also suggested that K can be explored for an alternative therapeutic approach to developing drugs for age related disorder. Antioxidative properties of melatonin and K were tested in another experiment using liver of aging mice which showed that there was inhibition of protein carbonyls (PC) formation suggesting the health benefits of the compounds (Dkhar and Sharma, 2010). Also when K was fed on developing larva, it triggers pathways that help to extend the health span accompanied by marked increase in median and maximum longevities in adult fly (Soh et al, 2013). Recently it has also been reported in PINK1 knock down SH-SY5Y cell model that K rescue mitochondrial dysfunction and cell death (van der Merve *et al*, 2017).

3. Neuroprotective Role of Curcumin

3.1. Studies in Mice

Studies in mice showed that K attenuates loss of striatal DA axons triggered by 6hydroxyDA (6-OHDA) and reduces glial response and maintains SOD1 level in the 6-OHDA lesioned striatum (Tripanichkul *et al*, 2013). K treatment was also shown to diminish the A β 40, A β 42 and clumping of A β -derived diffusible ligands in the mouse hippocampal Cronu Ammonis-1area; reduced the expression of the γ -secretase component presenilin-2; upregulate the expression of enzymes that digest β -amyloid, insulin and neprilysin (Wang *et al*, 2014). Study on mice with liver lymphoma have shown the anticarcinogenic properties through initiation of antioxidant defense mechanisms and also hamper the process of angiogenesis by repressing the stress active genes and glycolytic pathway (Das et al, 2014). K also markedly diminishes the quantity of apoptotic cells and repressed the upregulation of cytochrome-c, caspase-9, and caspase-3, in an experiment to understand mitochondrial injury and oligodendrocyte apoptosis. They suggested the ability of K to inhibit apoptosis and thereby confer mitochondria protection (Feng et al, 2014). Study conducted on the effect of fluoride in 30 days old mice found elevated lipid peroxidation that was accompanied by high amount neurodegenerative cells in the hippocampal sub-regions. When K was co-treated with fluoride, lipid peroxidation was markedly decreased with simultaneous decrease in neurodegenerative cells, suggesting K is useful in ameliorating the effect of fluoride in mice brain (Sharma et al, 2014). In another investigation in adult and D-galactose induced old mice, K markedly decreased the escape latency and improved D-galactose induced decline of cellular proliferation and neuroblast differentiation in the sub granular zone of hippocampal dentate gyrus, amplified the levels of phosphorylated CREB (cAMP response element binding protein) and brainderived neurotrophic factor in the sub granular zone of dentate gyrus suggesting the efficacy of K to alleviate cognitive impairment through action on CREB signaling in the hippocampal dentate gyrus (Nam et al, 2014).

3.2. Studies in Caenorhabditis elegans

Studies in *Caenorhabditis elegans* (*C.elegans*) model also provide several advantages to investigate the link between OS and DAergic neurodegeneration in PD. Human α -synuclein over-expression in DAergic neurons in *C.elegans* showed age-dependent neuronal loss (Cao *et al*, 2005). Study on the effects of K has reported lifespan extension and reduction in intracellular ROS and lipofuscin during aging in *C. elegans*. This extension in longevity is credited to antioxidative properties of K but not its antimicrobial

activity (Liao *et al*, 2011). K feeding also improves egg generation, offspring size and survival of the worm which was positively associated with DAergic neuroprotection and attenuation of acetylcholine esterase activity (Satapathy *et al*, 2016).

3.3. Studies in Drosophila

Using *Drosophila*, it is reported that K have gender and genotype specific life span extension and sequester OS mediated free radicals, enhance locomotor ability and show chemo preventive property, suggesting its potential treatment applicability in higher organisms (Lee *et al*, 2010). K decreases death of SH-SY5Y human neuroblastoma cells induced by rotenone; improve characteristic symptoms associated with PD in *Drosophila* via reduction in intracellular and mitochondrial ROS levels and acting against the caspase-3/caspase-9 activity (Liu *et al*, 2013). Transgenic fly expressing human α -synuclein was exposed to different concentrations of K and found considerable delay in the loss of activity pattern, decrease in the level of OS and apoptosis, and extended life span (Siddique *et al*, 2014) suggesting potential role of K in neuroprotection.

4. Use of Curcumin in PD Studies

Several other laboratories have studied the effect of K in PD models. K has been reported to chelate iron, copper and other metals consequently hampering the α -synuclein or LB accumulation (Perez *et al*, 2008). In a study using the fly and two-dimensional nuclear magnetic resonance, it was found that K reduce toxic action via interaction with α synuclein oligomers and fibrils, adjusting the morphology and enhancing their separation (Singh *et al*, 2013) clearly suggesting that K act against α -synuclein clustering and stopping LB formation and attenuate α -syn oligomer toxicity. K also attenuates reduction in DA levels and degeneration of DA neurons (Mythri *et al*, 2012) and assist improvement of macro-autophagy through action on transcription factor EB, consequently controlling cell loss and toxicity to neurons (Jiang *et al*, 2013). In another study it is reported that K slow down activity of monoamine oxidase like that of the monoamine oxidase (MAO) inhibitor, thereby reinstating the diminish DA levels (Khatri *et al*, 2016) and decreasing depression (Nam *et al*, 2014). It is also shown to modulate OS, memory deficits, motor impairments (Cole *et al*, 2007) and decrease ROS levels, preserve mitochondrial integrity and attenuate neuroinflammation which consequently help to shield DA neurons in brain. K also impairs the c-Jun N-terminal kinases (JNK) pathway and thus avoids DAergic neuronal loss by the mechanism of apoptosis (Jayaraj *et al*, 2014). K is also reported to ameliorate oxidative injury to DAergic neuronal by activation of the Akt/Nrf2 pathway (Cui *et al*, 2016) suggesting its protective properties.

All these findings have shown the neuroprotective effectiveness of K by employing health stages of animal models. It may be noted that the investigation of different phases of life in *Drosophila* have shown that each life stages are distinguished by diverse pattern of gene expression. This pattern is comparable to equivalent life phase in human. It is also known that in *Drosophila* there is a marked variation of about 23% in genome-wide transcript patterns with age (Pletcher *et al*, 2002).

Therefore, it is important to follow the age specific study for late onset NDD such as PD. Although K has been shown to have protective effect in numerous studies as discussed above, there is an indispensable need to further decipher its action because all the model organisms employed in the studies are in health span stage.

Considering this fact, I try to further understand the action of K in both health and transition stage of *Drosophila*.

5. Materials and Method:

5.1. Fly Stock

Male *Drosophila melanogaster* Oregon K strain was used in the present study. The fly was obtained from National *Drosophila* Stock Centre of University of Mysore, Mysore, Karnataka, India. The flies were raised at 22°C-24°C in a *Drosophila* environmental chamber (Percival, DR41VL) and fed on a standard culture medium with composition of sucrose, yeast, agar agar, and propionic acid.

5.2. Chemicals

Curcumin and Methyl viologen dichloride hydrate or paraquat was purchased from Sigma Aldrich (St. Louis, MO). Dimethly Sulfoxide (DMSO), Sucrose was procured from Sisco Research Laboratory (India). Whatman filter paper no.1 disc was used as a feeding medium in the experiment.

5.3. Methods of Treatment

Two different methods of treatments are used for this experiment viz-

- 1. Co-feeding regime
- 2. Pre-feeding regime

5.3.1. Co-feeding Regime

For this regime, the flies are exposed to PQ and K simultaneously. That is one group of flies are treated with PQ while other groups are treated with PQ along with K at the same time. 10mM PQ was prepared by dissolving the PQ in 5% sucrose. Different concentrations of K (100μ M, 500μ M, 1mM, 1.5mM and 2mM) are prepared by dissolving the K in PQ solution. A volume of 250μ L of the same solution is pipetted into each vial containing filter disc at the bottom. The control flies remain in 5% sucrose while K *per se*

group was fed with 5% sucrose along with K. The flies were transferred to fresh vial every 24 hrs. The mortality and climbing ability were noted for every 24 hrs.

5.3.2. Pre-feeding Regime

In this regime, the flies are first fed with multiple concentrations of K (100μ M, 500μ M, 1mM, 1.5mM and 2mM) for 5 days. After feeding the flies with K, the flies are then exposed to 10mM PQ. The control and K *per se* group remain in 5% sucrose only. A volume of 250µL of the same solution is pipetted into each vial containing filter disc at the bottom. The flies were transferred to fresh vial every 24 hrs. The mortality and climbing ability were noted for every 24 hrs.

5.4. Negative Geotaxis Assay

The Assay was discussed in detail in Chapter 2, page no. 53.

5.5. Quantification of Dopamine Levels

The HPLC apparatus (Thermo Scientific Dionex Ultimate 3000) consisted of a XPG 3000 series pump feeding MD-TM mobile phase containing Acetonitrile, phosphate buffer and an ion pairing agent (Thermo Scientific, 70-1332) to the stationary phase built in BDS Hypersil C-18 column with a dimension of 150x3mm and a particle allowance size of 3µ (Thermo Scientific, 13741) connected to an ultra analytical electrochemical detector cell (ECDRS 1) followed by a Omni coulometric cell (ECDRS 2) to reduce noise during catecholamine detection. The sample was injected to the Column/ECDRS compartment by a temperature regulated Auto-sampler and the reduction and oxidation potential for ECDRS 1 was kept at -175 mV and 225 mV while ECDRS 2 was kept at oxidation potential of 500 mV. The acquisition of data was performed in Chromaleon-7 within a

gain range of 1μ A and was collected at a rate of 5Hz while keeping the data acquisition filter at 2.0 for the analytical cell.

For the sample, 15 heads were homogenized in 150µl ice cold phosphate buffer (0.1M, pH=7.4) and centrifuged for 10 minutes in 6000rpm at 4°C and supernatant was collected. 1:1 ratio of the supernatant and 5% TCA was again centrifuge in 5000rpm for 10 minutes 4°C. 50µl of the resulting supernatant was injected into HPLC column/ECDRS compartment. Data processing was accomplished using Thermo Scientific Chromaleon-7. Quantification was performed by integration of peak areas to pure catecholamine standards (Sigma-Aldrich) as obtained by processing through the HPLC.

6. Results

6.1. Curcumin rescues the mobility defects induced by PQ under Cotreatment and Pre-treatment regime during Health stage (4 days old) of *Drosophila* model of PD

To understand the efficacies of K to rescue the motory defects induce by PQ, as some of the studies have already reported (Inamdar et al, 2012), negative geotaxis assay also called as the climbing assay was employed. This assay will show the motor impairment due to action of PQ that serves as a perceptive display of degeneration of DAergic neuron and consequently the onset of movement dysfunction. As expected, PQ feeding made the flies to exhibit tremor at rest and bradykinesia when the motor ability was assessed after 24 hrs. These motor symptoms are the distinguishing features linked to PD in human. It was noticed that a number of flies tried to climb up the wall of climbing tube but they could not hold on the tube firmly and fall on the bottom of tube. Some flies showed restlessness with continuous wing flipping. The flies treated with PQ exhibited reduced motor function as seen by 30% decreased in their ability to climb at 12 seconds when compared to controls. Where as on those flies pre-treated with K or co-treated with K and PQ, the motor function were significantly rescued, as seen from the flies climbing higher distance when compared to PQ flies (Fig.2; 4). Comparision between the K only fed flies and controls showed that the distance flies travelled were comparable, indicating that K per se has no negative action on fly mobility. In the co-feeding regime where the motor function was assessed even at 72 hrs of exposure to PQ, the protective effect of K was consistent even at this time point (Fig. 2) which clearly affirms the effectiveness of K. Apart from co-treatment regime, inorder to understand if the neuroprotective efficiency is conferred through antagonistic interaction with PQ, the pre-treatment regime was employed. The flies were first fed with selected concentrations of K for 5 days followed by treatment with

PQ. When the flies were subjected to negative geotaxis assay, it was found that K attenuated the PQ-induced motor dysfunction even in this method of feeding, which is similar to the one observed in co-treatment regime (Fig. 4).

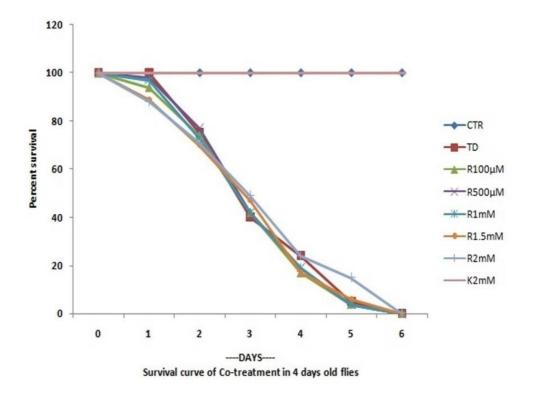


Figure.1: The survival curve of co-treatment in 4 days old flies (health stage). (CTR=control, TD=treated with PQ, R(rescue)=K+PQ, K=Curcumin).

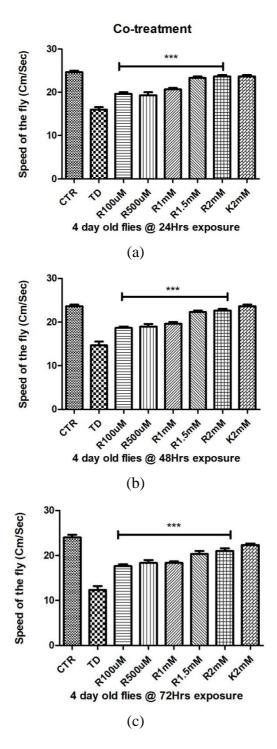


Figure.2: Negative geotaxis assay for co-treatment regime in 4 days old flies (health stage). K rescues the mobility defects induced by PQ under co-treatment regime during health span. 10mM PQ induces mobility defect in 4 days adult flies at different time points. The mobility defect is significantly altered when the flies are co-fed along with K (100 μ M, 500 μ M, 1, 1.5 and 2mM for 24hrs (a), 48hrs (b) and 72hrs (c). The ingestion of K alone does not make any difference in climbing ability of the fly when compared to control (***p<0.0001). (CTR=control, TD=treated with PQ, R (rescue)=K+PQ, K=Curcumin).

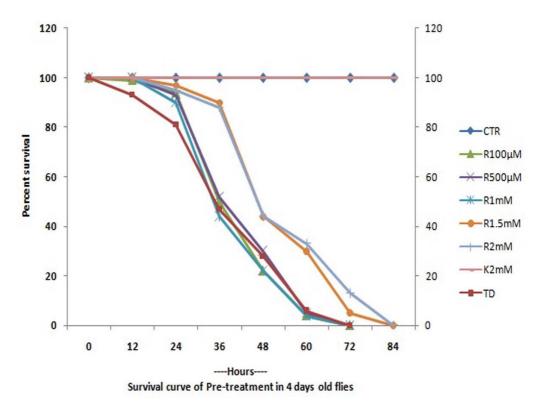


Figure.3: The survival curve of pre-treatment in 4 days old flies (health stage). (CTR=control, TD=treated with PQ, R (rescue) = K + PQ, K = Curcumin).

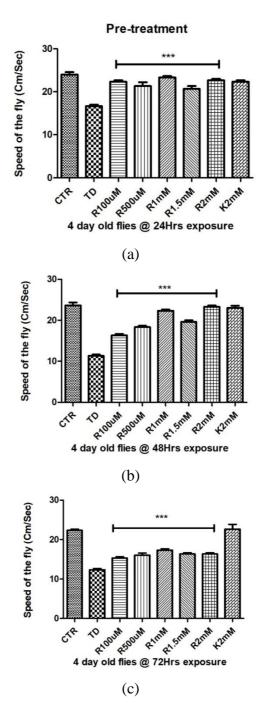


Figure 4: Negative geotaxis assay for pre-treatment regime in 4 days old flies (health stage). K rescues the mobility defects induced by PQ under pre-treatment regime during health span. 10mM PQ induces mobility defects in 4day adult flies at different time points. The mobility defect is significantly altered when the flies are pre-fed with K (100 μ M, 500 μ M, 1mM, 1.5mM and 2mM for 24hrs (a), 48hrs (b) and 72hrs (c). The ingestion of K alone does not make any difference in climbing ability of the fly when compared to control. (***p<0.0001). (CTR=control, TD=treated with PQ, R (rescue)=K+PQ, K=Curcumin).

6.2. Curcumin rescues the mobility defects induced by PQ under Cotreatment and Pre-treatment regime during Health stage (30 days old) of *Drosophila* model of PD

The health span ranges from adult eclosion from pupa to 30 days old. To proceed with assessment for K neuroprotection in transition phase, it is necessary to also check the negative geotaxis assay even in later part of health stage which will serve as study on wide window of health phase. Therefore, in order to have this confirmative knowledge, K was also tested in 30 days old (health stage) Drosophila and the flies were subjected to negative geotaxis assay after treatment of flies with both the feeding method. K could significantly modulate the motor dysfunction cuased by PQ even during this stage of health span in fly (Fig.6; 8). PQ exposure reduced the fly's motor ability by 30% when compared to control. The mobility defect is significantly altered when the flies are co-fed with K (100µM, 500µM, 1mM and 2mM) for 24hrs, 48hrs and 72hrs (Fig.6) respectively. Also in the pretreatment method the PQ induced motor defect was rescued by all the concentrations of K employed (100µM, 500µM, 1mM and 2mM for 24hrs and 48hrs (Fig.8) respectively. It was found that ingestion of K alone does not affect any difference in climbing ability of the fly when compared to controls in both the treatment methods employed. Therefore it is clear from the result that protective efficacy of K was not limited only to 4-5 days old but also efficient even at 30 days old flies in the health stage. The results confirm the neuroprotective efficacy of K during the health stage in *Drosophila* model of PD.

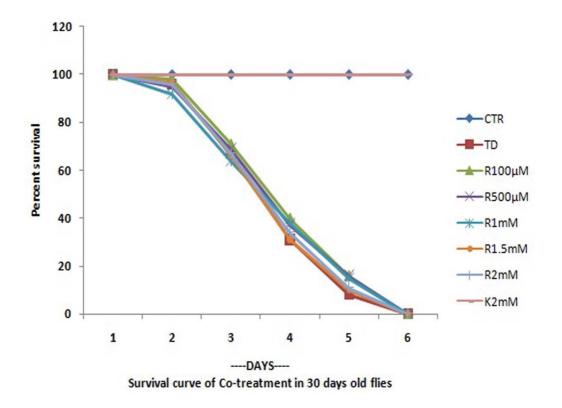


Figure.5: The survival curve of co-treatment in 30 days old flies (health stage). (CTR=control, TD=treated with PQ, R (rescue) = K + PQ, K= Curcumin)

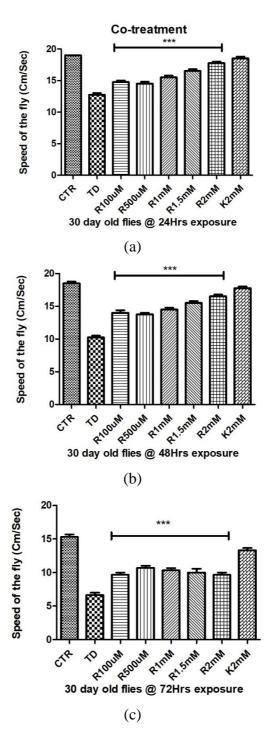


Figure 6: Negative geotaxis assay for co-treatment regime in 30 days old flies (health stage). Co-treatment regime for 30days adult flies for 24hrs (a), 48hrs (b) and 72hrs (c) indicates a similar pattern as that of the 5 day adult flies. The mobility defect induced by 10mM PQ is significantly altered when the flies are treated with PQ along with K concentrations of 100 μ M, 500 μ M, 1mM, 1.5mM and 2mM. Feeding K alone does not alter the mobility defect (***p<0.0001). (CTR=control, TD=treated with PQ, R (rescue)=K+PQ, K=Curcumin).

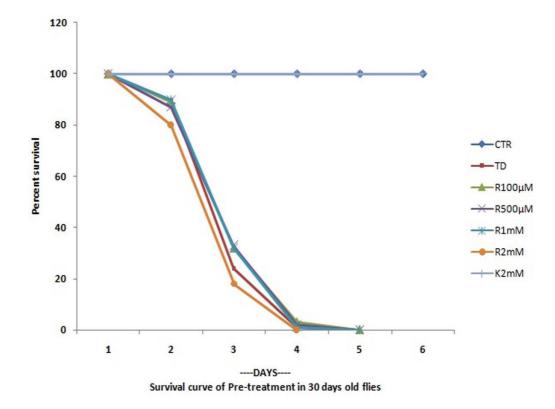


Figure.7: The survival curve of pre-treatment in 30 days old flies (health stage). (CTR=control, TD=treated with PQ, R (rescue) = K + PQ, K= Curcumin).

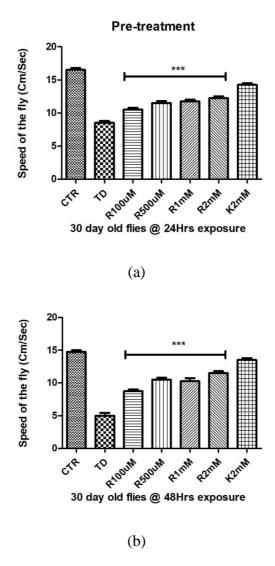


Figure.8: Negative geotaxis assay for pre-treatment regime in 30 day old flies (health stage). Pre-treatment regime for 30days adult flies for 24hrs (a) and 48hrs (b) indicates a similar pattern as that of the 4 day adult flies. The mobility defect induced by 10mM for is significantly altered when the flies are treated with PQ along with K concentrations of 100 μ M, 500 μ M, 1mM, and 2mM. Feeding K alone does not alter the mobility defect. (***p<0.0001). (CTR=control, TD=treated with PQ, R (rescue)=K+PQ, K=Curcumin).

6.3. Curcumin fails to rescue the mobility defects induced by PQ under Co-treatment and Pre-treatment regime during Transition stage of *Drosophila* model of PD

K efficacy was then tested in the age group of the transition phase (55 days old) with the same feeding methods employed for health stages (4-5 days and 30 days old). Interestingly, all the concentrations of K employed in the experiment could not improve the speed of the flies and the motor activity was similar between PQ fed flies and PQ+k groups in both co-treatment and pre-treatment regimen (Fig 10; 12). Importantly it suggests broad implications with regard to the neuroprotective effect of K and its therapeutic applications in NDD such as late onset PD that is manifested during the age period of 60 years in humans. Thus it is pertinent to properly screen and understand the efficacy of therapeutic molecules at the time point where the disease sets in.

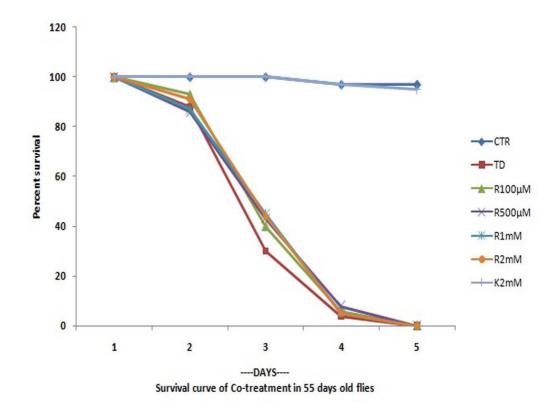
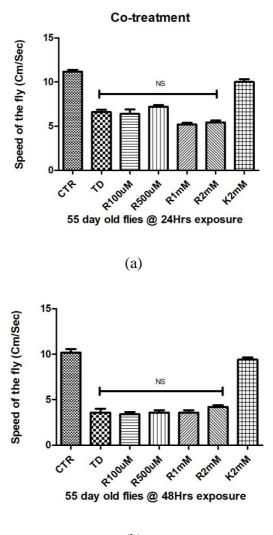


Figure.9: The survival curve of co-treatment in transition stage (55 days old flies). (CTR=control, TD=treated with PQ, R (rescue)=K+PQ, K=Curcumin).



(b)

Figure.10: Negative geotaxis assay for co-treatment regime in transition stage (55 days old flies). Co-treatment regime for 55 days old adult flies for 24hrs (a) and 48hrs (b) indicates a different pattern than that of the 4 day and 30 days adult flies. All the K concentrations used (100 μ M, 500 μ M, 1mM, and 2mM) fails to rescue the mobility defect induced by 10mM PQ. Feeding K alone does not alter the mobility defect (NS-Not Significant). (CTR=control, TD=treated with PQ, R (rescue)=K+PQ, K=Curcumin).

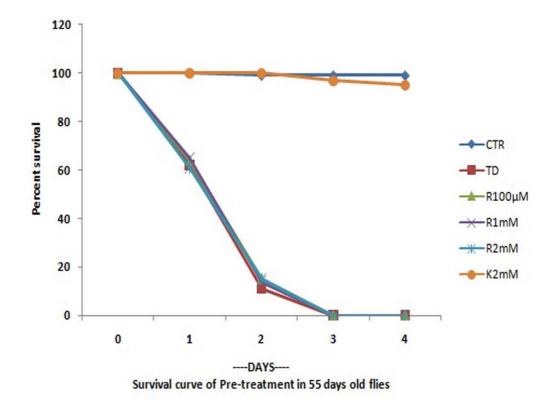


Figure.11: The survival curve of pre-treatment in transition stage (55 days old flies). (CTR=control, TD=treated with PQ, R (rescue)=K+PQ, K=Curcumin).

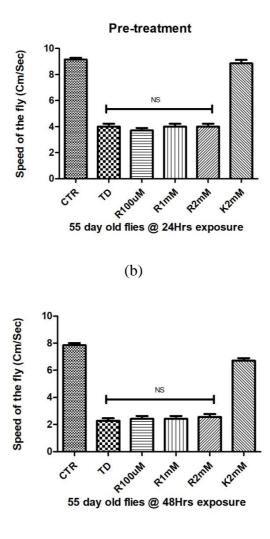
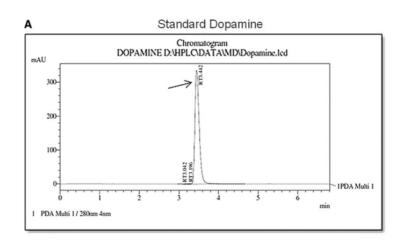




Figure.12: Negative geotaxis assay for pre-treatment regime in transition phase (55 days old flies). Pre-treatment regime for 55 days old adult flies for 24hrs (a) and 48hrs (b) indicates a different pattern than that of the 5 day and 30 days adult flies. All the K concentrations used (100 μ M, 1mM, and 2mM) fails to rescue the mobility defect induced by 10mM PQ. Feeding K alone does not alter the mobility defect (NS- Not Significant). (CTR=control, TD=treated with PQ, R (rescue)=K+PQ, K=Curcumin).

6.4. Curcumin could replenish decreased DA levels caused by PQ exposure in *Drosophila* during Health stage but not in Transition stage

It was found that although K clearly rescues mobility defect during health stages, it did not rescue the motor dysfunction during the transition stage. This raised the questions of whether the inability of K to rescue the observed climbing defect in transition stage is related to its failure to replenish the brain DA levels during that particular stage. Thus it was necessary to test the K effectiveness in DA activity at this phase. To confirm this phenomenon, employing the co-treatment method, I estimated DA levels in head tissue extracts of control flies and flies treated with PQ, PQ and K, and K alone in both the life stages. It was found that diminished brain DA levels were clearly replenished on K co-treatment during the health stage but it failed to replenish the decreased DA levels in transition stage (Fig.14). This result indicates the neuroprotective effectiveness of K could be in stage specific pattern where it may exert positive action at one stage but neutral or negative action during another stage of *Drososphila* life span. The present findings also observation suggests that K has limitation of its therapeutic efficacy in late onset NDD disease such as PD.



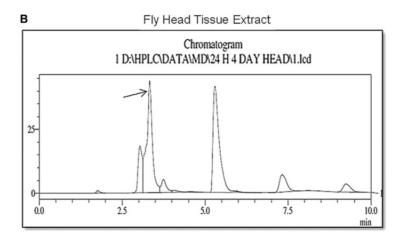


Figure.13: Quantification of brain DA level with high-performance liquid chromatography (HPLC). Chromatogram of standard DA showing a retention time between 3.196 and 3.442min (A) and chromatogram for *Drosophila* head tissue extract, showing a peak during the observed time window for standard DA (B) (peak for DA is pointed with an arrow in both the panels). Peaks can be seen (at 5min and 7.5min) much away from retention time window observed for standard DA that amount to artifact.

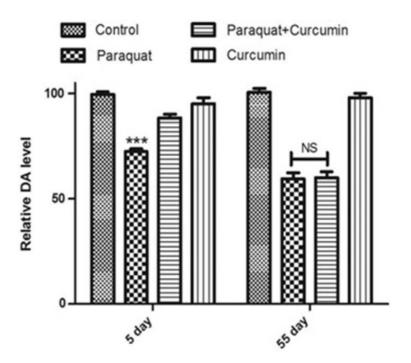


Figure.14: Relative DA levels in health stage (5 days old) and transition stage (55 days old) fly brains exposed to 10mM paraquat (PQ) for 24hrs. (***) p<0.0001 compared to control and K fed alone. NS, lack of significance between toxin treated group (PQ) and rescue group, i.e., PQ+K).

7. Discussion

In this chapter, multiple concentrations of K (100μ M, 500μ M, 1mM, 1.5mM and 2mM) along with 10mM PQ was employed to decipher the efficacy of K in mitigating the neurodegeneration against PQ induced mobility defects by employing co-feeding and pre-feeding regime.

I evaluated the PQ-induced locomotory deficits through negative geotaxis assay at 24 hrs, 48 hrs and 72 hrs time point after treatment. The experimental flies were exposed to 5% sucrose, 10mM PQ prepared in 5% sucrose, 10mM PQ with various K concentrations and K per se. Each experimental fly was allowed to climb up the tube towards the light source for 12 seconds. The total distance each fly travelled in 12 seconds was recorded both in co-treatment (Fig 2 and 6) and pre-treatment regime (Fig 4 and 8) in health stages (4 days and 30 days old flies). While control flies could climb up the climbing tube to over 20cm distance in 12 seconds, the patterns of movement of the flies treated with PQ was entirely different and their motor ability was reduced by 30% when compared with the controls. Some flies tend to remain at the bottom of the tube with signs of restlessness and others tried to climb up but would fall back to the bottom of tube implying that PQ caused neurotoxic effects in the fly, which was clearly manifested as impairment of locomotion ability. This observation is consistent with the previous report that PQ affect the behavior patterns in flies including altered climbing activity, continuous wing flipping, rotation and unsteady body and postural, that are identified characters as the human PD symptoms (Chaudhuri et al, 2007). The behavioral changes induced by PQ may be due to impairment of DAergic neuronal function and manifested through reduced climbing ability (Brooks et al, 1999). It has also been shown that PQ cause considerable death of DAergic neurons in flies (Chaudhuri et al, 2007).

Interestingly, there was no effect on the motor function of fly among the group to which K alone was fed, whereas ingestion of PQ alone negatively affected mobility as evident from inability of the flies to normally climb up the wall of climbing tube. K co-treatment with PQ showed recovery of the motor deficit as seen from its improved climbing speed (Fig 2 and 6). Even in the pre-treatment regime (Fig 4 and 8), it is clear that K effectively rescue the motor defects caused by PQ during all the tested time period of exposure. I observed the similar pattern of recovering the defect associated with PQ exposure in both the K pre-feeding and co-feeding regime, indicating that the efficacy of K is not due to its antagonistic interaction with PQ but by activation of molecular targets in the organism that confers protection to DAergic neurons.

However, in the case of transition stage, the phenomenon is completely different. All the concentrations of K used for the experiment could not rescue the mobility defect induced by PQ in both the co-treatment (Fig 10) and pre-treatment regimens (Fig 12) which is an interesting phenomenon.

It has been reported that there exist dissimilar patterns of gene expression at different life stages. About 23% in genome-wide transcript patterns variation are in *Drosophila* (Pletcher *et al*, 2002) has been shown indicating that natural compounds like K which has a genotropic action may not have its targets during the whole life period.

This finding was well substantiated by the results obtained from estimation of DA levels in health stage and transition stage (Fig.14). HPLC quantification of brain DA levels in health stage reveal that K co-treatment efficiently replenishes the PQ induced decreased level of DA. Whereas in the transition span there is no replenishment clearly indicating that targets of K is absent at this stage of *Drosophila* life span.

106

The property of K to alleviate expression intensity of age associated genes in young flies suggests the action of K on such genes is a cause of its longevity extending effects. These results signify that compounds like K may have definite positive effect at one stage while having neutral or detrimental action in another stage of adult life span. As it has been demonstrated that developmental or health phase feeding of K extends life span, however it shows harmful effect if fed for whole adult stages or during later stage such as the transition and senescent phases (Soh *et al*, 2013). They have demonstrated from the expression analysis that when flies are fed in stage specific manner pathways such as target of rapamycin (TOR) are affected, clearly signifying that K is a genotropic nutraceutical that confer its action by means of developmental stage-specific inducer of extended functional longevity in *Drosophila*. The genetic targets of K may be absent in some point of life span, implying its limitations with reference to a long-life phenotype.

8. Conclusion

Several studies employ co-treatment or pre-treatment method in young model animals. They treat the prospective molecules for a short duration and try to understand the effectiveness of therapeutic efficacy in PD models. They investigate if the toxin mediated DA neuronal toxicity is protected by the prospective therapeutic molecule through assessment of behavioral markers like motor defect, and biochemical indicator like the quantification of DA levels or cytological pointers like the degeneration of DAergic neurons.

The above findings demonstrate that K improves motor dysfunction caused due to PQ exposure in *Drosophila* during the health stages (4-5 days old and 30 days old flies) in both the co- and pre-treatment regimens. This motor dysfunction assay was assessed at 24

107

hrs time point after treatment where there was no apparent mortality seen in longevity experiment. K also recovers the motor defects associated with PQ exposure even at 48 hrs and 72 hrs of treatment, suggesting its underlying neuroprotective effectiveness. On the contrary, K could not improve the motor dysfunction in transition phase (55 days old flies) clearly signifying its constraint as a therapeutic agent in late onset NDD such as PD.

Considering the present experimental findings, I further try to understand the mechanism of K neuroprotection by looking into the OS markers, antioxidant and neurotransmitter enzyme activities employing both the health and transition span of *Drosophila*.

CHAPTER V

MECHANISTIC INSIGHTS INTO THE THERAPEUTIC PROPENSITY OF CURCUMIN IN *DROSOPHILA* MODEL OF PARKINSON'S DISEASE

1. Introduction

Oxidative Stress (OS) is classically defined as a state of redox imbalance caused by an excess formation of oxidants or a defect in antioxidants (Sies, 2015). It has been proposed that OS act as one of the factors that possibly play a role in the pathogenesis of neurodegenerative disorders. Clinical and preclinical studies point out that neurodegenerative diseases like PD are characterized by increased level of OS biomarkers and by decreased level of antioxidant defense biomarkers in the brain and peripheral tissues (Niedzielska *et al*, 2016).

In normal physiological state the tissues have a functional antioxidative system, GSH that is depleted due to OS. Unwarranted increase and accumulation of free radicals lead to imbalance of cellular oxidative homeostasis and decrease of GSH concentration contributes in OS and consequent brain damage (Jain *et al*, 1991). Physiologic maintenance of the redox potential is critical for normal functioning of neurons and disruptions on this balance interfere with a number of biological processes, which eventually lead to cell death. A number of machinery that are responsible for ROS production are known which include the metabolism of DA itself, mitochondrial impairement, iron, neuroinflammatory cells, calcium, and aging (Dias, 2013).

The mechanism leading to OS due to the action of ROS can be via the intracellular change in redox potential and oxidative modification in the protein molecules. Such alteration in redox potential of intracellular cystol of the cells take place under reduced conditions due to redox buffering capacity of intracellular thiol groups like GSH and thioredoxin (TRX) (Ravindran *et al*, 2012). The oxidized form of GSH and TRX are maintained in stable ratio by their reductase enzyme activities. Both of these enzymes are responsible for the reduction of hydrogen peroxide and lipid peroxidase that are carried out by the peroxidase enzymes. They have the ability to act as the antioxidant enzymes and are involved in the cell signaling mechanism (Sesti *et al*, 2010). GSH also perform redox signaling process by transporting out the differentiation in both the total level of GSH and also in the ratio of its oxidized to reduced forms. Under physiological condition the reduced form of GSH is higher than its entire oxidized species (Aquilano *et al*, 2014).

Further, the ROS have been accounted for the structural modification in protein molecules by causing protein dimerization and also by altering the molecular structure in amino acid residues during the second stage of oxidase modification in protein molecules (Ravindran *et al*, 2012). The protein molecules are altered oxidatively in ways that include the cysteine residue. The sulfhydryl group (-SH) gets oxidized to produce the sulfenic (-SOH), sulfinic (-SO₂H), sulfonic (SO₃H) moieties. Such modification results in alteration of enzyme biological functions, when cysteine is located inside the catalytic domain (Wu *et al*, 2008).

2. Parkinson's Disease and Oxidative Stress

Oxidative modifications of enzymes and structural proteins play a significant role in the etiology and progression of several human neurodegenerative diseases. Accumulation of free radicals and subsequent neurodegeneration in specific brain regions have been proposed as the underlying factors in neurodegenerative diseases such as Alzheimer's and PD (Halliwell, 2006).

Studies on post-mortem PD brains link the role of oxidative injure in the progression of PD (Yuan *et al*, 2016; Zeevalk *et al*, 2008; Bosco *et al*, 2006).

3. Oxidative Stress and its Markers

3.1. Non Enzymatic Markers

3.1. 1. Reactive Oxygen Species (ROS)

ROS are molecular entities that react to cellular components which result in detrimental effects on their function. Neurons and glias are prone to produce ROS as an inevitable outcome of oxygen dependent respiration (Dumont *et al*, 2011). ROS can be generated via a number of pathways which may include direct interactions between redox active metals and reactive oxygen species through the Fenton and Haber-Weiss reactions; through indirect pathway via activation of enzymes such as nitric oxide synthase (NOS) or nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidases). As a common principle, the chemical origin of the majority of free radicals requires the activation of molecular oxygen (Smith *et al*, 2007). Examples of ROS include the superoxide ($O_2^{2^-}$), hydroxyl (OH.), peroxyl (RO₂.), alkoxyl (RO.), hydroperoxyl (HO₂.), and nonradical species such as hydrogen peroxide (H_2O_2), hypochlorous acid (HOCl), ozone (O₃), singlet oxygen (1O₂), and peroxynitrite (ONOO-).

Several investigations have suggested the ROS to be the major factor contributing to DAergic neuronal loss in the PD brain, as a consequence of DA metabolism, decreased GSH activity, and elevated levels of iron and calcium in the SN*pc* (Jenner and Olanow, 2006).

The problem with ROS arises when free radical concentration is more than the antioxidant concentration and results in programmed cell death (Brieger *et al*, 2012). Thus in order to prevent the subsequent OS damage of cell, the inhibition of ROS must be established. Apart from estimation of general ROS level, several enzymes and non enzymatic function are studied as biomarkers to understand their activity variation under normal and disease conditions.

111

3.1.2. Lipid Peroxidation (LP)

LP is another common illustration of oxidative damage in cell membranes, lipoproteins, and other lipid-containing molecules. Peroxidative modification of unsaturated phospholipids, glycolipids, and cholesterol can take place in reactions triggered by free radical species such as oxyl radicals, peroxyl radicals and hydroxyl radicals derived from iron-mediated reduction of hydrogen peroxide or by non-radical species like singlet oxygen, ozone, and peroxynitrite generated by the reaction of superoxide with nitric oxide. Lipid hydroperoxides are well known non-radical intermediates of lipid peroxidation and its identification can often give essential mechanistic details such as whether a primary reaction is mediated by singlet oxygen or oxyradicals. Unsaturated phospholipids, glycolipids, and cholesterol in cell membranes and other organized systems are major targets of oxidant attack. This can result in lipid peroxidation, a degenerative process that disturbs structure and function in the target systems resulting in cytopathological consequences (Halliwell and Gutteridge, 1990; Girotti, 1985). Several products from lipid peroxidation are chemically reactive and are thought to be the major effectors of tissue damage (Mattson, 1998). Lipid peroxidation is measured by levels of MDA which is an intermediate compound and a major pointer of lipid peroxidation. The brain has high concentrations of polyunsaturated fatty acids that cause peroxidation of lipids and in the process accumulates toxic products when there is a condition of OS.

3.1.3. Protein Carbonyl (PC)

PC content is the most general and well used biomarker of severe oxidative protein damage. Carbonyl groups (aldehydes and ketones) are produced on protein sidechains especially of Proline, Argenine, Lysine, Threonine, when they are oxidized (Fig.1). These moieties are chemically stable, and important for both their detection and storage.

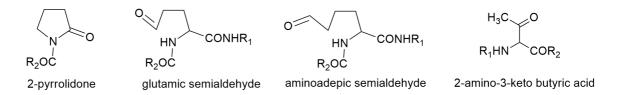


Fig.1. The structure of carbonyl derivatives produced by direct oxidation of amino acid side chains: 2-pyrrolidone from prolyl residue, glutamicsemialdehyde from arginyl and prolyl residue, a-aminoadipicsemialdehyde from lysyl residue, and 2-amino-3-ketobutyric acid from threonyl residue (Dalle-Donne *et al*, 2003).

Carbonylation of proteins is a permanent oxidative injury resulting in functional loss often forming a clumping of damaged unfolded proteins. Such aggregates can inhibit proteasomal function which is directly linked to the increase in aggregates of carbonylated proteins in tissues that are resistant to proteolytic process. Uneven protein structures and folding that arise due to functional impairment are closely associated with several human diseases (Bossy-wetzel *et al*, 2004). Bioinformatics Analyses on pesticide treated mice showed that protein carbonylation affects cellular junctions, cytoskeleton, and the proteasome (Coughlan *et al*, 2015). Elevated levels of protein carbonylation associated with PD have been reported (Floor *et al*, 1998). In some case, this elevations correlate well with the progression or severity of the disease.

3.1.4. Hydroperoxides (HP)

HP is one of the major modifications of organic compounds under the influence of wellrecognized reactive oxygen species. HP are the intermediate to produce more reactive oxygen species and are well established biomarkers for OS. Commonly used HP bio markers are protein HP and lipid HP. Studies indicate that proteins are most likely to be among the first targets of ROS cells and protein HP is the main products of this interaction. Formation of Protein HP has an important biological implication since it results in modification of the structure and properties of amino acid residues. Moreover, HP groups are powerful oxidants that can induce secondary damage through inactivation of thioldependent enzymes essential to cell functions. Hydroxyl radical (•OH), superoxide radical anion (O²*), and singlet oxygen (1O₂) are the major contributors to the overall amount of protein HP formed in living organisms. This ROS modify the amino acid residues with an addition of HP group (-OOH). The protein HP (PrOOH) has a tendency to react with the metals and proteins and induces DNA damage, deactivation of enzyme, free radical formation (Fig.2).

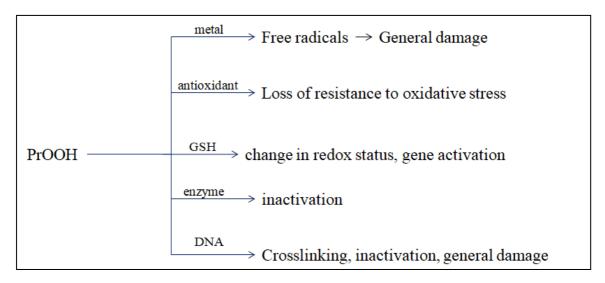


Fig.2. A summary of the reactions of protein hydroperoxides and their potential consequences in vivo (Gebicki, 1997).

3.2. Antioxidant Enzymes

3.2.1. Superoxide Dismutase Enzyme (SOD)

SOD which convert superoxide to hydrogen peroxide (H_2O_2) in the cytoplasm and mitochondria are often regarded as the first line of defense against ROS (Zhou *et al*, 2008) among the ROS-scavenging enzymes. They convert naturally occurring superoxide radicals to molecular oxygen and hydrogen peroxide. Three different SOD isoenzymes, that are well compartmentalized, have been characterized in humans (Zelko *et al*, 2002). Study on PD brain showed an increased superoxide dismutase like activity most prominent in SN and basal nucleus (Marttila *et al*, 1998). Post mortem studies in PD showed increased activity of encephalic superoxide dismutase (Taylor *et al*, 2012). Also recent study on patient's blood has reported that PD is positively associated with increased levels of SOD activity (de Farias *et al*, 2016).

3.2.2. Catalase (CAT)

CAT is known as a major antioxidant enzyme that neutralizes the harmful effects of ROS and as such, is considered beneficial in the treatment of many diseases. CAT protects cells by separation of hydrogen peroxide into water and oxygen and by prevention of hydroxyl radical generation. Thus it helps to remove the oxygen free radicals, which is essential for maintaining a stabilized redox status of cells (Halliwell and Gutteridge, 1990). Study on CAT-loaded, poly (lactic co-glycolic acid) nanoparticles (NP) in human neuronal protection against oxidative damage showed very efficient CAT encapsulation capable of retaining ~99% enzymatic activity. The NP-mediated CAT delivery effectively protected cultured neurons from H₂O₂-induced OS. CAT loaded nanoparticles significantly reduced H₂O₂-induced protein oxidation, DNA damage, mitochondrial membrane transition pore opening and loss of cell membrane integrity and restored neuronal morphology, neurite network and microtubule-associated protein-2 levels (Singhal *et al*, 2013).

3.2.3. Glutathione S-Transferase (GST)

GST is a group of enzymes important in the detoxification of many xenobiotics in animals. They guard cells against toxicants by conjugating the thiol group of the GSH to electrophilic xenobiotics, and thereby protect cells against the mutagenic, carcinogenic, and toxic effects of both exogenously and endogenously derived compounds (Hayes and Strange, 2000). GST activity is present in plants, insects, yeast, bacteria, and most mammalian tissues particularly in the liver, which plays a key role in detoxification.

Human GSTs are a functionally diverse family of soluble enzymes of detoxification that use GSH in conjugation and reduction reactions. Toxic electrophiles, including a variety of carcinogens, are substrates for the GSTs and after conjugation or reduction they are more easily excreted into bile or urine. There are at least 42 GST-like genes in *Drosophila melanogaster* belonging to six putative families including genes from the Delta, Epsilon and Sigma classes (Rubin *et al*, 2000).

3.2.4. Reduced Glutathione (GSH)

GSH is an important antioxidant enzyme capable of preventing the ROS mediated cellular damage. GSH activity loss is linked with incidental LB disease and it may represent the initial biochemical marker of nigral cell loss. GSH depletion alone may not result in damage to nigral neurons but may increase susceptibility to subsequent toxic or free radical In a study to access the alterations in GSH levels in PD and other exposure. neurodegenerative disorders affecting basal ganglia, GSH and oxidized GSH activity were monitored in various brain areas from patients dying with PD. GSH levels were found to be reduced by 40% as compared to control subjects in SN in PD patients and oxidized GSH levels were insignificantly elevated while there were no changes in other brain areas. In spite of severe nigral cell loss in the SN in PD, multiple-system atrophy, and progressive supranuclear palsy, significant reduction in GSH level was seen only in PD. The result suggests that nigral cell death alone is not responsible for alteration in GSH level (Sian et al, 1994). Decreased synthesis due to inhibition of GSH reductase or from increased levels of GSH disulfide (GSSG) and change the GSH: GSSG ratio could be the cause for the drop down in GSH level (Genestra, 2007).

3.2.5. Total Thiols

Thiol redox balance regulation is crucial for multiple cellular metabolic, signaling and transcriptional processes. Thiol groups are highly reactive and susceptible to oxidation that may cause significant loss of biological activity. In proteins, oxidation of free thiol groups produces modifications that may impact on the structure, catalytic activity or ability to engage in protein-protein interactions depending on their location. An important function of cell based thiol redox buffering systems is to protect thiol groups from oxidation and to repair those that may have become oxidized as a result of normal or aberrant cellular metabolism. The key components of the thiol redox buffering system are the cysteine/cystine and GSH/ GSH disulphide redox pairs, and the thiol disulphide oxidoreductases that include thioredoxin (Trx), glutaredoxins (Grx) and peroxiredoxins (Prx). Grx1 plays an important role in maintaining cell viability in model DAergic neurons in culture (Rodriguez *et al*, 2012). In vitro and in vivo study of potential impact of Grx1 upregulation in the CNS on DAergic viability suggests that the Grx1 upregulation promotes toxic neuroinflammation, potentially contributing to PD (Miller *et al*, 2016).

3.3. Neurotoxicity Marker

3.3.1. Acetylcholinesterase (AChE)

AChE is one of the most efficient enzymes which is concentrated at the cholinergic synapses throughout the CNS and at neuromuscular synapses where it rapidly hydrolyses the neurotransmitter acetylcholine (Ach) into choline and acetate thus playing an essential role in cholinergic neurotransmission (Tripathi and Srivastava, 2008). AChE enzyme is present in high concentration in all types of conducting tissue, nerve and muscle, central and peripheral tissues, motor and sensory fibers, sympathetic and parasympathetic so called cholinergic and noncholinergic fibers and all the regions where cell bodies and junctions are located (Nachmensohn, 1959). AChE plays an essential role in

acetylcholine-mediated neurotransmission. It is neurotransmission activity, rather than reuptake by transporters as with other neurotransmitter systems, that terminates cholinergic neurotransmission (Massoulie *et al*, 1993). The importance of AChE in mammals is illustrated by the effect of abrupt blockade of AChE catalytic activity, such as by exposure to the nerve gas sarin. Within minutes, inhibition of AChE leads to excess acetylcholine at neuromuscular synapses, continued activation of acetylcholine receptors, subsequent receptor inactivation, respiratory and/or cardiac dysfunction and death (Karalliedde, 1999). Therapeutically, controlled application of AChE inhibitors is used to increase synaptic levels of acetylcholine in diseases that impair acetylcholine neurotransmission.

4. Aging as a factor for Oxidative Stress

Protein, lipids and nucleic acids are biological macromolecules which are oxidatively altered due to ROS, resulting in genetic mutations and cellular senescence (Luceri *et al*, 2017). In complications and pathologies associated with aging, increasing evidence for causal role of cell senescence have been experimentally shown (Correia-Melo *et al*, 2015). Due to stress during aging there is increase in cell senescent. This phenomenon is controlled by a number of counting mechanisms including telomeres shortening, gradual increase in damage of DNA, deviation in normal oncogenes behavior, change in metabolic action and increased ROS generation (Kuilman *et al*, 2010). Such processes lead to impairment of cellular function and thus produce and accumulate ROS which in turn disturb the homeostasis, a vital factor for safeguarding the senescent phenotype. Apart from injuring DNA and damage mitochondrial activity, OS trigger the activity of p53, a gene coding for cell cycle regulator protein, which in turn induces prooxidant genes. Disturbances in redox homeostasis by generating ROS lead to deterioration of cell and impair its function.

To limit over-accumulation of ROS in the body, there exist both enzymatic and nonenzymatic systems that maintain ROS balance. Reports have also shown that natural antioxidant defense systems have limited capacity in the brain as compared to peripheral tissues (Somani *et al*, 1995; Del *et al*, 1987) and buildup of molecular oxidative damage, initiated by ROS is the main causal factor underlying senescence-associated losses in physiological functions (Sohal, 2002).

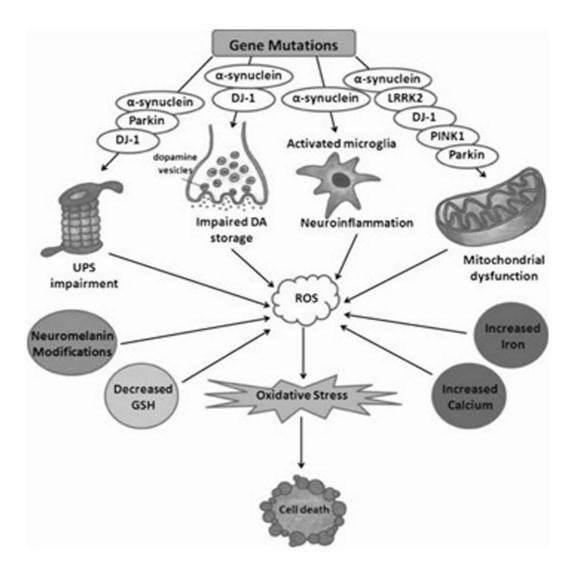


Figure.3. Mechanisms leading to Oxidative Stress in PD and the role of PD-related gene products in this process. DA = dopamine; ROS = reactive oxygen species; UPS = ubiquitin-proteasome system; GSH = glutathione (Adapted from Dias *et al*, 2013).

5. Studies on Efficacy of Neuroprotective Compounds using Fly Model

Using OS induced depletion as a gold standard, many laboratories have studied neuroprotective efficacy of multiple compounds showing modulation of biochemical markers, antioxidant enzymes and phenotype associated rescue on exposure to neurotoxic agents and suggested for therapeutic approach (Jia *et al*, 2018; González-Burgos *et al*, 2017; Anandhan *et al*, 2012; Khan *et al*, 2010; Rajasankar *et al*, 2009; Rojas *et al*, 2008; White *et al*, 1996).

Studies have also shown a number of natural compounds and molecules derived from them have the ability to protect cells from OS and ameliorate various OS-related diseases. Some of the list of pathways through which neuroprotective compounds confer neuroprotection are given in table 1.

Table 1: Listing of pathways through which neuroprotective compounds confer neuroprotection: Lessons from *Drosophila* model of PD (*All treatments were administered as dietary complement) (Ayajuddin *et al*, 2018)

Pathway/ Process	Compound treatment	Drosophila model	Modified phenotype/s
¥	Sulforaphane and	Parkin	DA neuron number
	allyldisulfide		
	S-methyl-Lcysteine	α-synuclein	Locomotor activity
	Polyphenols	α-synuclein	Lifespan, locomotor
			acitvity
	α-tocopherol	PQ and iron	Locomotor activity
	SOD	DJ-1β	Lifespan
Oxidative Stress	Melatonin	PINK1	Lifespan
	Bacopa monieri	PINK1	Ommatidial degeneration
	leaf extract	DJ-1β	Ommatidial degeneration
		PQ	Lifespan, locomotor
		_	activity
		Rotenone	Locomotor activity, DA
			neurons
		PQ	Oxidative marker levels
Oxidative Stress /	Miocycline	DJ-1a	DA neuron numbers, DA
Inflammatory process	Celastrol		levels, locomotor activity
			and survival under OS
TOD signalling	Danamusin	Doultin /DINI/ 1	condition Thorasis indeptations
TOR signalling	Rapamycin	Parkin/PINK1	Thoracic indentations, locomotor activity, DA
			neuron number and
			muscle integrity
Removal of excess or toxic	Geldanamycin	α-synuclein	DA neuron number
protein forms	Geldununiyeni	a synderenn	Di incuroni number
Zinc homeostasis	Zinc chloride	parkin	Lifespan, locomotor
		r	activity, and percentage of
			adulthood survivors
Chaperone therapies (HSF-1	Celastrol Carbenoxolone	α-synuclein	DAergic neuroprotection
modulators) Trigger HSF-1		α-synuclein	
activation Induces			
downstream Hsp70 expression			
Hsp90 inhibitors Inhibits the	Geldanamycin	α-synuclein	decrease α- synuclein
interaction between Hsp90	17-AAG		aggregation and reduce
and HSF-1, leading to	17-DMAG		cell toxicity
increased Hsp70 expression	SNX-2112		
and activity			
mTOR-dependent	Metformin	Drosophila	Reduced cell death
pathways/AMPK	AICAR	melanogaster mutated	
		for LRRK2	
mTORC1	Rapamycin and Rp	Drosophila	Reduced mitochondrial
	analogues (CCI-779,	<i>melanogaster</i> mutated	dysfunction
	RAD001 and AP23573)	for PINK-1 and Parkin	Deduced meet
mTor-independent	Spermidine	α-synuclein	Reduced motor
pathways/unknown			dysfunction, increased lifespan; Reduced
			neuronal cell loss
LRKK2 kinase inhibitors	GW5074, and sorafenib	α-synuclein	Protect against DA neuror
	G w J074, and solatenild	u-synucienn	degeneration locomotor
			activity
Histone Deacetylase inhibitors		α-synuclein	Protect against DA
instone Deacetylase minonons		u-synucioni	locomotor activity
Antitumor agents	Geldanamycin	α-synuclein	Protect against DA
	Sciumuniyeni	a synderenn	Mobilized the stress
			response and increase
	1		levels of chaperon HSP70

6. Necessity of Stage Specific studies in *Drosophila* Model of PD

For understanding the pathology of late onset neurodegenerative diseases, it is important to employ organisms belonging to adult stages/phases where the disease sets in, such as the transition stage of Drosophila PD model. Because different stages of the life stage have different patterns of gene expression and the cellular, physiological and molecular phenomenon at old stage may be completely different from young stage. Studies in Drosophila using expression analysis has demonstrated that there are about 23% genomewide transcript patterns variations (Pletcher *et al*, 2002) indicating that the genetic targets of a particular therapeutic molecule may be present at one stage but lacking in another stage of life span. Therefore it may exert positive action at one stage and neutral or negative action in another stage. Genotropic drugs will only exhibit its efficacy in the stage of life cycle where their target molecules are present (Soh et al, 2013). Hence, it is possible that the targets of K may be absent in some stages of life, which is a vital and interesting paradigm. But there are no reports showing the effectiveness of K during the later stages of adult fly life. Therefore, it is necessary to understand the neuroprotective efficacy of compounds at transition phase because this is the period during which late-onset neurodegenerative diseases such as idiopathic PD sets in.

7. Neuroprotective Studies of Curcumin employing adult Health Stages in Fly Model

Many laboratories have studied the efficacy of K in fly model. They estimate biochemical markers, antioxidant enzymes and phenotype associated rescue on exposure to neurotoxic agents and suggested K for therapeutic approach.

One such study using 8-10 days old fly has reported that upon treatment with K the neurotoxin acrylamide induced levels of reduced GSH, total thiols and acetylcholine esterase activity was also restored and mitochondrial dysfunctions was alleviated and also

restored the decreased DA levels (Prasad and Muralidhara, 2014). Pre-treatment with synthesized K bioconjugate K monoglucoside also protected against rotenone neurotoxicity and exerted antioxidant effects by replenishing cellular GSH levels and considerably decreasing reactive species, restored mitochondrial complex I and IV activities, restored nuclear damage and induced anti-apoptotic effects. Q-PCR (quantitative polymerase chain reaction) analysis of redox genes showed up-regulation of Nitric Oxide Synthase-2 (NOS2) and down-regulation of NAD(P)H Quinone Dehydrogenase 1 (NQO1) when flies were exposed to rotenone which was attenuated by CMG pre-treatment (Paandaresh et al, 2016). Employing newly eclosed adult flies it is also reported that DAergic neuron-specific knockdown of dUCH (a homolog of human UCH-L1, Ubiquitin terminal-hydroxylase-L1) led to locomotor deficit and DAergic neuronal loss and induction of OS. K could improve motor function, decrease the elevated ROS level and minimize the extent of neurodegeneration (Nguyen et al, 2018). Another investigation has shown K treatment improve the motor impairment caused by rotenone toxicity in 7-8 days old fly (Khatri et al, 2016).

However, it is imperative to note that all the above studies were performed only in young flies in the age group of 1-10 days old. Basing on this result it may be lacking to say that neuroprotective efficiency of K is active in all the life stages of adult fly because it is known that there is 23% variation in genome-wide transcript pattern with respect to age in *Drosophila* (Pletcher *et al*, 2002) signifying that targets of genotropic compounds under investigation may be absent at some life stage; they would be active only during those stages where their target molecules are present (Soh *et al*, 2013). Thus, I decided to understand neuroprotective efficacy of K by studying the stress markers and enzyme activities in both the young and old stage groups of flies through the model developed.

8. Materials and Methods

8.1. Fly Stock

Drosophila melanogaster of Oregon K strain was obtained from National *Drosophila* Stock Centre of University of Mysore, Mysore, Karnataka, India. Male flies were used in the present study. They were kept at 22°C-24°C and fed on a standard culture medium made of sucrose, yeast, agar agar, and propionic acid.

8.2. Chemicals

Bovine Serum Albumin (A-2153), Curcumin (C-1386), Paraquat (methyl viologen dichloride hydrate, 856177), 2,7-dichlorofluorescin diacetate (D-6883), Thiobarbituric acid (T5500), Hydrogen peroxide (323381), Acetylthiocholine iodide (A5751), Quercetin (fluka, 200595), N'N'N'N-Tetramethylethylenediamine (T9281), Glutathione (G-4251), were purchased from Sigma Aldrich St. Louis, USA. Dimethyl sulfoxide (DMSO), 2,4-Dinitro phenyl hydrazine (DNPH), 1,1-dithio nitro-bi-benzoicacid (DTNB), 1-Chloro-2,4-dinitrobenzene (CDNB), Ethyldiaminetera acetic acid (EDTA), Dimethlysulfoxide (DMF), Sodiumdodecyl sulphate (SDS), Ortho-pthaldehyde (OPA), Xylenol orange, Acetone, Glacial acetic acid, Trichloroacetic acid and all other analytical grade chemicals were procured from Sisco Research Laboratory Chemicals and Merck, India. Whatman filter paper no.1 disc was used as a feeding medium in the experiment.

8.3. Treatment Protocol

Male flies were aged for 4 days and 55 days feeding in sucrose-agar media. The flies were transferred to freshly prepared media every 3^{rd} days while aging. The flies were then transferred to vials (30mm x100mm) containing single disc of Whatman filter paper no. 1 saturated with 275µl of 5% sucrose, 10mM PQ in 5% sucrose, Curcumin (500µM, 1mM) with PQ (10mM) and Curcumin in DMSO. At 24hrs of exposure, flies were freezed @-

80°C. For dissection of head, an aluminium tray which was positioned on the ice block and flies were placed on it. Using Carl Zeiss stereozoom (Stemi 305) microscope the flies were then dissected separating head from body with a sharp razor.

8.4. Preparation of Homogenate

100 heads were homogenized with pestle motor mixer (Argos technologies) in $175\mu 10.1M$ Phosphate Buffer Saline (PBS) (pH 7.4). It was then centrifuged at 5000rpm for 10mins @ 4°C. The supernatant was again centrifuge under the same conditions. The resulting clear supernatant was stored at -80°C/used for biochemical assay.

8.5. Estimation of Protein

The Protein concentrations of the tissue homogenates were determined by the modified version of the method initially described by Bradford (1976) using Bio-Rad protein assay dye reagent concentrate. The Bradford assay is a colorimetric assay for protein determination based on absorbance shift in the dye Coomassie brilliant blue-G250. Coomassie brilliant blue which is red in unbound form, on binding to protein change to stable blue form with absorbance shift from 465nm to 595nm. Since the increase of absorbance at 595nm is proportional to the amount of bound dye and thus to the amount of protein present in the sample. This can be used as a measure for the protein concentration of the unknown sample. Bovine serum albumin was used as the standard prepared in concentration range of 0.5μ gP to 3.5μ gP. The measurement was performed using NanoDrop 2000 (Thermo Scientific).

9. Biochemical Assays

9.1. Reactive Oxygen Species (ROS)

Generation of ROS was determined using 2',7'-dichlorofluorescein diacetate (DCFDA), a non polar compound that can quickly react with ROS to form the highly fluorescent compound, 2',7'-dichlorofluorescein (DCF) after conversion to a polar derivative by intracellular esterases. DCF can be detected by fluorescence spectroscopy.

50µgP of brain sample was incubated in Locke's buffer (NaCl-154, KCl-5.6, NaHCO₃-3.6, HEPES-5, CaCl₂-2, glucose-10 in mM concentrations, pH 7.4) containing 5µM DCFDA for 45 minutes at room temperature and the fluorescence was measured with excitation 480nm,emission 530nm using Cary Eclipse Fluorescence Spectrophotometer (Agilent Technologies). The ROS generation was expressed as pmol DCF/min/mg protein.

9.2. Lipid Peroxide (LP)

Lipid peroxidation was assessed by measured by estimation of MDA by thiobarbituric acid (TBA) reaction method. MDA a product of lipid peroxidation react with TBA to form 1:2 adduct (MDA-TBA2) measured spectrophotometrically at 532nm.

200µgP of test samples were allowed to react in 1.5 ml of 20% acetic acid (pH 3.5), 1.5 ml of 0.8% TBA, 0.2 ml of 8% SDS (w/v) and vortex, followed by 100°C incubation in water bath for 30 minutes. The mixture was cool down to room temperature and optical density absorbance was measured at 532nm using NanoDrop 2000 (Thermo Scientific) (Ohakawa *et al*, 1979).

9.3. Protein Carbonyls (PC)

This technique involves detecting and quantifying oxidative modification of proteins. 2,4dinitrophenylhyrdazine (DNPH) reacts with protein carbonyls to form a stable dinitrophenyl (DNP) hydrazone adducts proportional to carbonyls present which can be detected spectrophotometrically at an absorbance of 360 nm.

Protein carbonyl (PC) levels in the samples were quantified following reaction of $100\mu gP$ test samples with 2,4-dinitrophenyl hydrazine (DNPH) for 1hr. The protein is precipitated by adding 20% trichloroacetic acid, kept on ice for 5 minutes and it is then centrifuged at 3000rpm for 10 minutes at 4°C. The pellets were washed with 1ml ice cold acetone, followed by dissolving in 20mM Tris-HCl buffer (pH 7.4, containing 2% SDS, 10mM EDTA). The optical density was measured at 360 nm using NanoDrop 2000 (Thermo Scientific) and expressed as nmol carbonyl/mg protein (MEC=22,000/M/cm) (Levine *et al*, 1990).

9.4. Hydroperoxides (HP)

The H_2O_2 assay was based on the oxidation of ferrous ions (Fe²⁺) to ferric ions (Fe³⁺) wherein the ferric ions (Fe³⁺) react with the indicator dye xylenol orange to produce a purple colored complex measurable at 560nm. Sorbitol was included in the assay to amplify the color intensity.

HP levels were measured according to a previously described method using Ferrous ion oxidation xylenol orange (FOX1) reagent (100μ M xylenol orange, 250μ M ammonium ferrous sulphate, 100μ M sorbitol, 25mM H₂SO₄). 100μ gP of test sample was added to 1ml FOX reagent and incubated in dark at room temperature for 30 minutes. The color developed was read at 560nm in a NanoDrop 2000 (Thermo Scientific). The concentration

of HP was calculated using molar extinction coefficient of 2.2 x 105 M^{-1} cm⁻¹ and expressed as nmol HP/mg protein (Wolff, 1994).

9.5. Superoxide Dismutase (SOD)

Superoxide dismutase activity was measured indirectly by monitoring the inhibition of quercetin auto-oxidation. Quercetin is oxidized by O_{2-} produced by TEMED, which is effectively inhibited by SOD in the sample. The rate of inhibition of Quercetin oxidation is monitored at 406nm.

100µgP of test sample was mixed with phosphate buffer (0.016 M, pH 7.8, containing TEMED- 0.8mM and EDTA-0.08 mM) followed by addition of quercetin (1.5mg/10 ml DiMethyl Formamide). The rate of its auto-oxidation was monitored at 406 nm for 1 minute with 10 seconds interval using NanoDrop 2000 (Thermo Scientific). 50% inhibition of quercetin oxidation in the test sample is defined as one unit of the enzyme and activity expressed as units/mg protein (Kostyuk and Potapovich, 1989).

9.6. Catalase (CAT)

Catalase is involved in the detoxification of H_2O_2 , a toxic product of both normal aerobic metabolism and pathogenic ROS production. This enzyme catalyze the conversion of two molecules of H_2O_2 to molecular oxygen and two molecules of water and the rate of decomposition of H_2O_2 is measured at 240nm.

CAT activity was estimated by adding 25μ gP of samples to phosphate buffer (0.1M, pH 7.4, containing 10 mM H₂O₂). H₂O₂ degradation was monitored at 240nm for 2 minutes with 15 seconds interval in NanoDrop 2000 (Thermo Scientific) and expressed as nmol substrate/min/mg protein (MEC=44/mM/cm) (Aebi, 1984).

9.7. Glutathione S-Transferase (GST)

GST represents a family of enzymes that play important role in detoxification of xenobiotics. GST catalyzes the conjugation of the thiol group of the GSH to electrophilic xenobiotics to protect cell against toxicants. It utilizes GSH to scavenge toxic compounds including those produced due to OS and is part of the defense mechanism against the mutagenic, carcinogenic and toxic compounds (Boyland and Booth, 2003). GST activity was estimated by monitoring the conjugation of glutathione to CDNB.

The reaction was initiated by addition of 40μ gP test sample to phosphate buffer (0.1M, pH 6.5) containing 0.5mM EDTA, 0.075mM CDNB, 0.05mM GSH. The increase in optical density was observed for 5 minutes at 340nm using NanoDrop 2000 (Thermo Scientific). The activity was expressed as nmol conjugate formed/min/mg protein using molar extinction coefficient 9.6 mM⁻¹ cm⁻¹ (Guthenberg *et al*, 1985).

9.8. Reduced Glutathione (GSH)

GSH activity was determined based on fluorimetric method using Orthopaldehyde (OPA) following the standard procedure. GSH react with OPA to form a stable highly fluorescent tricyclic derivate at pH 8.0 which is measured spectrophotometrically.

 50μ gP of sample protein was mixed with 5% tricarboxylic acid (TCA) and centrifuge at 3000rpm for 5 minutes at 4°C. The supernatant was added to a reaction mixture of buffered formaldehyde (1:4 (v/v) 37% formalin: 0.1M Na₂HPO₄). 900µl of Sodium phosphate buffer (0.1M, 5mM EDTA, pH 8.0) was added to each tube followed by 100µl OPA. Then after dark incubation for 30 minutes at room temperature, the fluorescence was measured at excitation and emission wavelengths of 345 and 425 nm respectively using Fluorimeter (Agile Life technologies) (Cohn *et al*, 1966).

9.9. Total Thiols

The procedure is based on the reaction of the thiol with 5,5'-dithiobis 2-nitrobenzoic acid (DTNB) to give the mixed disulfide and 2-nitro-5-thiobenzoic acid (TNB) which is quantified by the absorbance of the anion (TNB²⁻) at 412 nm. The reagent has been widely used for the quantitation of thiols in peptides and proteins.

Total thiols activity was determined by taking 25µgP of test sample containing Tris buffer (pH 8.0) reacted by adding 25µl DTNB and 1775µl methanol. The reaction was incubated in dark for 30minutes with occasional mixing, followed by centrifugation at 3000rpm for 5 minutes at 4°C. The supernatant was used to measure the ability of thiols to oxidize DTNB using spectrophotometer at 412 nm in NanoDrop 2000 (Thermo Scientific). Total thiols activity was then expressed as nmol/mg protein (Ellmann, 1959).

9.10. Acetylcholinesterase (AChE)

Acetylcholinesterase efficiently catalyze the hydrolysis of acetyl-thiocholine sulfur analogs of its natural substrate, acetylcholine. Upon hydrolysis, these substrate analogs produce acetate and thiocholine. Thiocholine in the presence of the highly reactive dithiobisnitro-benzoate (DTNB) ion reacts to generate the yellow of 5-thio-2-nitrobenzoate anion. The yellow color product can be measured by its absorbance at 405 nm (Ellman *et al*, 1961).

Acetylcholinesterase (AChE) activity was determined by taking 25µgP test sample containing in phosphate buffer (0.1M, pH 8.0) and DTNB (10mM). To the mixture, 20µl of acetylthiocholine iodide (ATCI, 78 mM) was added. The reaction absorbance was monitored at 412 nm for 5 minutes with 30 seconds interval in a NanoDrop 2000 (Thermo Scientific). AchE activity was then expressed as nmol substrate hydrolyzed/min/mg protein (Ellmann *et al*, 1961).

131

11. Results

11.1. Curcumin diminishes PQ induced ROS level during Health and Transition phase of adult life stage in fly model of PD

The modulatory activity of K on PQ induced elevation in ROS levels was determined. It was found that in 4 day old flies 34% enhanced level of ROS by PQ exposure was effectively decreased by 20% and 60% upon $R_{500\mu M}$ and R_{1mM} K co-treatment. Also, in 55 day old flies there was 5 fold increases in ROS level by PQ treatment when compared to control, suggesting that at later stage of life, the organism becomes more susceptible to OS conditions. K co-treatment could decrease the elevated ROS levels by 60% and 35% in both the concentrations ($R_{500\mu M}$ and R_{1mM}). PQ elevated the ROS levels in both the age group of flies and K *per se* did not have any detrimental effects on 4-day flies. However, in the 55 day old flies it was found that concentration of K *per se* ($K_{500\mu M}$ and K_{1mM}) increased the ROS level when compared to control (Fig.4) suggesting that K itself may act as potential pro-oxidant and therefore the necessity to carefully screen the compound before prescribing them for therapeutic agents.

11.2. Curcumin diminishes PQ induced Lipidperoxide level during Health and Transition phase of adult life stage in fly model of PD

To study the neuroprotective action of K by estimating Lipid Peroxidation, the levels of MDA an end product of LP was analyzed as one of the stress markers. PQ exposure upregulated LP by 60% and K co-treatment attenuated the level by 23% and 31% in $R_{500\mu M}$ and R_{1mM} respectively in health stage. In the transition stage, PQ treatment upregulated LP by 24% when compared to control. The increase in LP level was attenuated by 30% with both the K concentration used ($R_{500\mu M}$ and R_{1mM}) which indicate the effective free radical sequestration properties of K (Fig.5). K *per se* did not show any toxic effect in

transition stage, but in health stage it was evident that the K *per se* exerts toxicity at this stage with the selected concentration.

11.3. Curcumin diminishes PQ induced Protein carbonyl level during Health and Transition phase of adult life stage in fly model of PD

K neuroprotective efficacy was also studied by estimating the PC level. While there was marked increase in PC upon PQ exposure (44% during health stage and 27% increase in transition phase), K co-treatment could effectively attenuate the PC level by all the concentrations used in health stage. There was 18%, 28% and 20% reduction in PC levels with all the tested concentrations of K ($R_{100\mu M}$, $R_{500\mu M}$ and R_{1mM} respectively). At transition stage there was reduction by 25% ($R_{100\mu M}$,), 24% ($R_{500\mu M}$) and 20% (R_{1mM}). At highest concentrations used suggesting K itself may be toxic at this concentration stage and can have detrimental effect which is also evident from K *per se* in higher concentrations used (K_{1mM}) in both stages (Fig.6).

11.4. Curcumin diminishes PQ induced Hydroperoxide level during Health and Transition phase of adult life stage in fly model of PD

Hydroperoxidation was increased by 30% in health stage and 13% in transition stage on PQ exposure. K co-treatment decreased the HP level by 20% and 30% in health stage and 15% and 18% in transition stage with both $R_{500\mu M}$ and R_{1mM} concentrations respectively (Figure.7). There was no toxicity associated with K *per se* treatment. This result further confirms the potent free radical scavenging activity of K. It is evident that K can actively

sequester the OS level in both the health and transition stage of adult life stages of *Drosophila*.

11.5. Curcumin diminishes PQ induced SOD level during Health and Transition phase of adult life stage in fly model of PD

SOD, the first line of antioxidant defense system was elevated by 20% in health stage and 27% in transition stage (Fig.8) by PQ treatment. K co-exposure significantly reduced the antioxidant enzyme activity. It was found that R_{1mM} concentration could bring down the elevated SOD activity by 40% in health stage and 20% in transition stage. It suggests lowered level of OS in the fly brain with K co-treatment. In transition stage, it was found that K *per se* was also found to actively sequester OS as indicated by decreased SOD activity when compared to control.

11.6. Curcumin diminishes PQ induced CAT level during Health and Transition phase of adult life stage in fly model of PD

K effectively decreased the CAT activity which was elevated by PQ. Exposure of PQ increased the CAT level by 43% in health stage and 30% in transition stage (fig.9). Co-treatment of K decreased the level by 44% and 35% in health stage and 40% and 15% in transition stage by both $R_{500\mu M}$ and R_{1mM} respectively. While K *per se* was also found to increase the CAT activity by 30% in health stage, there was no significance in transition stage. The result shows antioxidant activity of K which could reduce the elevated antioxidant enzyme activity suggesting lowered level of OS by K action.

11.7. Curcumin diminishes PQ induced GST level during Health and Transition phase of adult life stage in fly model of PD

The antioxidant property of K was evident even by the measurement of GST activity. It was found that PQ elevated GST level by 20% during health stage and 30% in transition phase when compared to controls (Fig.10). The elevated level was decreased by 18% and 22% in health stage and 15% and 19% in transition stage in both the concentrations used ($R_{500\mu M}$ and R_{1mM}) respectively. This result further reaffirms the antioxidant property of K and its modulatory effect against PQ induced OS in fly model of PD.

11.8. Curcumin replenishes PQ induced Glutathione activities during Health and Transition phase of adult life stage in fly model of PD

PQ exposure led to significant reduction in GSH activities in both the age groups of flies. There was reduction of 30% during health stage and 27% during transition stage (Fig.11) suggesting enhanced levels of neuronal OS. K significantly rescues the diminished level of GSH in both the age groups. It was found that 20% and 15% rescue in health stage and 23% and 27% in transition stage with both R500µM and R1mM concentration respectively. These results further suggest the efficacy of K as an antioxidant therapeutic agent in PD condition.

11.9. Curcumin replenishes PQ induced Total Thiols activity during Health and Transition phase of adult life stage in fly model of PD

K could also rescue the reduced thiols levels caused by PQ treatment. In health stage 18% inhibition due to PQ was rescued by 14% and 12% with $R_{500\mu M}$ and R_{1mM} concentration

whereas in transition stage 27% inhibition in thiols levels was rescued by 17% and 14% in both the $R_{500\mu M}$ and R_{1mM} concentration respectively (Fig.12). It further supports the findings in other markers that K has anti-oxidative property and potential for therapeutic agent in PD.

11.10. Curcumin replenishes PQ inhibited AChE level during Health and Transition phase of adult life stage in fly model of PD

K also rescued the PQ induced marginal inhibition of AChE activity in both the age groups. In health stage 20% inhibition in AChE activity was rescued by 13% and 16% and in transition stage 16% inhibition was rescued by 15% and 8% in both the $R_{500\mu M}$ and R_{1mM} concentrations respectively (Figure.13). This result further suggests the neuromodulatory properties of K and its potential application as therapeutic agent in PD.

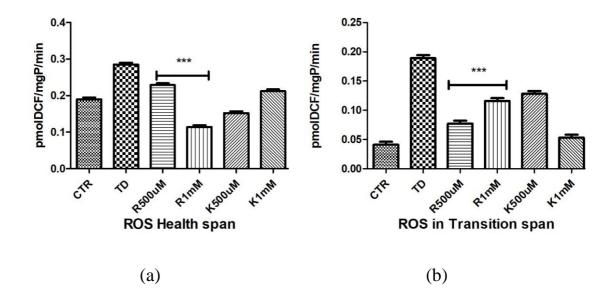


Figure.4: Measurement of ROS levels in (a) Health stage and (b) Transition stage. PQ enhances ROS significantly up to 34% during health stage (4a) and 5 fold increase in transition phase (4b) of adult *Drosophila* model. On co-treatment with K the enhanced levels of ROS were significantly diminished during both the adult life stages of fly. It was found that 34% enhanced level of ROS in PQ treated sample were brought down by 20% and 60% in $R_{500\mu M}$ and R_{1mM} in health stage respectively (fig.4a); and PQ induced 5 fold ROS increase was brought down by 60% and 35% in $R_{500\mu M}$ and R_{1mM} in transition stage respectively (fig.4b) suggesting K acting as an antioxidant in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

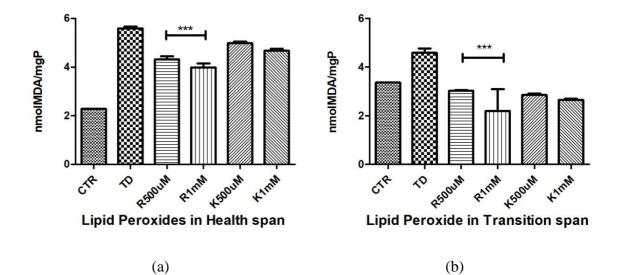


Figure.5: Measurement of MDA levels in (a) Health stage and (b) Transition stage. PQ induces lipid peroxidation as indicated by measurement of Malondialdehyde (MDA) an end product of LP. It was found that there was up to 60% increase in MDA levels during health stage (5a) and 24% increase in transition phase (5b) of adult *Drosophila* model. On co-treatment with K the enhanced levels of MDA were significantly diminished during both the adult life stages of fly. It was found that 60% enhanced level of MDA in PQ treated sample were brought down by 23% and 31% in $R_{500\mu M}$ and R_{1mM} in health stage respectively (fig.5a); and PQ induced 24% PC increase was brought down by 30% in both the $R_{500\mu M}$ and R_{1mM} concentrations in transition stage (fig.5b) suggesting the effective modulatory action of K against PQ induced OS in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

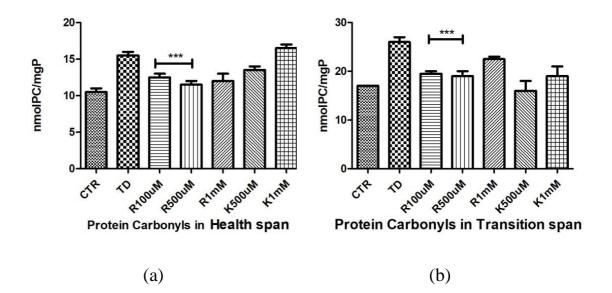


Figure.6: Measure of Protein Carbonyl levels in (a) Health stage and (b) Transition stage. PQ enhances protein carbonylation up to 44% during health stage (6a) and 27% increase in transition phase (6b) of adult *Drosophila* model. On co-treatment with K the enhanced levels of PC were significantly diminished during both the adult life stages of fly. It was found that 44% enhanced level of PC in PQ treated sample were brought down by 18%, 28% and 20% in R_{100µM}, R_{500µM} and R_{1mM} in health stage respectively (fig.6a); and PQ induced 27% PC increase was brought down by 25%, 24% and 20% in R_{100µM}, R_{500µM} and R_{1mM} in transition stage respectively (fig.6b) suggesting the modulatory effect of K against PQ induced OS in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

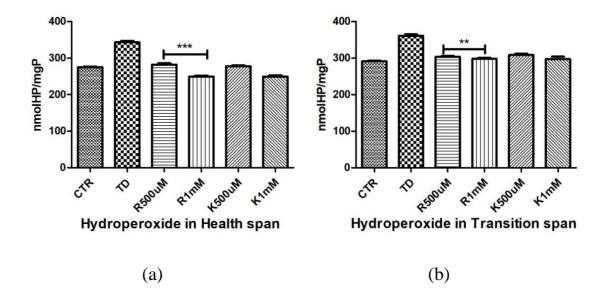


Figure.7: Measurement of HP levels in (a) Health stage and (b) Transition stage. Treatment of PQ enhanced the hyderperoxidation by 20% and 13% during health stage and transition phase (fig.7) of adult *Drosophila* model. On co-treatment with K the enhanced levels of HP were significantly diminished during both the adult life stages of fly. It was found that 20% increased levels of HP in PQ treated sample were brought down by 20% and 30% in $R_{500\mu M}$ and R_{1mM} in health stage respectively (fig.7a); and PQ induced 13% HP increase was brought down by 15% and 18% in both the $R_{500\mu M}$ and R_{1mM} concentrations in transition stage (fig.7b) suggesting that K has modulatory effect against PQ induced OS in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

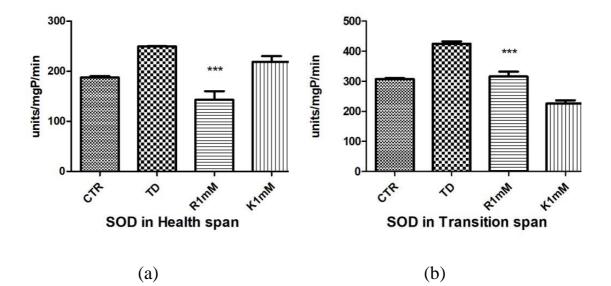


Figure.8: Measurement of SOD activity in (a) Health stage and (b) Transition stage. PQ increases the SOD enzyme activity by 20% and 27% during health stage and transition phase (Fig.8) of adult *Drosophila* model. On co-treatment with K the increased activity of SOD was significantly diminished in both the adult life stages of fly. It was found that the increased levels of SOD in PQ treated sample were brought down by 40% and 20% with R_{1mM} concentration during both the health stage and transition stage respectively (fig.8a,b) suggesting that K has a potent anti-oxidative effect against PQ induced OS in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

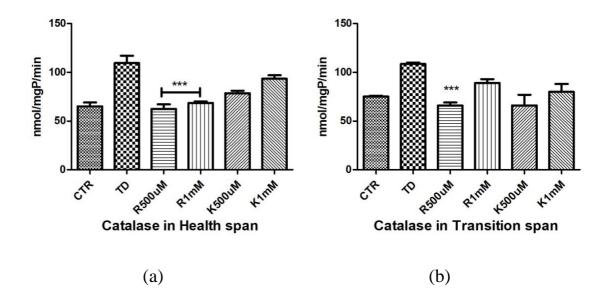


Figure.9: Measurement of Catalase activity in (a) Health stage and (b) Transition stage. PQ increases the CAT enzyme activity by 43% and 30% during health stage and transition phase (fig.9a, b) of adult *Drosophila* model. On co-treatment with K the increased activity of CAT was significantly diminished during both the adult life stages of fly. It was found that 43% increased levels of CAT in PQ treated sample were brought down by 44% and 35% in $R_{500\mu M}$ and R_{1mM} in health stage respectively (fig.9a); and PQ induced 30% CAT activity increase was brought down by 40% and 15% in both the $R_{500\mu M}$ and R_{1mM} concentrations in transition stage (fig.9b) suggesting that K has a potent anti-oxidative effect against PQ induced OS in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

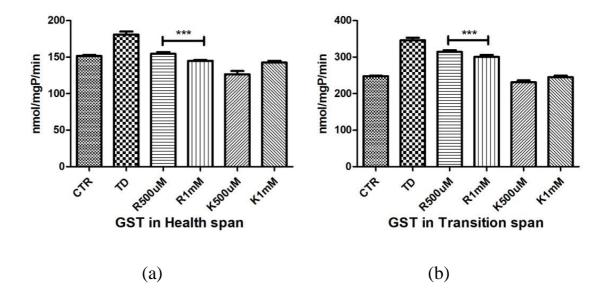


Figure.10: Measurement of GST level in (a) Health stage and (b) Transition stage. PQ enhances the GST enzyme activity by 20% and 30% during health stage and transition phase (Fig.10a,b) of adult *Drosophila* model. On co-treatment with K the activity of GST was significantly decreased in both the adult life stages of fly. It was found that 20% increased levels of GST in PQ treated sample were brought down by 18% and 22% in $R_{500\mu M}$ and R_{1mM} concentrations in health stage respectively (fig.10a); and PQ induced 30% increase GST activity was brought down by 15% and 19% in both the $R_{500\mu M}$ and R_{1mM} concentrations stage respectively (fig.10b) suggesting that K has modulatory effect against PQ induced OS in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

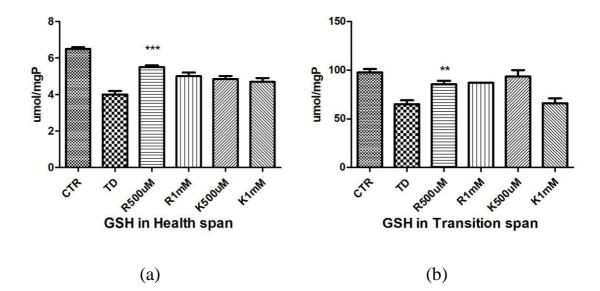


Figure.11: Measurement of GSH levels in (a) Health stage and (b) Transition stage. PQ treatment inhibits the GSH level by 30% during health stage and 27% during transition phase (Fig.11a, b) in the adult *Drosophila* model. On co-treatment with K the activity of GSH were significantly increased in both the adult life stages of fly. It was found that 30% GSH activity inhibition upon PQ exposure were rescued by 20% and 15% in $R_{500\mu M}$ and $R1_{mM}$ concentrations health stage respectively (fig.11a); and 27% GSH activity inhibition was rescued by 23% and 27% in both the $R_{500\mu M}$ and R_{1mM} concentrations stage respectively (fig.11b) suggesting the efficacy of K as a modulant against the PQ induced OS in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

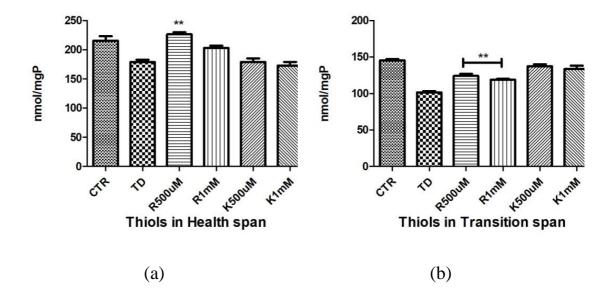


Figure.12: Measurement of total Thiols level in (a) Health stage and (b) Transition stage. PQ treatment also inhibits thiols level by 18% during health stage and 27% during transition phase (Fig.12a, b) in the adult *Drosophila* model. On co-treatment with K the activity of thiols was significantly increased in both the adult life stages of fly. It was found that K co-exposure could rescue the inhibited thiols activity by 14% and 12% in $R_{500\mu M}$ and R_{1mM} concentrations health stage respectively (fig.12a); and rescued by 17% and 14% in both the $R_{500\mu M}$ and R_{1mM} concentrations in transition stage respectively (fig.12b) further suggesting the efficacy of K as a modulant against the PQ induced OS in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

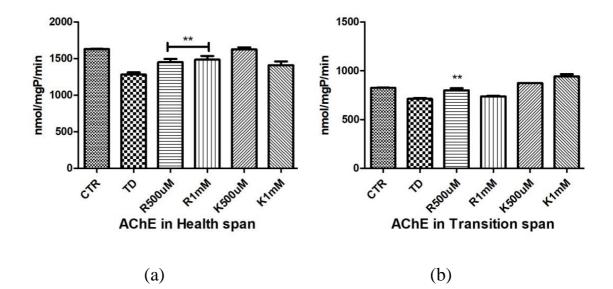


Figure.13: Measurement of AChE activity in (a) Health stage and (b) Transition stage. PQ treatment inhibits AChE level by 20% during health stage and 16% during transition phase (Fig.13a, b) in the adult *Drosophila* model. On co-treatment with K the AChE level was significantly increased in both the adult life stages of fly. It was found that K co-exposure could rescue the inhibited AChE activity by 13% and 16% in $R_{500\mu M}$ and R_{1mM} concentrations in health stage respectively (fig.13a); and rescued by 15% and 8% in both the $R_{500\mu M}$ and R_{1mM} concentrations in transition stage respectively (fig.13b) suggesting the efficacy of K as a modulant against the impairment of neurotransmitter enzyme activity induced by PQ in fly model of PD. (CTR=Control, TD=Treated with PQ, R (Rescue) =PQ+K, K= Curcumin *per se*).

12. Discussion

Evidence suggesting the involvement of OS mechanism arises from both in vitro and in vivo models relevant to PD. It is considered to be the common underlying mechanism that leads to cellular dysfunction and demise (Kurtishi *et al*, 2018). Studies on post-mortem brains from PD patients have implicated the role of oxidative damage in the pathogenesis of PD (Zeevalk *et al*, 2008; Nakabeppu *et al*, 2007; Bosco *et al*, 2006). This illustrates the role of OS in DA degeneration is linked to PD.

12.1. Susceptibility to Oxidative Stress with Age

Protein, lipids and nucleic acids are biological macromolecules which are oxidatively altered due to ROS, resulting in genetic mutations and cellular senescence (Luceri *et al*, 2018). In complications and pathologies associated with aging, increasing evidence for causal role of cell senescence have been experimentally shown (Correia-Melo *et al*, 2015). Due to stress during aging there is increase in cell senescent. This phenomenon is controlled by a number of counting mechanisms including telomeres shortening, gradual increase in damage of DNA, deviation in normal oncogenes behavior, change in metabolic action and increased ROS generation (Kuilman *et al*, 2010). Such processes lead to impairment of cellular function and thus produce and accumulate ROS which in turn disturb the homeostasis, a vital factor for safeguarding the senescent phenotype. Apart from injuring DNA and damage mitochondrial activity, OS trigger the activity of p53, a gene coding for cell cycle regulator protein, which in turn induces prooxidant genes. Disturbances in redox homeostasis by generating ROS lead to deterioration of cell and impair its function and as a response to increased ROS levels, mechanisms for cellular protection are excessively activated including up-regulated expression of antioxidant

enzymes (Ren and Zhang, 2017; Zarse *et al*, 2012) suggesting OS is intricately linked to aging and neurodegeneration.

12.2. Insights from Quantification of Non Enzymatic Markers

In the present study in *Drosophila* model of sporadic PD condition the levels of non enzymatic OS markers such as ROS, PC, MDA, HP were increased significantly during both the phases (health and transition) of adult life. Sequestration of OS is evident from the significant decreased levels of these markers (fig.4-7).

While there was about 35% increase in ROS level in health stage of PQ treated flies when compared to control, in transition stage the level was increased by 5 fold suggesting stress vulnerability of the fly with age.

Increased in ROS levels and MDA an intermediate in the lipid peroxidation process upon PQ exposure has also been shown by other laboratories (Soares *et al*, 2017). Lipid peroxidation is a well-known example of oxidative damage in cell membranes, lipoproteins, and other lipid-containing structures. Study on postmortem PD brain have shown Lipid peroxidation was increased in Parkinsonian nigra compared with other brain regions and control tissue (Dexter *et al*, 1994). MDA levels are also found to be significantly higher in PD patients than in controls suggesting that high plasma lipid peroxidation rates might contribute as a risk factor for PD (Sanyal *et al*, 2009). PQ induced increase in the MDA level was effectively reduced with K co-treatment in both the age groups (Fig.5), as also shown by other reports of K ability to attenuate elevated MDA levels (Wang *et al*, 2017).

There was significant protein damage in flies as evident by the high level of PC which indicate the increased rate of protein oxidation (fig.6). Protein carbonylation is an irreversible oxidative damage which leads to loss of protein function and it is considered

148

as a widespread indicator of severe oxidative damage and disease-derived protein dysfunction. Elevated levels of PC associated with PD have been reported in postmortem PD brains tissues (Alam *et al*, 1997). The efficacy of K to decrease the PC levels has been reported (Kolodziejczyk *et al*, 2011; Dekhar *et al*, 2010).

HP groups are powerful oxidants that can induce secondary damage through inactivation of thiol-dependent enzymes essential to cell functions. Study on α -synuclein implicated in PD catalyzing the formation of hydrogen peroxide has been reported (Turnbull *et al*, 2001). Protein hydroperoxides results in modification of the structure and properties of amino acid residues. Lipid hydroperoxides are also prominent non-radical intermediates of lipid peroxidation that can often provide important mechanistic information. Increased levels of lipid hydroperoxides in Parkinsonian SN has been demonstrated (Dexter *et al*, 1994).

12.3. Estimation of Antioxidant Enzymes Activity

In *Drosophila*, SOD/CAT system is one of the intracellular protective mechanisms against ROS induced damage. Studies have demonstrated that in response to the OS generated by PQ, levels of SOD, CAT and GST are upregulated. K treatment results in the significant reduction of these antioxidant enzymes which suggest lowered level of OS in the brain of K co-treated flies. Post mortem studies have shown an increased SOD activity in PD brains (de Farias *et al*, 2016) where SOD activity was significantly associated with late PD stage. The ability of CAT coated nanoparticles to reduce protein oxidation, DNA damage, mitochondrial, restored neuronal morphology has also been shown (Singhal *et al*, 2013). Investigation of two synthetic superoxide dismutase/catalase mimetics protecting against PQ-induced DAergic cell death in both the rat DAergic cell line and primary mesencephalic cultures in vitro and in adult mice in vivo has also been reported (Peng *et*

al, 2005). There was significant increase in the activity of antioxidant enzymes such as SOD (fig.8), Catalase (fig.9) and GST (fig.10). These findings substantiate well with the increased in level of ROS and other stress marker measured. K therapeutic treatment could attenuate the altered activities of these biomarkers measured in both the age groups.

The GST activity was also elevated under PD condition and K treatment markedly decreased its activity in the present study (fig.10). Antioxidant role of GST in protection against oxidant toxicity and regulation of stress-mediated apoptosis has been carefully reviewed (Sharma *et al*, 2004). GSTs are a functionally diverse family of soluble enzymes of detoxification that use reduced GSH in conjugation and reduction reactions. Depletion in GSH content and an increase in GST activity have been reported in the brains of human PD patients and also in other experimental models of PD (Johnson *et al*, 2012; Zhu *et al*, 2007).

Reduced GSH level was decreased under PD conditions further supporting the evidence of OS generation (fig.11). Earlier investigation have also suggested the ROS to be the major factor contributing to DAergic neuronal loss in the PD brain, as a consequence of DA metabolism, low GSH, and high levels of iron and calcium in the SN*pc* (Jenner and Olanow, 2006). GSH confers the neuronal cells with multiple defenses not only to sequester but also against different types of toxic products and by products of cellular mechanism. Reduction in GSH levels as compared to control subjects in SN in PD patients have been reported (Sian *et al*, 1994). Depletion in GSH level may impair mitochondrial function via inactivation of complex I and increase in ROS production and the relation between decreased GSH function and severity of PD has been reported (Smeyne and Smeyne, 2013). In the present study K therapeutic intervention markedly rescued the GSH level in both the age groups (fig.11a,b). Efficacy of K to elevate the reduced GSH activity

150

against Acrylamide induced toxicity has also been reported (Prasad and Muralidhara, 2014).

Measurement of total Thiols showed reduced activity under PD condition and K could marginally increase the total thiol levels (fig.12). Thiol groups are highly reactive and susceptible to oxidation that may cause significant loss of biological activity. A critical function of cell-based thiol redox buffering systems is to protect thiol groups from oxidation and to repair those that may have become oxidized as a result of normal or aberrant cellular metabolism. Studies have shown that thiols levels are reduced under disease conditions (Isik *et al*, 2017).

12.4. Acteylcholine esterase as Neurotoxicity Marker

Neurotransmitter enzyme AChE activity was also estimated in the present study. It was found that the enzyme activity was inhibited under PD condition in both the age groups (fig.13a,b). Such action will impair the neurotransmission process since AChE is involved in maintaining the free Acetylcholine in the synaptic region. Earlier studies have shown reduced AChE activity in several brain disorder including neurodegenerative diseases (Mendez *et al*, 2011; Shinotoh *et al*, 2003). The alteration in activities of these enzymes is indirectly responsible for neuronal loss and probably plays a role in pathogenesis of PD. AChE inhibition has been used as a biomarker for the influence of organophosphate and carbamate pesticides in invertebrate and vertebrate model. PQ exposure lead to marginal inhibition of AChE activity and inhibited levels were rescued by K co-treatment in both the age groups (fig.13) reaffirming the efficacy of K to attenuate the PQ induced toxicity in both the age groups. It has been proposed that exposure to low levels of pesticides leads to several neurological and neurobehavioral changes that cannot be accounted for AChE

inhibition only (Salvi *et al*, 2003; Stephens *et al*, 1995). The particular enzyme associated with the different sub-cellular fractions can exert different effects.

The present results show that K can sequester the enhanced levels of OS not only during early phase (health stage) but importantly, also during late phase (transition stage) of adult life stage in fly PD model as seen from the above results (fig.4-13). Further it has been observed that at higher concentration treatment, K *per se* can act as potential pro-oxidant, suggesting the necessity to properly screen the compounds before prescribing them as a therapeutic agent.

13. Oxidative Stress markers-based studies alone may not be sufficient to decipher the Neuroprotective efficacy of Nutraceuticals

Most of the laboratories analyze the neuroprotective action of nutraceuticals using OS markers as a gold standard in their studies considering the OS as a factor responsible for neurodegenerative diseases such as PD (Niveditha *et al*, 2017; Ravi *et al*, 2017; Jhonsa *et al*, 2016). Employing the same approach, I found that K can mitigate the OS induced by PQ exposure in health stage flies as evident from the above results. This finding confirms with several other studies of K neuroprotection employing health stage in adult *Drosophila* PD model (Nguyen *et al*, 2018). Also K could reduce the OS even in the transition stage flies as seen from the above study on stress markers, antioxidant and neurotransmitter enzyme activities which is an important and interesting phenomenon. It is shown that dietary feeding of K and other phytochemicals modulate PQ induced alteration in the expression levels of several genes that are associated with anti-oxidant and anti-aging effect in 2-3 days old flies (Park *et al*, 2012). K also is reported to improve PQ induce motor deficit and replenish the decreased DA level in adult health stage (Abbaiou *et al*, 2017; Phom *et al*, 2014). However, K is unable to rescue either the motor deficits or

diminished DA level in adult transition stage of Drosophila exposed to PQ (Phom et al, 2014). But contrary to this finding, K could reduce the OS levels in both the fly age groups as seen from the above results, clearly suggesting that sequestration of boosted OS alone is not enough to prevent the DAergic neurodegeneration in PD. Therefore apart from OS, there is the synergic effect of other pathways that could be responsible for the DAergic neurodegeneration in PD. Several laboratories have shown the effective role of various therapeutic compounds that modulates the altered enzymatic functions arising out of OS They suggested for therapeutic approach using such active in model organisms. compounds in NDD such as PD. However, according to the present findings it is clear that accessing OS level alone is not sufficient enough to understand the neuroprotective efficacy of therapeutic molecule in late onset NDD such as PD. Studies in addition have also shown that K increases longevity when administered in the developmental of health stage but exerts negative result when administered in other life stage, suggesting that target molecules of K are present in the health stage (Soh et al, 2013) but not in transition and senescence stage.

14. Conclusion

Present study reveals that OS may be a necessary factor, but alone it may not be sufficient enough for DA degeneration which is characteristic pathological feature of PD. Therefore, studies in animal models through which neuroprotective efficacy of multiple natural products being assessed based on OS markers alone may not be good enough to recommend them as neuroprotective agents. Hence, further it is necessary to look into the molecular targets along with the oxidative stress markers. SUMMARY

Parkinson's disease is the second most common neurodegenerative disease and it is characterized by degeneration of DAergic neurons in the SN of mid brain that lead to impairments of motor functions (Cacabelos *et al*, 2017). Progressive loss of DAergic neurons and intraneuronal protein inclusions, called LB in the mid brain are characteristic of PD (Olanow and Tatton, 1999). The neuronal loss in this region gives rise to motor symptoms like bradykinesia, rest tremor, postural instability, and gait impairment whereas non-motor symptoms like impaired olfaction, constipation, depression, increased daytime sleep, rapid eye movement sleep disorder, and behavioral deficits are commonly observed (Saleem *et al*, 2013). During the time of diagnosis of the symptoms, about 50 to 60% of the DAergic neurons have already been degenerated. Over the time, individuals suffering from PD are unable to perform even the basic function and they become completely dependent on care and support.

Although the disease was first described in the 18th century, there is no therapy available that will cure the disease and medication is limited only to alleviate the progressing symptoms and with more of side effects. Presently, the treatment strategies for PD include Deep Brain Stimulation (DBS) and Levo-dopa (L-dopa) therapy. These therapies are considered as the most effective, but placement of lead in DBS often result in infection and intracranial hemorrhage (Zrinzo *et al*, 2012) and long-term treatment of L-dopa leads to abnormal involuntary movement known as L-dopa induced dyskinesia (Fahn *et al*, 2000). Developing a therapeutic strategy for NDD such as PD remains a challenge till date.

Here lies the importance of model organism based development of therapeutic strategies and their further validation in humans. Over the year's researchers have developed several animal models including *Drosophila* while attempting to understand the PD progression and thereby find means to its therapeutic approach. A suitable model for sporadic PD should show histopathologically characterizable progressive loss of DA neurons together with other neurons and significant reduction in DA level. Moreover, the onset of the disease should be in a stage of adulthood. The model animal should also manifest disease in such a way that it would mimic the PD affected human motor symptom such as bradykinesia, rigidity, postural instability and resting tremor, with motor features being responsive to L-DOPA (L-dopa, the precursor of DA) or any anti PD drug therapy. Low cost of maintenance, shorter life cycle and defined neuropathological profile is making *Drosophila* amongst the emerging and more interesting model of PD, though nonhuman primate and mouse has been the traditional model of PD (Pienaar, 2010).

Environmental toxins are implicated to be the causative agents of idiopathic PD. Several case studies have reported that the subjects having exposure to pesticides, herbicides showed symptoms similar to Parkinsonism. Laboratory exposure of model organisms to environmental toxins like PQ is therefore productively employed to study the disease progressions that would help in understanding the mechanisms involved. Animal models such as *Drosophila*, allow researchers to replicate human diseases symptoms or abnormal behavior, enabling to understand the many biological functions without ethical concerns inherent of human studies. Complex human behavior such as aggression, mating, circadian rhythms, sleep, learning and memory are also observed in animals. Therefore, by studying these processes and behaviors in animal models, one can understand the basic biology underlying them and thus applying this information to figure out how diseases progression take place and find the remedial measures.

155

Several researchers have suggested the neuroprotective efficacy of natural products showing modulation of biochemical markers, antioxidant enzymes and phenotype associated rescue upon exposure to neurotoxic agents in different disease models. Natural product K is largely used in food as spices, coloring agent, and traditional medicines in India, China, Southeast Asia (Aggarwal *et al*, 2007) and properties of K performing neuroprotective effect, anti-oxidant, anti-inflammatory and anti-cancer are well known. The most central biological role of K associated to neuroprotection has been shown to be its anti-oxidant effect which protects the SN neurons and increases striatal DA count and chelates Fe^{2+} in the 6-OHDA (6-hydroxyDA) rat models of PD (Mythri *et al*, 2012).

Using *Drosophila*, it is reported that K have gender and genotype specific life span extension and sequester OS mediated free radicals, enhance locomotor ability and show chemo preventive property, suggesting its potential use in treatment applicability in higher organisms (Lee *et al*, 2010). K decreases death of SH-SY5Y human neuroblastoma cells induced by rotenone; improve characteristic symptoms associated with PD in *Drosophila* via reduction in intracellular and mitochondrial ROS levels and acting against the caspase-3/caspase-9 activity (Liu *et al*, 2013). Transgenic fly expressing human α -synuclein was exposed to different concentrations of K and found considerable delay in the loss of activity pattern, decrease in the level of OS and apoptosis, and extended life span (Siddique *et al*, 2014) suggesting potential role of K in neuroprotection.

For understanding the pathology of late onset neurodegenerative diseases like PD, it is important to employ organisms at the adult phases where the disease sets in, such as the transition stage of *Drosophila* PD model. Because different stages of the life stage have different patterns of gene expression and the cellular, physiological and molecular phenomenon at old stage may be completely different from young stage. There exists significant change of about 23% in genome-wide transcript profiles with age in *Drosophila*

(Pletcher *et al*, 2002) suggesting that targets of genotropic compounds under study may well not be present in all life stages. Genotropic drugs would be effective only during those life cycle stages when their target molecules are available (Soh *et al*, 2013). Therefore, it is possible that targets of genotropic compounds such as K may not be present in all life stages, which is an interesting paradigm. However, no reports are available regarding the efficacy of K in PD models during later phases of adult life. Therefore, it is necessary to understand the neuroprotective efficacy of compounds at transition phase because this is the period during which late-onset neurodegenerative diseases such as idiopathic PD sets in.

Oxidative modifications of enzymes and structural proteins play a significant role in the etiology and progression of several human neurodegenerative diseases. Accumulation of free radicals and subsequent neurodegeneration in specific brain regions have been proposed as the underlying factors in neurodegenerative diseases such as Alzheimer's and PD (Halliwell, 2006). Studies on post-mortem brains from PD patients have implicated the role of oxidative damage in the pathogenesis of PD (Yuan *et al*, 2016; Zeevalk *et al*, 2008; Bosco *et al*, 2006).

K rescues the mobility defects induced by PQ during health stages of *Drosophila* but fails to rescue the mobility defects induced by PQ during transition phase of *Drosophila*. This finding is well substantiated by results that K could replenish decreased DA levels caused by PQ exposure in *Drosophila* during health stage but not in transition phase (Phom *et al*, 2014). But K effectively sequester the elevated level of OS and attenuated levels of antioxidant enzyme activities and neurotransmitter enzyme in PD condition during both health span and transition phases of adult life span *Drosophila* suggesting that for a therapeutic compound such as K, the OS pathway may be one of the factors through which

it act to attenuate the disease aggravated abnormal cellular and neural pathways, including one responsible for DAergic neuronal degeneration.

The study reveals that OS may be a necessary factor, but alone it may not be sufficient enough for DA degeneration which is characteristic pathological feature of PD. Therefore, studies in animal models through which neuroprotective efficacy of multiple natural products are being assessed based on OS markers alone may not be good enough to recommend them as neuroprotective agents. Hence, further it is necessary to look into the molecular targets along with the oxidative stress markers. REFERENCES

Abbaoui A, Chatoui H, El Hiba O, *et al* (2017). Neuroprotective effect of curcumin-I in copper induced dopaminergic neurotoxicity in rats: A possible link with Parkinson's disease. *Neurosci Lett.* 660; 103-108.

Abbas M, Bhatti IA, Shahidadeel (2010). Influence of gamma radiation on the colour strength and fastness properties of fabric using turmeric (*Curcuma longa L.*) as natural dye. *Radiat Phys Chem.*79(5); 622-625.

Abou-Sleiman PM, Healy DG, Quinn N, *et al* (2003). The role of pathogenic DJ-1 mutations in Parkinson's disease. *Ann Neurol.* 54;283–286.

Acharya MM, Hattiangady B, Shetty AK (2008). Progress in neuroprotective startegies for preventing epilepsy. *Prog Neurobiol.* 84;363–404.

Aebi H (1984). Catalase in vitro. Meth Enzymol. 105; 121-125.

Aggarwal BB, Sundaram C, Malani N, et al (2007). Curcumin: the Indian solid gold. Adv Exp Med Biol. 595; 1-75.

Alam MA, Ali NA, Sultana N, *et al* (2008). Newborn umbilical cord and skin care in Sylhet District, Bangladesh: Implications for the promotion of umbilical cord cleansing with topical chlorhexidine. *J Perinatol*.28;61–68.

Alhusaini A, Fadda L, Hassan I, et al (2018). Liposomal Curcumin Attenuates the Incidence of Oxidative Stress, Inflammation, and DNA Damage Induced by Copper Sulfate in Rat Liver. *Dose Response*. 16(3); 1559325818790869.

Anandhan A, Tamilselvam K, Radhiga T, *et al* (2012). Theaflavin, a black tea polyphenol, protects nigral dopaminergic neurons against chronic MPTP/probenecid induced Parkinson's disease. *Brain Res.* 1433; 104-13.

Appel-Cresswell S, Vilarino-Guell C, Encarnacion M, *et al* (2013). Alpha synuclein p.H50Q, a novel pathogenic mutation for Parkinson's disease. *Mov Diord*. 28(6); 811-813.

Appiah-Opong R, Commandeur JN, van Vugt-Lussenburg B, *et al* (2007). Inhibition of human recombinant cytochromeP450s by curcumin and curcumin decomposition products. *Toxicology*. 235(1-2); 83-91.

Aquilano K, Baldelli S, Ciriolo M (2014). Glutathione: new roles in redox signaling for an old antioxidant. *Front Pharmacol.* 5(196); 1-12.

Ardley HC, Scott GB, Rose SA, *et al* (2004). UCH-L1 aggresome formation in response to proteasome impairment indicates a role in inclusion formation in Parkinson's disease. *J Neurochem.* 90;379-391.

Arking R (2005). Multiple longevity phenotypes and the transition from health to senescence. *Ann N Y Acad Sci.* 1057;16-27.

Arking R, Novoseltseva J, Hwangho DS, *et al* (2002). Different age specific demographic profiles aregenerated in the same normal-lived Drosophila strain bydifferent longevity stimuli. *J Gerontol A Biol Sci Med Sci*.57;390–398.

Ataie A, Sabetkasaei M, Haghparast A, *et al* (2010). Curcumin exerts neuroprotective effects against homocysteine intracerebroventricular injection-induced cognitive impairment and oxidative stress in rat brain. *J Med Food*. 13(4); 821-826.

Ayajuddin M, Das A, Phom L, et al (2018). Parkinson's disease: Insights from *Drosophila* model. In: *Drosophila melanogaster*: Model for Recent Advances in Genetics and Therapeutics. Ed. Farzana Khan Perveen. *Intech Open.* 8;157-92. *European Union: In-tech Croatia.* 156-192.

Babin PJ, Goizet C, Raldua D (2014). Zebrafish models of human motor neuron diseases: advantages and limitations. *Prog Neurobiol.* 118; 36-85.

Bachli G (1998). Family Drosophilae. In: Papp L., Darvas B., editors. Contributions to a manual of palearctic Diptera. III. Higher Brachteera. *Science Herald*.

Bai Q, Mulett SJ, Garver JA, *et al* (2006). Zebrafish DJ-1 is evolutionarily conserved and expressed in dopaminergic neurons. *Brain Res.* 1113(1); 33-44.

Barrientos A, Moraes CT (1999). Titrating the effects of mitochondrial complex, I impairment in the cell physiology. *J Biol Chem.* 274(23); 16188-16197.

Bastías-Candia S, Zolezzi JM, Inestrosa NC (2018). Revisiting the Paraquat-Induced Sporadic Parkinson's Disease-Like Model. *Mol Neurobiol*. doi- https://doi.org/10.1007/s12035-018-1148-z.

Bateman RJ, Xiong C, Benzinger LST, et al (2012). Clinical and biomarker changes in dominantly inherited Alzheimer's disease. N Engl J Med. 367; 795-804.

Belguendouz L, Fremont L, Linard A (1997). Resveratrol inhibits metal ion-dependent and independent peroxidation of porcine low-density lipoproteins. *Biochem Pharmacol.* 53;1347-1355.

Berry JA, Cervantes-Sandoval I, Nicholas EP, *et al* (2012). Dopamine is required for learning and forgetting in Drosophila. *Neuron*. 74;530–542.

Bilen J, Bonini NM (2005). Drosophila as a model for human neurodegenerative disease. Annu Rev Genet.39; 153-71.

Bill BR, Petzold AM, Clark KJ, et al (2009). A primer for morpholino use in zebrafish. Zebrafish. 6(1); 69-77.

Biswas SK, McClure D, Jimenez LA, *et al* (2005). Curcumin induces glutathione biosynthesis and inhibits NF-kappaB activation and interleukin-8 release in alveolar epithelial cells: mechanism of free radical scavenging activity. *Antioxid Redox Signal*.7(1-2);32-41.

Bjorklund A, Dunnett SB, Stenevi U, *et al* (1980). Reinnervation of the denervated striatum by substantia nigra transplants: functional consequences as revealed by pharmacological and sensorimotor testing. *Brain Res.*199;307–333.

Blenau W, Baumann A (2001). Molecular and pharmacological properties of insect biogenic amine receptors: lessons from *Drosophila melanogaster* and *Apis mellifera*. *Arch Insect Biochem Physiol*. 48(1);13-38.

Blesa J, Trigo-Damas I, Quiroga-Varela A, et al (2015). Oxidative stress and Parkinson's Disease. Front Neuroanat. 8(9);91.

Bogaerts V, Nuytemans K, Reumers J, *et al* (2008). Genetic variability in the mitochondrial serine protease HTRA2 contributes to risk for Parkinson disease. *Hum Mutat.* 29;832-840.

Bonifati V, Rizzu P, Van Baren MJ, et al (2003). Mutations in the DJ-1 Gene associated with autosomal recessive early-onset parkinsonism. Science. 299; 256–259.

Bonini NM, Fortini ME (2003). Human neurodegenerative diseases modelling using *Drosophila*. Annu Rev Neurosci. 26; 627-656.

Bosco DA, Fowler DM, Zhang Q, *et al* (2006). Elevated levels of oxidized cholesterol metabolites in lewy bodies accelerate alpha synuclein fibrilization. *Nat Chem Biol.* 2(5); 249-53.

Bossy-Wetzel E, Schwarzenbacher R, Lipton SA (2004). Molecular pathways to neurodegeneration. *Nat Med*. 10(S2–S9).

Botella JA, Ulschmid JK, Gruenewald C, *et al* (2004). The Drosophila carbonyl reductase sniffer prevents oxidative stress-induced neurodegeneration. *Curr Biol.* 14(9):782-6.

Botella JA, Bayersdorfer F, Gmeiner F, et al (2009). Modelling Parkinson's disease in Drosophila. Neuromolecular Med. 11; 268–280.

Boyland E, Booth J (2003). The Metabolic Fate and Excretion of Drugs. Ann Rev Pharm. 2; 129-142.

Braak H, Tredicia KD, Rüba U, et al (2000). Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol Aging*. 24(2);197-211.

Bradford MM (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem*.72; 248-54.

Bretaud S, Allen C, Ingham PW, *et al* (2006). p53-dependent neuronal cell death in a DJ-1 deficient zebrafish model of Parkinson's disease. *J Neurochem*. 00(6); 1626-1635.

Brettschneider J, Del Tredici K, Toledo JB, *et al* (2013). Stages of pTDP-43 pathology in amyotrophic lateral sclerosis. *Ann Neurol*. 74;20-38.

Brichta L, Greengard P, Flajolet M (2013). Advances in the pharmacological treatment of Parkinson's disease: targeting neurotransmitter systems. *Trends Neurosci.* 36(9); 543-554.

Brieger K, Schiavone S, Miller FJ Jr, et al (2012). Reactive oxygen species: from health to disease. Swiss Med Wkly. 142; w13659.

Brooks AI, Chadwick CA, Gelbard HA, *et al* (1999). Paraquat-elicited neurobehavioral syndrome caused by dopaminergic neuron loss. *Brain Res.* 823;1–10.

Buchhave P, Minthon L, Zetterberg H, *et al* (2012). Cerebrospinal fluid levels of beta-amyloid 1-42, but not of tau, are fully changed already 5 to 10 years beforethe onset of Alzheimer dementia. *Arch. Gen. Psychiatry.* 69;98–106.

Bundy R, Walker AF, Middleton RW, *et al* (2004). Turmeric extract may improve irritable bowel syndromesymptomology in otherwise healthy adults: A pilot study. *J Altern Complement Med*. 10;1015–1018.

Busche MA, Konnerth A (2016). Impairments of neural circuit function in Alzheimer's disease. *Philos Trans R Soc Lond B Biol Sci.* 371(1700); 20150429.

Butler MS (2004). The role of natural product chemistry in drug discovery. J Nat Prod.67;2141–2753.

Cacabelos R (2017). Parkinson's Disease: From Pathogenesis to Pharmacogenomics. Int J Mol Sci. 18(3).

Calabrese B, Halpain S (2005). Essential role for the PKC target MARCKS in maintaining dendritic spine morphology. *Neuron*.48(1); 77-90.

Cao S, Gelwix CC, Caldwell KA, et al (2005). Torsin-mediated protection from cellular stress in the dopaminergic neurons of *Caenorhabditis elegans*. J Neurosci. 25(15); 3801-3812.

Cao J, Jia L, Zhou HM, *et al* (2006). Mitochondrial and Nuclear DNA Damage Induced by Curcumin in Human Hepatoma G2 Cells. *Toxicol Sci.* 91(2); 476–483.

Carlsson A (2001). A paradigm shift in brain research. Science. 294;021-1024.

Carlsson A, Lindqvist M, Magnusson T (1957). 3,4-Dihydroxyphenylalanine and 5-hydroxytryptophan as reserpine antagonists. *Nature*. 180(4596); 1200.

Carroll RE, Benya RV, Turgeon DK, et al (2011). Phase IIa clinical trial of curcumin for the prevention of colorectal neoplasia. Cancer Prev Res (Phila). 4(3); 354-364.

Casjens S, Eckert A, Woitalla D, *et al* (2013). Diagnostic value of the impairment of olfaction in Parkinson's disease. *PloS One*. 8(5); e64735.

Cassar M, Issa AR, Riemensperger T, et al (2015). A dopamine receptor contributes to paraquat-induced neurotoxicity in Drosophila. Hum Mol Genet. 24(1); 197-212.

Cavaco S, Goncalves A, Mendes A, *et al* (2015). Abnormal olfaction in Parkinson's disease is related to faster disease progression. *Behav Neurol*. 2015; 976589.

Chang KH, Chen CM, Chen YC, *et al* (2013). Association between PARK16 and Parkinson's disease in the Han Chinese population: a meta-analysis. *Neurobiol Aging*. 34(10); 2442.e5-9.

Charcot JM (1872). De la paralysieagitante. In Oeuvres Comple`tes (t 1) Lec_.ons sur les maladies du syste`menerveux, pp. 155–188. A Delahaye, Paris. [In English: Charcot J-M. 1877. On Parkinson's disease. In Lectures on diseases of the nervous system delivered at the Salpe^trie`re (transl. Sigerson G). New Sydenham Society, London. 129–156.

Chartier-Harlin MC, Dachsel J, Hulihan M, et al (2009). EIF4G1 mutations in familial parkinsonism. Parkinsonism RelatDisord. 15; 145-146.

Chaudhuri A, Bowling K, Funderburk C, *et al* (2007). Interaction of genetic and environmental factors in a *Drosophila* parkinsonism model. *J Neurosci.* 27(10);2457-2467.

Chauvin C, De Oliveira F, Ronot X, *et al* (2001). Rotenone inhibits the mitochondrial permeability transition-induced cell death in U937 and KB cells. *J Biol Chem.* 276(44); 41394-41398.

Chavko M, Auker CR, McCarron RM (2003). Relationship between protein nitration and oxidation and development of hyperoxic seizures. *Nitric Oxide*. 9(1); 18-23.

Chesselet MF (2008). In vivo alpha-synuclein overexpression in rodents: a useful model of Parkinson's disease? *Exp Neurol*. 209(1); 22-27.

Chiosis G, Tao H (2006). Purine-scaffold Hsp90 inhibitors. I Drugs. 9(11); 778-782.

Chou KL, Taylor JL, Patil PG (2013). The MDS-UPDRS tracks motor and non-motor improvement due to subthalamic nucleus deep brain stimulation in Parkinson's disease. *Parkinsonism Relat Disord*. 19(11); 966-9.

Cohly HH, Taylor A, Angel MF, *et al* (1998). Effect of turmeric, turmerin and curcumin on H2O2-induced renal epithelial (LLC-PK1) cell injury. *Free Radic Biol Med.* 24(1); 49-54.

Cohn VH, Lyle J (1966). A fluorometric assay for glutathione. Anal Biochem. 14(3); 434-440.

Cole GM, Teter B, Frautschy SA (2007). Neuroprotective effects of curcumin. *Adv Exp Med Biol.* 595; 197-212.

Cools AR, Van Rossum JM (1976). Excitation-mediating and inhibition-mediating dopamine-receptors: a new concept to- wards a better understanding of electrophysiological, biochemi- cal, pharmacological, functional and clinical data. *Psychopharmacologia*. 45; 243-254.

Correia-Melo C, Passos JF (2015). Mitochondria: Are they casual players in cellular senescence?. *Biochim Biophys Acta*. 1847(11); 1373-9.

Coughlan C, Walker DI, Lohr KM, *et al* (2015). Comparative Proteomic Analysis of Carbonylated Proteins from the Striatum and Cortex of Pesticide-Treated Mice. *Parkinsons Dis.* 812532;1-11.

Coulom H, Birman S (2004). Chronic exposure to rotenone models sporadic Parkinson's disease in *Drosophila melanogaster*. J Neurosci. 24(48); 10993-10998.

Craufurd D, Snowden JS (2002). Neuropsychological and neuropsychiatric aspects of cytogenetic damage in patients suffering from oral submucous fibrosis. *Cancer Lett.* 116; 265–269.

Creese I, Burt DR, Snyder SH (1975). Dopamine receptor binding: differentiation of agonist and antagonist states with 3H-dopamine and 3H-haloperidol. *Life Sci.* 17;933–1001.

Cui Q, Li X, Zhu H (2016). Curcumin ameliorates dopaminergic neuronal oxidative damage via activation of the Akt/Nrf2 pathway. *Mol Med Rep*.13(2);1381-1388.

Daher JP, Ying M, Banerjee R, *et al* (2009). Conditional transgenic mice expressing C-terminally truncated human alpha-synuclein (alphaSyn119) exhibit reduced striatal dopamine without loss of nigrostriatal pathway dopaminergic neurons. *Mol Neurodeneger*. 4; 34.

Dai MC, Zhong ZH, Sun YH, et al (2013). Curcumin protects against iron induced neurotoxicity in primary cortical neurons by attenuating necroptosis. *Neurosci lett.* 536(1); 41-6.

Dalle-Donne I, Giustarini D, Colombo R, et al (2003). Protein carbonylation in human diseases. Trens Mol Med. 9(4); 169-176.

Daniel S, Limson JL, Dairam A, *et al* (2004). Through metal binding, curcumin protects against lead- and cadmium-induced lipid peroxidation in rat brain homogenates and against lead-induced tissue damage in rat brain. *J Inorg Biochem.* 98(2);266-75.

Das L, Vinayak M (2014). Long Term Effect of Curcumin in Regulation of Glycolytic Pathway and Angiogenesis via Modulation of Stress Activated Genes in Prevention of Cancer. *PLoS ONE*. 9(6); e99583.

Dawson TM, Dawson VL (2003). Molecular Pathways of Neurodegeneration in Parkinson's Disease. *Science*. 302(5646);819-822.

de Farias CC, Maes M, Bonifácio KL, *et al* (2016). Highly specific changes in antioxidant levels and lipid peroxidation in Parkinson's disease and its progression: Disease and staging biomarkers and new drug targets. *Neurosci Lett.* 617; 66–71.

de Silva HR, Khan NL, Wood NW (2000). The genetics of Parkinson's disease. *Curr Opin Genet Dev.* 10; 292–298.

Dekhar P, Sharma R (2010). Effect of dimethylsulphoxide and curcumin on protein carbonyls and reactive oxygen species of cerebral hemispheres of mice as a function of age. *Int J Dev Neurosci*. 28; 351-57.

Del Maestro R, Mc Donald W (1987). Distribution of superoxide dismutase, glutathione peroxidase and catalase in developing rat brain. *Mech Ageing Dev.* 41; 29–38.

den Brock MG, van Dalen JW, van Gool WA, *et al* (2015). Apathy in Parkinson's disease: A systematic review and meta-analysis. *Mov Disord*. 30(6); 759-769.

Dexter DT, Holley AE, Flitter WD, et al (1994). Increased levels of lipid peroxidases in the parkinsonian substantia nigra: an HPLC and ESR study. Mov Disord. 9(1); 92-97.

Dhillon AS, Tarbutton GL, Levin JL, *et al* (2008). Pesticide/environmental exposure and Parkinson's disease in East Texas. *J Agromedicine*. 13(1); 37-48.

Di Fonzo A, Tassorelli C, De Mari M, *et al* (2006). Comprehensive analysis of the LRRK2 gene in sixty families with Parkinson's disease. *Eur J Hum Genet*. 14;322–331.

Di Monte DA, Lavasani M, Manning Bog AB (2002). Environmental factors in Parkinson's disease. *Neurotoxicol*.23;487-502.

Dias V, Junn E, Mouradian M (2013). The Role of Oxidative Stress in Parkinson's Disease. *J Parkinsons Dis.* 3(4); 461–491.

Dick S, Semple S, Dick F, et al (2007). Occupational titles as risk factors for Parkinson's disease. Occup Med (Lond). 57(1); 50-56.

Dixit VP, Jain P, Joshi SC (1988). Hypolipidaemic effects of *Curcuma longa* L and Nardostachysjatamansi DC in triton-induced hyperlipidaemicrats. *Ind J Physiol Pharmacol.* 32; 299-304.

Dkhar P, Sharma R (2013). Attenuation of age-related increase of protein carbonylation in the liver of mice by melatonin and curcumin. *Mol Cell Biochem.* 380(1-2); 153-60.

Dobrossy MD, Svendsen CN, Dunnett SB (1996). Bilateral striatallesions impair retention of an operant test of short-term memory. *Brain Res Bull.* 41; 159–165.

Dogan VB, Koksal A, Dirican A, et al (2015). Independent effect of fatigue on health-related quality of life in patients with idiopathic Parkinson's disease. *Neurol Sci.* 36(12); 2221-2226.

Dono ND (2013). Turmeric (*Curcuma longa Linn.*) supplementation as an alternative to antibiotics in poultry diets. *Wartazoa.* 23; 41-49.

Du XX, Xu HM, Jiang H, *et al* (2012). Curcumin protects nigral dopaminergic neurons by iron chelation in the 6-hydroxydopamine rat model of Parkinson's disease. *Neurosci Bull.* 28(3):253-258.

Dumont M, Beal MF (2011). Neuroprotective strategies involving ROS in Alzheimer disease. *Free Radic Biol Med.* 51(5); 1014-26.

Dunnett SB, Nathwani F, Brasted PJ (1999). Medial prefrontal andneostriatal lesions disrupt performance in an operant delayed alter-nation task in rats. *Behav Brain Res.* 106;13–28.

Dutta S, Padhye S, Priyadarsini KI, et al (2005). Antioxidant and antiproliferative activity of curcumin semicarbazone. Bioorg Med Chem Lett. 15(11);2738-2744.

Edvardson S, Cinnamon Y, Ta-Shma A, *et al* (2012). A deleterious mutation in DNAJC6 encoding the neuronal-specific clathrin-uncoating co-chaperone auxilin, is associated with juvenile Parkinsonism. *PLoS One.* 7; e36458.

Eisen JS, Smith JC (2008). Controlling morpholino experiments: don't stop making antisense. *Developmental*. 135(10); 1735-43.

Elbaz A, Clavel J, Rathouz PJ, *et al* (2009). Professional exposure to pesticide and Parkinson's disease. *Ann Neurol.* 66(4); 494-504.

Elbaz A, Levecque C, Clavel J, *et al* (2004). CYP2D6 polymorphism, pesticide exposure, and Parkinson's disease. *Ann Neurol.* 55(3);430-4.

Ellman GL (1959). Tissue sulfhydryl groups. Arch Biochem Biophys. 82;70-77.

Ellmann GE, Courtney KD, Anderson V, *et al* (1961). A new calorimetric determination of acetyl cholinesterase activity. *Biochem Pharmacol.* 7; 88–95.

Fahn S (2000). The spectrum of levodopa-induced dyskinesias. Ann Neurol. 47(4 Suppl 1):S2-9.

Fasano A, Visanji NP, Liu LW, et al (2015). Gastrointestinal dysfunction in Parkinson's disease. Lancet Neurol. 14(60); 625-639.

Feany MB, Bender WW (2000). A Drosophila model of Parkinson's disease. Nature. 404(6776); 394-398.

Feng J, Tao T, Yan W, *et al* (2014). Curcumin Inhibits Mitochondrial Injury and Apoptosis from the Early Stage in EAE Mice. *Oxid Med Cell Longev* 1-10; 728751.

Finch CE, Pike MC, Witten M (1990). Slow mortality rate accelerations during aging in animals approximate that of humans. *Science*.249; 902–905.

Flinn L, Bretaud S, Lo C, et al (2008). Zebrafish as a new animal model for movement disorders. J Neurochem. 106(5); 1991-1997.

Floor E, Wetzel MG (1998). Increased protein oxidation in human substantia nigra pars compacta in comparison with basal ganglia and prefrontal cortex measured with an improved dinitrophenylhydrazine assay. *J Neurochem.* 70(1); 268-75.

Folstein MF, Folstein SE, Brandt J (1990). Huntington 's disease: Cummings J.L. (Ed.). Subcortical dementia. *Oxford University Press*, New York.

Fong CS, Wu RM, Shieh JC, *et al* (2007). Pesticide exposure on southern Taiwanese with MnSOD and NQO1 polymorphisms is associated with increased risk of Parkinson's disease. *Clin Chim Acta*. 378(1-2); 136-141.

Friggi-Grelin F, Coulom H, Meller M, *et al* (2003). Targeted gene expression in Drosophila dopaminergic cells using regulatory sequences from tyrosine hydroxylase. *J Neurobiol.* 54;618-627.

Fritsch T, Smyth KA, Wallendal MS, et al (2012). Parkinson's disease: research update and clinical management. South Med J. 105(12); 650-660.

Funayama M, Hasegawa K, Kowa H, *et al* (2002). A new locus for Parkinson's Disease (PARK8) maps to chromosome 12p11.2–q13.1. *Ann Neurol*.51;296–301.

Furtado S, Payami H, Lockhart PJ, et al (2004). Profile of families with parkinsonism predominant spinocerebellar ataxia type 2 (SCA2). Mov Disord. 19(6); 622-629.

Gaiano N, Fishell G (2002). The role of Notch in promoting glial and neural stem cell fates. *Annu Rev Neurosci*. 25;471-490.

García Ruiz PJ (2004). "Prehistoria de la enfermedad de Parkinson" (Prehistory of Parkinson's disease). *Neurologia (in Spanish).* 19, 35–37.

Garodia P, Ichikawa H, Malani N, *et al* (2007). From ancient medicine to modern medicine: Ayurvedic concepts of health and their role in inflammation and cancer. *J Soc Integr Oncol.*5;25–37.

Gebicki JM (1997). Protein hydroperoxidase as new reactive oxygen species. Redox Rep. 3(2); 99-110.

Genestra M (2007). Oxyl radicals, redox-sensitive signaling cascades and antioxidants. *Cell Signal*. 19;1807–1819.

George EB, Alexander I, Valentina EM (2014). Beneficial effects of nicotine, cotinine and its metabolites as potential agents for Parkinson's disease. *Front Aging Neurosci.* 6; 1-13.

Geser F, Brandmeir NJ, Kwong LK, *et al* (2008). Evidence of multisystem disorder in whole-brain map of pathological TDP-43 in amyotrophic lateral sclerosis. *Arch Neuro*. 65; 636–641.

Girish C, Muralidhara (2012). Propensity of *Selaginella delicatula* aqueous extract to offset rotenoneinduced oxidative dysfunctions and neurotoxicity in *Drosophila melanogaster*: Implications for Parkinson's disease. *Neuro Toxicology*.33(3); 444-456.

Girotti AW (1985). Mechanisms of lipid peroxidation. J Free Radic Biol Med. 1(2); 87-95.

Gleason K, Shine JP, Shobnam N, *et al* (2014). Contaminated turmeric is a potential source of lead exposure for children in rural Bangladesh. *J Environ Public Health.* 2014; 730636.

Goel A, Kunnumakkara AB, Aggarwal BB (2008). Curcumin as "Curecumin": from kitchen to clinic. *Biochem Pharmacol.* 75(4);787-809.

González-Burgos E, Fernández-Moriano C, Lozano R, *et al* (2017). Ginsenosides Rd and Re co-treatments improve rotenone-induced oxidative stress and mitochondrial impairment in SH-SY5Y neuroblastoma cells. *Food Chem Toxico*.109(1);38-47.

Goodpasture CE, Arrighi FE (1976). Effects of food seasonings on the cell cycle and chromosome morphology of mammalian cells in vitro with special reference to turmeric. *Food Cosmet Toxicol*.14(1);9-14.

Goud VK, Polasa K, Krishnaswamy K (1993). Effect of turmeric on xenobiotic metabolising enzymes. *Plant Foods Hum Nutr*.44(1);87–92.

Gowda NK, Ledoux DR, Rottinghaus GE, *et al* (2008). Efficacy of turmeric (*Curcuma longa*), containing a known level of curcumin, and a hydrated sodium calcium aluminosilicate to ameliorate theadverse effects of aflatoxin in broiler chicks. *Poult Sci*.87;1125–1130.

Goyal NA, Mozaffar T (2014). Respiratory and nutritional support in amyotrophic lateral sclerosis. *Curr Treat Options Neurol*. 16(2);270.

Green DR and Reed JC (1998). Mitochondria and apoptosis. Science. 281(5381); 1309-1312.

Guehne U, Riedel-Heller S, Angermeyer MC (2005). Mortality in dementia. *Neuroepidemiology*. 25(3);153-162.

Gupta SC, Patchva S, Aggarwal BB (2012). Therapeutic Roles of Curcumin: Lessons Learned from Clinical Trials. *The AAPS Journal*. 15(1);195-218.

Gusella JF, MacDonald ME (1995). Huntington's disease: CAG genetics expands neurobiology. *Curr Opin Neurobiol.* 5;656–662.

Guthenberg C, Alin P, Mannervik B (1985). Glutathione transferase from rat testis. *Meth Enzymol.* 113; 507–510.

Gwinn-Hardy K, Chen JY, Liu HC, *et al* (2000). Spinocerebellar ataxia type 2 with parkinsonism in ethnic Chinese. *Neurology*. 55(6); 800-805.

Halliwell B (2006). Reactive Species and Antioxidants. Redox Biology Is a Fundamental Theme of Aerobic Life. *Plant Physiol.* 141(2); 312–322.

Halliwell B, Gutteridge JM (1990). Role of free radicals and catalytic metal ions in human disease: an overview. *Meth Enzymol.* 186:1-85.

Hampshire DJ, Roberts E, Crow Y, *et al* (2001). Kufor-Rakeb syndrome, pallido-pyramidal degeneration with supranuclear up gaze paresis and dementia, maps to 1p36. *J Med Genet*, 38;680-682.

Hamre K, Tharp R, Poon K, *et al* (1999). Differential strain susceptibility following 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) administration acts in an autosomal dominant fashion: quantitative analysis in seven strains of Mus musculus. *Brain res.* 828(1-2); 91-103.

Hastak K, Lubri N, Jakhi SD, et al (1997). Effect of turmeric oil and turmeric oleoresin on cytogenetic damage in patients suffering from oral submucous fibrosis. Cancer Lett. 116(2);265-9.

Hagl S, Kocher A, Schiborr C, *et al* (2015). Curcumin micelles improve mitochondrial function in neuronal PC12 cells and brains of NMRI mice - Impact on bioavailability. *Neurochem Int.* 89;234-42.

Hayes JD, Strange RC (2000). Glutathione S-transferase polymorphism and their biological consequences. *Pharmacology*. 61; 154–166.

Healy DG, Falchi M, O'Sullivan SS, *et al* (2008). Phenotype, genotype, and worldwide genetic penetrance of LRRK2-associated Parkinson's disease: a case-control study. *Lancet Neurol.* 7;583–590.

Hearn MG, Ren Y, McBride EW, et al (2002). A Drosophila dopamine 2-like receptor: Molecular characterization and identification of multiple alternatively spliced variants. Proc Natl Acad Sci U S A. 99(22); 14554-14559.

Heemels MT (2016). Neurodegenerative diseases. Nature. 539;179.

Hegde SN, Naseerulla MK, Krishna MS (2000). Variability of morphological traits in *Drosophila* bipectinata complex. *Indian J Exp Biol.* 38(8); 797-806.

Henrotin Y, Clutterbuck AL, Allaway D, et al (2010). Biological actions of curcumin on articular chondrocytes. Osteoarth and Cartil. 18(2);141–149.

Hertzman C, Wiens M, Bowering D, et al (1990) Parkinson's disease: a case-control study of occupational and environmental risk factors. Am J Ind Med. 17; 349–355.

Holtzman DM, Bales KR, Tenkova T, *et al* (2000). Apolipoprotein E isoform-dependent amyloid deposition and neuritic degeneration in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A*.97(6);2892-2897.

Hornykiewicz O, Kish SJ (1987). Biochemical pathophysiology of Parkinson's disease. Adv Neurol. 45;19–34. https://doi.org/10.1016/C2009-0-48725-7.

Huang L, Chen C and Zhang X, *et al* (2018). Neuroprotective Effect of Curcumin Against Cerebral Ischemia-Reperfusion Via Mediating Autophagy and Inflammation. *J Mol Neurosci.* 64(1);129-139.

Inamdar AA, Chaudhuri A, O'Donnell J (2012). The Protective Effect of Minocycline in a Paraquat-Induced Parkinson's Disease Model in *Drosophila* is Modified in Altered Genetic Backgrounds. *Parkinsons Dis.*2012;938528.

Isik H, Sahbaz A, Timur H, et al (2017). The Use of Thiol/Disulfide as a Novel Marker in Premature Ovarian Failure. *GynecolObstet Invest.* 82; 113-118.

Iversen SD, Iversen LL (2007). Dopamine: 50 years in perspective. Trends Neurosci. 30;188-193.

Iwunze MO (2004). Fluorescence Quenching Studies of Curcumin by Hydrogen Peroxide in Acetonitrile Solution. *Chemical Monthly*. 135(3); 231-240.

Jagatha B, Mythri RB, Vali S, *et al* (2008). Curcumin treatment alleviates the effects of glutathione depletion in vitro and in vivo: Therapeutic implications for Parkinson's disease explained via in silico studies. *Free Radic Biol Med.* 44;907–917.

Jahromi SR, Haddadi M, Shivanandappa T, *et al* (2015). Attenuation of neuromotor deficits by natural antioxidants of *Decalepishamiltonii* in transgenic *Drosophila* model of Parkinson's disease. *Neurosci*. 293;136-150.

Jain A, Martensson J, Stole E, *et al* (1991). Glutathione deficiency leads to mitochondrial damage in brain. *Proc Natl Acad Sci USA*. 88; 1913–1917.

Jayaraj RL, Elangovan N, Manigandan K, et al (2014). CNB-001 a Novel curcumin derivative, guards dopamine neurons in MPTP model of Parkinson's disease. *Biomed Res Int.* 2014; 236182.

Jellinger KA (2009). Formation and development of Lewy pathology: a critical update. *J Neurol*.256 (Suppl 3); 270-279.

Jenner P (2003). Dopamine agonists, receptor selectivity and dyskinesia induction in Parkinson's disease. *Curr Opin Neurol*.1;3-7.

Jenner P, Olanow CW (2006). The pathogenesis of cell death in Parkinson's disease. *Neurology*. 66 (10 suppl 4); S24-S36.

Jhonsa DJ, Badgujar LB, Sutariya BK, *et al* (2016). Neuroprotective effect of flavonoids against paraquat induced oxidative stress and neurotoxicity in drosophila melanogaster.*Curr top nutraceutical res.* 14(4); 283-294.

Jia Z, Babu PV, Che W, *et al* (2018). Natural Products Targeting on Oxidative Stress and Inflammation: Mechanisms, Therapies, and Safety Assessment. *Oxid Med Cell Longev*. 1-3; 6576093.

Jiang TF, Zhang YJ, Zhou HY, *et al* (2013). Curcumin ameliorates the neurodegenerative pathology in A53T α -synuclein cell model of Parkinson's disease through the downregulation of mTOR/p70S6K signaling and the recovery of macroautophagy. *J Neuroimmune Pharmacol.* 8(1);356-369.

Jiao Y, Wilkinson J, Di X, *et al* (2009). Curcumin, a cancer chemopreventive and chemotherapeutic agent, is a biologically active iron chelator. *Blood*.113;462-469.

Johnson WM, Wilson-Delfosse AL, Mieyal JJ (2012). Dysregulation of glutathione homeostasis in neurodegenerative diseases. *Nutrients*. 4(10); 1399-1440.

Kamel F, Tanner C, Umbach D, *et al* (2007). Pesticide exposure and self-reported Parkinson's disease in the agricultural health study. *Am J Epidemiol*. 1656(4); 364-367.

Karalliedde (1999). Organophosphorus poisoning and anaesthesia. Anaesthesia. 54; 1073-1088.

Kebabian JW, Greengard P (1971). Dopamine-sensitive adenyl cyclase: Possible role in synaptic transmission. *Science*. 174;1346-1349.

Khan MM, Ahmad A, Ishrat T, *et al* (2010). Resveratrol attenuates 6-hydroxydopamine- induced oxidative damage and dopamine depletion in rat model of Parkinson's disease. *Brain Res.* 1328; 139-51.

Khan NL, Brooks DJ, Pavese N, *et al* (2002). Progression of nigrostriatal dysfunction in a parkin kindred: an [18F] dopa PET and clinical study. *Brain*,125;2248–2256.

Khatri DK, Juvekar AR (2016). Kinetics of Inhibition of Monoamine Oxidase Using Curcumin and Ellagic Acid. *Pharmacogn Mag*.12(Suppl 2);116-120.

Khatri DK, Juvekar AR (2016). Neuroprotective effect of curcumin as evidenced by abrogation of rotenoneinduced motor deficits, oxidative and mitochondrial dysfunctions in mouse model of Parkinson's disease. *Pharmacol Biochem Behav.* 150-151; 39-47.

Khoo TK, Yarnall AJ, Duncan GW, *et al* (2013). The spectrum of nonmotor symptoms in early Parkinson disease. *Neurology*. 80(3); 276-281.

Kim JS, Youn J, Shin H, *et al* (2013). Nonmotor symptoms in drug-induced Parkinsonism and drug-naïve Parkinson's disease. *Can J Neurol Sci*. 40(1); 36-41.

Kirkwood TBL, Austad SN (2000). Why do we age? Nature. 408;233-238.

Kirtikar KR, Basu BD, Blatter E, et al (1993). Indian Medicinal Plants. 2nd Ed. 2;1182.

Kitada T, Asakawa S, Hattori N, *et al* (1998). Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. *Nature*. 392;605–608.

Knudsen K, Damholdt FM, Mourisden K, *et al* (2015). Olfactory function in Parkinson's disease-effect of training. *Acta Neurol Scand*, 132(6); 395-400.

Kojro E, Fahrenholz F (2005). The non-amyloidogenic pathway: structure and function of alpha-secretases. *Subcell Biochem*. 38;105–127.

Koller WC (1986). Paraquat and Parkinson's disease. Neurology.36 (8);1147.

Kolodziejczyk J, Olas B, Saluk-Juszczak J, et al (2011). Antioxidative properties of curcumin in the protection of blood platelets against oxidative stress in vitro. *Platelets*. 22; 270-76.

Kopan R, Ilagan MXG (2009). The Canonical Notch Signaling Pathway: Unfolding the Activation Mechanism. *Cell*. 137(2);216–233.

Kostyuk VA, Potapovich AI (1989). Superoxide-driven oxidation of quercetin and a simple sensitive assay for determination of superoxide dismutase. *Biochem Int.* 19(5); 1117-1124.

Krebs CE, Karkheiran S, Powell JC, *et al* (2013). The Sac1 domain of SYNJ1 identified mutated in a family with early-onset progressive Parkinsonism with generalized seizures. *Hum Mutat.* 34;1200–1207.

Kriscenski-Perry E, Durham HD, Sheu SS, *et al* (2002). Synergistic effects of low-level stressors in an oxidative damage model of spinal motor neuron degeneration. *Amyotroph Lateral Scler Other Motor Neuron Disord.*3(3);151-157.

Krishnan S, Sarma G, Sarma S, *et al* (2011). Do nonmotor symptoms in Parkinson's disease differ from normal aging?. *Mov Disord*. 26(11); 2110-3.

Kroemer G, Reed JC (2000). Mitochondrial control of cell death. Nat Med. 6(5); 513-519.

Kuilman T, Michaloglou C, Mooi WJ, et al (2010). The essence of senescence. Genes Dev. 24(22); 2463-79.

Kuroda M, Mimaki Y, Nishiyama T, *et al* (2005). Hypoglycemic effects of turmeric (*Curcuma longa* L. rhizomes) on genetically diabetic KK-Ay mice. *Biol Pharm Bull*.28;937–939.

Kurtis MM, Rodriguez-Blazquez C, Martinez-Martin P, *et al* (2013). Relationship between sleep disorders and other non-motor symptoms in parkinson's disease. *Parkinsonism Relat Disord*. 19(12); 1152-1155.

Kurtishi A, Rosen B, Patil KS, *et al* (2018). Cellular Proteostasis in Neurodegeneration. *Mol Neurobiol*. 1-14. doi: 10.1007/s12035-018-1334-z.

Kuttan R, Sudheeran PC, Josph CD (1987). Turmeric and curcumin as topical agents in cancer therapy. *Tumori*. 73;29–31.

L'opez-Ot'ın C, Blasco MA, Partridge L, et al (2013). The hallmarks of aging. Cell. 153(6); 1194–1217.

Lal B, Kappor AK, Astha, et al (1999). Efficacy of Curcumin in the Management of Chronic Anterior Uveitis. *Phytother Res.* 13(4); 318-322.

Lang AE, Lozano AM (1998). Parkinson's disease. First of two parts. N Engl J Med. 339; 1044-1053.

Langston JW, Ballard P, Tetrud JW, et al (1983). Chronic parkinsonism in humans due to a product of meperidine-analog synthesis. Science. 219; 979-980.

Lebestky T, Chang JS, DankertH, *et al* (2009). Two different forms of arousal in Drosophila are oppositely regulated by the dopamine D1 receptor ortholog DopR via distinct neural circuits. *Neuron*. 64(4); 522-36.

Lee KS, Lee BS, Semnnani S, *et al* (2010). Curcumin extends life span, improves health span, and modulates the expression of age-associated aging genes in *Drosophila melanogaster*. *Rejuvenation Res.* 13(5); 561–570.

Lees AJ (2007). Unresolved issues relating to the shaking palsy on the celebration of James Parkinson's 250th birthday. *Mov Disord*. 22(17); S327–334.

Lee WH, Loo CY, Bebawy M, et al (2013). Curcumin and its derivatives: their application in neuropharmacology and neuroscience in the 21st century. Curr Neuropharmacol. 11(4); 338-78.

Leroy E, Boyer R, Auburger G, et al (1998). The ubiquitin pathway in Parkinson's disease. Nature. 395; 451-452.

Lesage S, Anheim M, Letournel F, *et al* (2013). G51D α -synuclein mutation causes a novel parkinsonianpyramidal syndrome. *Ann Neurol.* 73(4); 459-71.

Lessing D, Bonini NM (2009). Maintaining the Brain: Insight into Human Neurodegeneration from *Drosophila* Mutants. *Nat Rev Genet*. 10(6); 359.

Levine RL, Garland D, Oliver C, *et al* (1990). Determination of carbonyl content in oxidatively modified proteins. *Meth Enzymol.* 186; 464–78.

Levy G, Schupf N, Tang MX, *et al* (2002). Combined effect of age and severity on the risk of dementia in Parkinson's disease. *Ann Neurol.* 51;722–729.

Li SH, Li XJ (2004). Huntingtin-protein interactions and the pathogenesis of Huntington's disease. *Trends Genet.* 20;146-154.

Liao VH, Yu CW, Chu YJ, et al (2011). Curcumin-mediated lifespan extension in *Caenorhabditis elegans*. *Mech Ageing Dev*. 132(10);480-487.

Lin X, Parisiadou L, Gu XL, et al (2009). Leucine-rich repeat kinase 2 regulates the progression of neuropathology induced by Parkinson's-disease-related mutant alpha-synuclein. Neuron. 64(6); 807-827.

Lincoln SJ, Maraganore DM, Lesnick TG, *et al* (2003). Parkin variants in North American Parkinson's disease: cases and controls. *Mov Disord*. 18;1306–1311.

Liou HH, Tsai MC, Chen CJ, et al (1997). Environmental risk factors and Parkinson's disease: a case-control study in Taiwan. Neurology. 48(6); 1583-1588.

Liu Z, Li T, Yang D, et al (2013). Curcumin protects against rotenone-induced neurotoxicity in cell and drosophila models of Parkinson's disease. Adv Parkinson's Dis. 2(1);18-27.

Lloyd TE, Taylor JP (2010).Flightless flies: *Drosophila* models of neuromuscular disease. *Ann N Y Acad Sci.* 1184; 1-20.

Lohmann E, Periquet M, Bonifati V, *et al* (2003). How much phenotypic variation can be attributed to parkin genotype?. *Ann Neurol.* 54;176–185.

Luceri C, Bigagli E, Femia AP, *et al* (2017). Aging related changes in circulating reactine oxygen species (ROS) and protein carbonyls are indicative of liver oxidative injury. *Toxicol Rep.* 5; 141-145.

Mackenzie IR, Bigio EH, Ince PG, *et al* (2007). Pathological TDP-43 distinguishes sporadic amyotrophic lateral sclerosis from amyotrophic lateral sclerosis with SOD1 mutations. *Ann Neurol*.61; 427-434.

Mancuso C, Barone E (2009). The Heme Oxygenase/Biliverdin Reductase Pathway in Drug Research and Development. *Curr Drug Metabol*. 10; 579-594.

Manning-Bog AB, McCormack AL, Li J, *et al* (2002). The herbicide paraquat causes up-regulation and aggregation of alpha-synuclein in mice: paraquat and alpha-synuclein. *J Biol Chem.* 277(3); 1641-4.

Manyam BV (1990). Paralysis agitans and levodopa in "Ayurveda": Ancient Indian medical treatise. Mov.Disord. 5, 47–48.

Maries E, Dass B, Collier TJ, et al (2003). The role of alpha-synucleininParkinson's disease: insights from animal models. *Nat Rev Neurosci.* 4;727–738.

Marshall JW, Ridley RM (1996). Assessment of functional impairment following permanent middle cerebral artery occlusion in a non-human primate species. *Neurodegeneration*. 5(3); 275-286.

Martin CA, Barajas A, Lawless G, *et al* (2014). Synergistic effects on dopamine cell death in a *Drosophila* model of chronic toxic exposure. *Neurotoxicology*. 44; 344-351.

Marttila RJ, Lorentz H, Rinne UK (1998). Oxygen toxicity protecting enzymes in Parkinson's disease: Increase of superoxide dismutase like activity in the substantia nigra and basal nucleus. *J Neurol Sci.* 86(2–3); 321–331.

Massano J, Bhatia KP (2012). Clinical approach to Parkinson's disease: features, diagnosis, and principles of management. *Cold Spring Harb Perspect Med.*2: a008870.

Masson P, Lockridge (2010). Butyrylcholinesterase for protection from organophosphate poisons: catalytic complexities and hysteretic behavior. *Arch. Biochem. Biophys.* 494; 107–120.

Massoulie J, Pezzementi L, Bon S, et al (1993). Molecular and cellular biology of cholinesterases. Prog Neurobiol. 41; 31–91.

Mata IF, Yearout D, Alvarez V, *et al* (2011). Replication of MAPT and SNCA, but not PARK16-18, as susceptibility genes for Parkinson's disease. *Mov Disord*. 26(5); 819-823.

Matsumine H, Yamamura Y, Hattori N, *et al* (1998). A microdeletion of D6S305 in a family of autosomal recessive juvenile parkinsonism (PARK2). *Genomics*. 49(1); 143-146.

Matthews RP, Plumb-Rudewiez N, Lorent K, *et al* (2005). Zebrafish vps33b, an ortholog of the gene responsible for human arthrogryposis-renal dysfunction-cholestasis syndrome, regulates biliary development downstream of the onecut transcription factor hnf6. *Development*. 132(23); 5295-306.

Mattson MP (1998). Modification of ion homeostasis by lipid peroxidation: roles in neuronal degeneration and adaptive plasticity. *Trends Neurosci.* 21(2); 53-7.

McHugh PC, Buckley DA (2015). The structure and function of the dopamine transporter and its role in CNS diseases. *VitamHorm.* 98; 339-69.

Means RT Jr (2009). Ironing out complementary medicine. Blood. 113;270-271.

Mehdi SH, Qamar A (2013). Paraquat-induced ultrastructural changes and DNA damage in the nervous system is mediated via oxidative-stress-induced cytotoxicity in *Drosophila melanogaster.Toxicol Sci.*134(2);355-365.

Mellick GD (2006). CYP450, genetics and Parkinson's disease: gene x environment interactions hold the key. *J Neural Transm Suppl*.70;159-165.

Mendez M, Mendez-Lopez M, Lopez L, et al (2011). Acetylcholinesterase activity in an experimental rat model of Type C hepatic encephalopathy. Acta Histochem. 113(3); 358-362.

Menegon A, Board PG, Blackburn AC, et al (1998). Parkinson's disease, pesticides, and glutathione transferase polymorphisms. Lancet. 352(9137);1344-1346.

Miller OG, Behring JB, Siedlak SL, *et al* (2016). Upregulation of Glutaredoxin 1 Activates Microglia and Promotes Neurodegeneration: Implications for Parkinson's Disease. *Antioxid Redox Signal.* 25(18); 967–982.

Miquel J, Bernd A, Sempere JM, et al (2002). The curcuma antioxidants: pharmacological effects and prospects for future clinical use. A review. Arch Geronto Geriatr. 34;37-46.

Missale C, Nash SR, Robinson SW, et al (1998). Dopamine receptors: from structure to function. Physiol Rev. 78;189–225.

Mochel F, Charles P, Seguin F, *et al* (2007). Early energydeficit in Huntington disease: identification of a plasma biomarker traceable duringdisease progression. *PloS one*.2;647.

Modi P, Mohamad A, Phom L, *et al* (2016). Understanding Pathophysiology of Sporadic Parkinson's Disease in *Drosophila* Model: Potential Opportunities and Notable Limitations. In Challenges in Parinson's Disease. Ed. Jolanta Dorszewska. *Intech Open.* 11; 217-44.

Munoz-Sanjuan I, Bates GP (2011). The importance of integrating basic and clinical research toward the development of new therapies for Huntington disease. *J Clin Invest.* 121;476-483.

Murray-Stewart T, Dunworth M, Lui Y, et al (2018). Curcumin mediates polyamine metabolism and sensitizes gastrointestinal cancer cells to antitumor polyamine-targeted therapies. PLoS One. 13(8);202677.

Mythri RB, Bharath MM (2012). Curcumin: a potential neuroprotective agent in Parkinson's disease. *Curr Pharm Des*. 18(1);91-99.

Nachmensohn D (1959). Chemical and molecular basis of nerve activity. Academic press, New York and London. 59; 1-235.

Najim al-Din AS, Wriekat A, Mubaidin A, *et al* (1994). Pallido-pyramidal degeneration, supranuclear upgaze paresis and dementia: Kufor-Rakeb syndrome. *Acta Neurologica Scandinavica*. 89;347-352.

Nakabeppu Y, Tsuchimoto D, Yamaguchi H, et al (2007). Oxidative damage in nucleic acids and parkinson's disease. J Neurosci Res. 85(5);919-934.

Nam SM, Choi JH, Yoo DY, *et al* (2014). Effects of curcumin (*Curcuma longa*) on learning and spatial memory as well as cell proliferation and neuroblast differentiation in adult and aged mice by upregulating brain-derived neurotrophic factor and CREB signaling. *J Med Food*. 17(6);641-649.

Natural Toxicology Program (1993). NTP Toxicology and Carcinogenesis Studies of Turmeric Oleoresin (CAS No. 8024-37-1) (Major Component 79%-85% Curcumin, CAS No. 458-37-7) in F344/N Rats and B6C3F1 Mice (Feed Studies). *Natl Toxicol Program Tech Rep Ser*.427;1-275.

Neumann M, Sampathu DM, Kwong LK, *et al* (2006). Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science*. 14;130 – 133.

Newman DJ, Cragg GM (2007). Natural products as sources of new drugs over the last 25years. J Nat Prod.70;461–477.

Ng C, Ko C, Koon C, *et al* (2016). The aqueous extract of rhizome of *Gastrodia elata* Blume attenuates locomotor defect and inflammation after traumatic brain injury in rats. *J Ethnopharmacol*.185;87-95.

Ng L, Talman P, Khan F (2011). Motor neurone disease: Disabilityprofile and service needs in an Australian cohort. *Int J Rehabil Res.* 34;151–159.

Nguyen TT, Vuu MD, Huynh MA, *et al* (2018). Curcumin Effectively Rescued Parkinson's Disease-Like Phenotypes in a Novel *Drosophila melanogaster* Model with dUCH Knockdown. *Oxid Med Cell Longev*.1-12; 2038267; doi: 10.1155/2018/2038267.

Niedzielska E, Smaga I, Gawlik M, et al (2016). Oxidative Stress in Neurodegenerative Diseases. Mol Neurobiol. 53(6); 4094–4125.

Ning YP, Kanai K, Tomiyama H, *et al* (2008). PARK9-linked parkinsonism in eastern Asia: Mutation detection in ATP13A2 and clinical phenotype. *Neurology*. 70;1491-1493.

Nishioka K, Hayashi S, Farrer MJ, et al (2006). Clinical heterogeneity of alpha-synuclein gene duplication in Parkinson's disease. Ann Neurol. 59(2); 298-309.

Niso-Santano M, Morán JM, García-Rubio L, *et al* (2006). Low Concentrations of Paraquat Induces Early Activation of Extracellular Signal-Regulated Kinase 1/2, Protein Kinase B, and c-Jun N-terminal Kinase 1/2 Pathways: Role of c-Jun N-Terminal Kinase in Paraquat-Induced Cell Death. *Toxicol Sci.* 92(2); 507-515.

Niu Y, Guo X, Chen Y, *et al* (2015). Early Parkinson's disease symptoms in alpha-synucleintransgenic monkeys. *Hum Mol Genet.* 24(8); 2308-2317.

Niveditha S, Ramesh SR, Shivanandappa T (2017). Paraquat induced movement disorder in relation to oxidative stress-mediated neurodegeneration in the brain of *Drosophila melanogaster*. *Neurochem Res.* 42(11); 3310-3320.

Nuytemans K, Bademci G, Inchausti V, *et al* (2013). Whole exome sequencing of rare variants in EIF4G1 and VPS35 in Parkinson disease. *Neurology*. 80; 982-989.

Nuytemans K, Theuns J, Cruts M, *et al* (2010). Genetic etiology of Parkinson disease associated with mutations in the SNCA, PARK2, PINK1, PARK7, and LRRK2 genes: a mutation update. *Hum Mutat.* 31; 763–780.

O'Brien RJ, Wong PC (2011). Amyloid precursor protein processing and Alzheimer's disease. *Annu Rev Neurosci.* 34;185-204.

Oetari S, Sudibyo M, Commandeur JN, *et al* (1996). Effects of curcumin on cytochrome P450 and glutathione S-transferase activities in rat liver. *Biochem Pharmacol.* 51(1); 39-45.

Ohakawa H, Ohishi U, Yagi K (1979). Assay of lipid peroxidation in rat tissues by thiobarbituric reaction. *Anal Biochem.* 95; 145–9.

Okray Z, Hassan BA (2013). Genetic approaches in *Drosophila* for the study of neurodevelopmental disorders. *Neuropharmacol.* 68; 150-156.

Olanow CW, Tatton WG (1999). Etiology and pathogenesis of Parkinson's disease. Annu Rev Neurosci.22;123-144.

Oliveira MAP, Balling R, Smidt MP, *et al* (2017). Embryonic development of selectively vulnerable neurons in Parkinson's disease. *NPJ Parkinsons Dis.* 3; 21.

Ossowska K, Wardas J, Kuter K, *et al* (2005). Influence of paraquat on dopaminergic transporter in the rat brain. *Pharm reports*. 57; 330-335.

Outeiro TF, Kontopoulos E, Altmann SM, *et al* (2007). Sirtuin 2 inhibitors rescue alpha-synucleinmediated toxicity in models of Parkinson's disease. *Science*. 317(5837); 516-9.

Paandaresh MD, Shrivash MK, Naveen Kumar HN, *et al* (2016). Curcumin Monoglucoside shows improved bioavailability and mitigates rotenone induced neurotoxicity in cell and Drosophila models of Parkinson's disease. *Neurochem* Res. 41(11); 3113-3128.

Paisan-Ruiz C, Bhatia KP, Li A, *etal* (2009). Characterization of PLA2G6 as a locus for dystoniaparkinsonism. *Annals of Neurology*. 65;19-23.

Paisán-Ruíz C, Jain S, Evans EW, *et al* (2004). Cloning of the gene containing mutations that cause PARK8-linked Parkinson's disease. *Neuron*. 44;595–600.

Pandey UB, Nichols CD (2011). Human disease models in Drosophila melanogaster and the role of the fly in therapeutic drug discovery. *Pharmacol Rev.* 63(2); 411–436.

Park JH, Jung JW, Ahn YJ, *et al* (2012). Neuroprotective properties of phytochemicals against paraquatinduced oxidative stress and neurotoxicity in Drosophila melanogaster. *PesticideBiochem Physiol*. 104(2); 118–125. Parkinson J (1817). An essay on the shaking palsy. Whittingham and Rowland for Sherwood, Needly and Jones, London.

Pavese N, Gerhard A, Tai YF, et al (2006). Microglial activation correlates with Cell BiochemBiophys severity in Huntington disease: A clinical and PET study. Neurology. 66; 1638–1643.

Peng J, Stevenson FF, Doctrow SR, *et al* (2005). Superoxide dismutase/catalase mimetics are neuroprotective against selective paraquat-mediated dopaminergic neuron death in the substantial nigra: implications for Parkinson disease. *J Biol Chem.* 280(32); 29194-29198.

Perez CA, Tong Y, Guo M (2008). Iron Chelators as Potential Therapeutic Agents for Parkinson's Disease. *CurrBioact Compd.* 4(3);150-158.

Pérez-Barrón, Ávila-Acevedo JG, García-Bores AM, *et al* (2015). Neuroprotective effect of *Buddleja cordata* methanolic extract in the 1-methyl-4-phenylpyridinium Parkinson's disease rat model. J Nat Med. 69(1); 86–93.

Pfeiffer RF (2016). Non-motor symptoms in Parkinson's disease. *Parkinsonism Relat Disord*. 22(Suppl. 1); 119–122.

Phan TT, See P, Lee ST, *et al* (2001). Protective Effects of Curcumin against Oxidative Damage on Skin Cells In Vitro: Its Implication for Wound Healing. *JTrauma: Injury, Infection, and Critical Care.* 51(5); 927-931.

Philippens (2008). Non-human primate models for Parkinson's disease. Drug Discovery Today Disease Models. 5(2); 105–111.

Phom L, Achumi B, Alone DP, *et al.* (2014). Curcumin's neuroprotective efficacy in Drosophila model of Parkinson's disease is phase specific: implications of its therapeutic effectiveness. *Rejuvenation Res.* 17(6); 481-489.

Pienaar IS, Gotz J, Feany MB (2010). Parkinson's disease: insights from non-traditional model organisms. *Prog Neurobiol*. 92(4);558-71.

Pihlstrom L, Rengmark A, BjørnaråKA, et al. (2015). Fine mapping and resequencing of the PARK16 locus in Parkinson's disease. J Hum Genet. 60(7); 357-362.

Pletcher SD, Macdonald SJ, Marguerie J (2002). Genome-Wide Transcript Profiles in Aging and Calorically Restricted *Drosophila melanogaster*. *Curr Biol*. 12(9); 712-723.

Polasa K, Raghuram TC, Krishna TP, et al (1992). Effect of turmeric on urinary mutagens in smokers. Mutagenesis. 7; 107–109.

Polymeropoulos MH, Higgins JJ, Golbe LI, *et al* (1996). Mapping of a gene for Parkinson's disease to chromosome 4q21–q23. *Science*, 274;1197–1199.

Polymeropoulos MH, Lavedan C, Leroy E, *et al* (1997). Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. *Science*. 276; 2045-2047.

Potashkin JA, Blume SR, Runkle NK (2010). Limitations of animal models of Parkinson's disease. *Parkinsons Dis.* 2011; 1-7; 658083.

Prasad S, Tyagi AK, Aggarwal BB (2014). Recent Developments in Delivery, Bioavailability, Absorption and Metabolism of Curcumin: the Golden Pigment from Golden Spice. *Cancer Res Treat.* 46(1); 2–18.

Prasad SN, Muralidhara (2014). Mitigation of acrylamide-induced behavioral deficits, oxidative impairments and neurotoxicity by oral supplements of geraniol (a monoterpene) in rat model. *Chem Biol Interact.* 5; 223-237.

Priyadarshi A, Khuder SA, Schaub EA, et al (2000). A meta-analysis of Parkinson's disease and exposures to pesticides. *Neurotoxicology*. 21; 435-440.

Proukakis C, Dudzik CG, Brier T, *et al* (2013). A novel α -synuclein missense mutation in parkinson's disease. *Neurology*. 80(11); 1062-1064.

Prubing K, Voigt A, Schulz JB (2013) Drosophila melanogaster as a model organism for Alzheimer's disease. *Mol Neurodegener*. 8; 35.

Prucksunand C, Indrasukhsri B, Leethochawalit M, *et al* (2001). Phase II clinical trial on effect of the long turmeric (*Curcuma longa* Linn.) on healing of peptic ulcer. *Southeast Asian J Trop Med Public Health*. 32;208–215.

Quadri M, Fang M, Picillo M, *et al* (2013). Mutation in the SYNJ1 gene associated with autosomal recessive, early-onset Parkinsonism. *Hum Mutat.* 34;1208–1215.

Ragland M, Hutter C, Zabetian C, *et al* (2009). Association between the ubiquitin carboxyl-terminal esterase L1 gene (UCHL1) S18Y variant and Parkinson's disease: A HuGE review and meta analysis. *Am J Epidemiol.* 170; 1344-1357.

RajaSankar S, Manivasagam T, Surendran S (2009). Ashwagandha leaf extract: A potential agent in treating oxidative damage and physiological abnormalities seen in a mouse model of Parkinson's disease. *Neurosci Lett.* 454; 11–15.

Ramanan VK, Saykin AJ (2013). Pathways to neurodegeneration: mechanistic insights from GWAS in Alzheimer's disease, Parkinson's disease, and related disorders. *Am J Neurodegener Dis*.2(3);145-175.

Ramirez A, Heimbach A, Grundemann J, *et al* (2006). Hereditary Parkinsonism with dementia is caused by mutations in ATP13A2, encoding a lysosomal type 5 P-type ATPase. *Nat Genet*. 38;1184–1191.

Ramsey CP, Tsika E, Ischiropoulos H, et al (2010). DJ-1 deficient mice demonstrate similar vulnerability to pathogenic Ala5Thr human alpha-syn toxicity. Hum Mol Genet. 19(8); 1425-37.

Rao SV, Greeshma M, Muralidhara, *et al* (2016). Neuromodulatory Potential of Aqueous Extracts of Cumin, Cinnamon: Evidence from Rotenone Model in Drosophila: Implications to Parkinson's Disease. *Intl J of Neur Res.* 2(3-4); 297-307.

Ravi SK, Narasingappa RB, Joshi CG, et al (2017). Neuroprotective effects of Cassia tora against paraquatinduced neurodegeneration: relevance for Parkinson's disease. *Natural Product Res.* 32(12); 1476-1480.

Ravindran A, Chandrasekaran N, Mukherjee A (2012). Studies on Differential Behavior of Silver Nanoparticles Towards Thiol Containing Amino Acids. *CurrNanosci.* 8(1): 1-9.

Reiter LT, Potocki L, Chien S, et al (2001). A systematic analysis of human disease-associated genes sequences in Drosophila melanogaster. Genome Res. 11(6); 1114-1125.

Rejinders JS, Ehrt U, Weber WE, et al (2008). A systemic review of prevalence studies of depression in Parkinson's disease. Mov Disord. 23(2); 183-189.

Ren Z, Zhang R, Li Y, *et al* (2017). Ferulic acid exerts neuroprotective effects against cerebral ischemia/reperfusion-induced injury via antioxidant and anti-apoptotic mechanisms in vitro and in vivo. *Int J Mol Med.* 40; 1444-1456.

Rentschler G, Covolo L, Haddad AA, *et al* (2012). ATP132A2 (PARK9) polymorphisms influence the neurotoxic effects of manganese. *Neurotoxicology*. 33(4); 697-702.

Riemensperger T, Isabel G, Coulom H, Neuser K, *et al* (2011). Behavioral consequences of dopamine deficiency in the *Drosophila* central nervous system. *Proc Natl Acad Sci U S A*. 108(2);834-8399.

Ritz BR, Manthripragada AD, Costello S, et al (2009). Dopamine transporter genetic variants and pesticides in Parkinson's disease. Environ Health Perspect. 117(6); 964-969.

Rodriguez-Rocha H, Garcia Garcia A, Zavala-Flores L, *et al* (2012). Glutaredoxin 1 protects dopaminergic cells by increased protein glutathionylation in experimental Parkinson's disease. *Antioxid Redox Signal*. 17; 1676–1693.

Rojas P, Serrano-García N, Mares-Sámano JJ, *et al* (2008). EGb761 protects against nigrostriatal dopaminergic neurotoxicity in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinsonism in mice: role of oxidative stress. *Eur J Neurosci.* 28; 41-50.

Rosen DR, Siddique T and Patterson D, *et al* (1993). Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature*. 362; 59–62.

Ross CA, Tabrizi SJ (2011). Huntington's disease: from molecular pathogenesis to clinical treatment. *Lancet Neurol.* 10(1); 83-98.

Rowland L (1994). Amyotrophic lateral sclerosis: theories and therapies. J Neorol Sci. 31(169);126-127.

Rubin GM, Yandell MD, Wortman JR, et al (2000). Comparative Genomics of the Eukaryotes. Science. 287: 2204–2215.

Saleem TZ, Higginson IJ, Chaudhuri KR, *et al* (2013). Symptom prevalence, severity and palliative care needs assessment using the Palliative Outcome Scale: A cross-sectional study of patients with Parkinson's disease and related neurological conditions. *Palliat Med*.27(8);722-731.

Salvi RM, Lara DR, Ghisolfi ES, *et al* (2003). Neuropsychiatric evaluation in subjects chronically exposed to organophosphate pesticide. *Toxicol Sci*.72(2); 267-271.

Samaranch L, Lorenzo-Betancor O, Arbelo JM, *et al* (2013). PINK1-linked parkinsonism is associated with Lewy body pathology. *Brain*. 133;1128–1142.

Sanyal J, Bandyopadhyaysk, Banerjee TK (2009). Plasma levels of lipid peroxides in patients with Parkinson's disease. *Eur Rev Med Pharmacol Sci.* 13(2); 129-32.

Satake W, Nakabayashi Y, Mizuta I, et al (2009). Genome-wide association study identifies common variants at four loci as genetic risk factors for Parkinson's disease. Nat Genet. 41;1303-1307.

Satapathy P, Salim C, M Naidu M, et al (2016). Attenuation of Dopaminergic Neuronal Dysfunction in *Caenorhabditis elegans* by Hydrophilic Form of Curcumin. J Neurosci Neuropharmacol. 2; 111.

Sathasivam K, Lane A, LegleiterJ, *et al* (2010). Identical oligomeric and fibrillar structures captured from the brains of R6/2 and knock-in mouse models of Huntington's disease. *Hum Mol Genet*. 19;65–78.

Schneider SA, Coro Paisan-Ruiz, Niall P, et al (2010). ATP13A2 mutations (PARK9) cause neurodegeneration with brain iron accumulation. *Mov Disord*, 25; 979-984.

Seidler A, Hellenbrand W, Robra BP, *et al* (1996). Possible environmental, occupational, and other etiologic factors for Parkinson's disease: a case-control study in Germany. *Neurology*. 46(5); 1275-1284.

Sejvar JJ, Holman RC, BreseeJS, *et al* (2005). Amyotrophic lateral sclerosis mortality in the United States, 1979-2001. *Neuroepidemiology*.25(3); 144-152.

Selkoe DJ (1998). The cell biology of beta-amyloid precursor protein and presenilin in Alzheimer's disease. *Trends Cell Biol.* 11; 447-53.

Seo SW, Bae GS and Kim SG, et al (2011). Protective effects of Curcuma longa against cerulein-induced acute pancreatitis and pancreatitis-associated lung injury. Int J Mol Med. 27(1);53-61.

Sesti F, Liu S, Cai SQ (2010). Oxidation of potassium channels by ROS: a general mechanism of aging and neurodegeneration. *Trends Cell Biol*. 20(1); 45-51.

Sharma C, Suhalka P, Sukhwal P (2014). Curcumin attenuates neurotoxicity induced by fluoride: An in vivo evidence. *Pharmacogn Mag.* 10(37); 61–65.

Sharma R, Yang Y, Sharma A, *et al* (2004). Antioxidant role of glutathione s transferase: protection against oxidant toxicity and regulation of stress-mediated apoptosis. *Antioxid Redox Signal.* 6(2); 289-300.

Sheerin UM, Houlden H, Wood NW. (2014). Advances in the genetics of Parkinson's disease: A guide for the clinician. *Mov Disord Clin Pract.* 1; 3-13.

Shimizu K, Ohtaki K, Matsubara K, et al (2001). Carrier-mediated processes in blood--brain barrier penetration and neural uptake of paraquat. Brain Res. 906(1-2):135-42.

Shimouchi A, Nose K, Takaoka M, et al (2008). Effect of dietary turmeric on breath hydrogen. Dig Dis Sci. 54(8);1725–1729.

Shinotoh H, Fukushi K, Nagatsuka S, *et al* (2003). The amygdala and Alzheimer's disease: positron emission tomographic study of the cholinergic system. *Ann N Y Acad Sci.* 985; 411-419.

Shirasaki DI, Greiner ER, Al-Ramahi I, et al (2012). Network organization of the huntingtin proteomic interactome in mammalian brain. Neuron. 75(1); 41-57.

Shojaee S, Sina F, Banihosseini SS, *et al* (2008). Genome-wide linkage analysis of a Parkinsonian-pyramidal syndrome pedigree by 500 K SNP arrays. *Am J Hum Genet*. 82;1375–1384.

Shungua D, Maoa X, Weiduschat N, *et al* (2017). Nigrostriatal glutathione deficit in Parkinson's disease measured in vivo with MRS supports oxidative stress in disease pathophysiology. *J Neurol Sci.* 381;561–756

Sian J, Dexter DT, Lees AJ, et al (1994). Alterations in glutathione levels in Parkinson's disease and other neurodegenerative disorders affecting basal ganglia. Ann Neurol. 3; 348–355.

Sibley DR (1999). New insights into dopaminergic receptor function using antisense and genetically altered animals. *Annu Rev PharmacolToxicol*. 39;313–341.

Siddique YH, Naz F, Jyoti S (2014). Effect of Curcumin on Lifespan, Activity Pattern, Oxidative Stress, and Apoptosis in the Brains of Transgenic *Drosophila* Model of Parkinson's Disease. *Bio Med Res Int.* 2014;606928.

Sies H (2015). Oxidative stress: a concept in redox biology and medicine. Redox Biol. 4; 180-183.

Simon-Sanchez J, Schulte C, Bras J (2009). Genome-wide association study reveals genetic risk underlying Parkinson's disease. *Nat Genet*. 41;1308-1312.

Simuni T, Luo ST, Chou KL, *et al* (2013). Rankin scale as a potential measure of global disability in early Parkinson's disease. *J Clin Neurosci*. 20(9); 1200-3.

Singh PK, Kotia V, Ghosh D, *et al* (2013). Curcumin modulates α-synuclein aggregation and toxicity.*ACS Chem Neurosci*.4(3);393-407.

Singh S, Kushwah AS, Singh R, *et al* (2012). Current therapeutic strategy in Alzheimer's disease. *Eur Rev Med Pharmacol Sci.* 16(12);1651-1664.

Singhal A, VB Morris, V Labhasetwar, et al (2013). Nanoparticle-mediated catalase delivery protects human neurons from oxidative stress. *Cell Death Dis.* 4, e903; doi:10.1038/cddis.2013.36.

Smeyne M, Smeyne RJ (2013). Glutathione metabolism and Parkinson's disease. *Free Radic Biol Med.* 62; 13-25.

Smith DG, Cappai R, Barnham KJ (2007). The redox chemistry of the Alzheimer's disease amyloid beta peptide. *BiochemBiophys Acta*. 1768;1976-90.

Soares JJ, Rodrigues DT, Goncalves MB, *et al* (2017). Paraquat exposure-induced Parkinson's disease-like symptoms and oxidative stress in *Drosophila melanogaster*: Neuroprotective effect of Bougainvillea glabra choisy. *Biomed Parmacother*. 95; 245-251.

Soh JW, Marowsky N, Nichols TJ, et al (2013). Curcumin is an early-acting stage-specific inducer of extended functional longevity in *Drosophila*. *Exp Gerontol*. 48(2);229-239.

Sohal RS, Mockett RJ, Orr WC (2002). Mechanisms of aging: an appraisal of the oxidative stress hypothesis. *Free Radic Biol Med.* 33(5); 575–586.

Sokolowski MB (2001). Drosophila: genetics meets behaviour. Nat Rev Genet.2(11);879-890.

Somani SM, Ravi R, Rybak LP (1995). Effect of exercise training on antioxidant system in brain region of rat. *Pharmacol Biochem Behav.* 50 (4): 635–639.

Son OL, Kim HT, Ji MH, *et al* (2003). Cloning and expression analysis of a Parkinson's disease gene, uch-L1, and its promoter in zebrafish. *Biochem Biophys Res Commun.* 312(3); 610-607.

Spillantini MG, Crowther RA, Jakes R, *et al* (1998). Alpha synuclein in filamentous inclusions of lewy bodies from Parkinson's disease and dementia with lewy bodies. *Proc Natl Acad Sci USA*. 95(11); 6469-6473.

Steenland K, MacNeil J, Seals R, *et al* (2010). Factorsaffecting survival of patients with neurodegenerative disease. *Neuroepidemiology*. 35(1);28-35.

Stelmashook EV, Isaev NK, Zorov DB (2007). Paraquat potentiates glutamate toxicity in immature cultures of cerebellar granule neurons. *Toxicol Lett.* 174(1-3);82-88.

Stephens R, Spurgeon A, Calvert IA, *et al* (1995). Neuropsychological effects of long-term exposure to organophosphates in sheep dip. *Lancet.* 345; 1135-1139.

Stork T, Engelen D, Krudewig A, *et al* (2008). Organization and function of the blood-brain barrier in Drosophila. *J Neurosci*. 28(3); 587-597.

Strauss KM, Martins LM, Plun-Favreau H, et al (2005). Loss of function mutations in the gene encoding Omi/HtrA2 in Parkinson's disease. Hum Mol Genet. 14; 2099-2111.

Sudati JH, Vieira FA, Pavin SS, et al (2013). Valeriana officinalis attenuates the rotenone-induced toxicity in Drosophila melanogaster. *Neurotoxicology*. 37;118-26.

Tadolini, Juliano C, Piu L, et al (2000). Resveratrol inhibition of lipid peroxidation. J. FreeRadic Res. 33;105-114.

Tan CC, Yu JT, Tan MS, *et al* (2014). Autophagy in aging and neurodegenerative diseases: implications for pathogenesis and therapy. *Neurobiol Aging*. 35(5);941-957.

Tang M, Larson-Meyer DE, Liebman M (2008). Effect of cinnamon and turmeric on urinary oxalate excretion, plasma lipids, and plasma glucose in healthy subjects. *Am J Clin Nutr.* 87(5); 1262-1267.

Tanner CM, Kamel F, Ross GW, et al (2011). Rotenone, Paraquat, and Parkinson's Disease. Environ Health Perspect. 119(6): 866–872.

Taylor JM, Main BS, Crack PJ (2012). Neuroinflammation and oxidative stress: co-conspirators in the pathology of Parkinson's disease. *Neurochem Int.* 62 (2013): 803–819.

Thapliyal R and Maru GB (2001). Inhibition of P450 isoenzymes by curcumin in vitro and in vivo. *Food Chem Toxicol*. 39(6); 541-547.

Thiruchelvam MJ, Powers JM, Cory-Slechta DA, *et al* (2004). Risk factors for dopaminergic neuron loss in human alpha-synuclein transgenic mice. *Eur J Neurosci.* 19(4); 845-854.

Tizabi Y, Hurley LL, Qualls Z, *et al* (2014). Relevance of the anti-inflammatory properties of curcumin in neurodegenerative diseases and depression. *Molecules*. 19(12); 20864-20879.

Toda S, Miyase T, Arichi H, *et al* (1985). Natural antioxidants. III. Antioxidative components isolated from rhizome of Curcuma longa L. *Chem Pharm Bull (Tokyo)*.33 (4); 1725-8.

Trabucchi M, Cheney DL, Racagni G, *et al* (1975) In vivo inhibition of striatal acetylcholine turnover by L-DOPA, apomorphine and (1) amphetamine. *Brain Res.* 85; 30–134.

Tripanichkul W, Jaroensuppaperch EO (2013). Ameliorating effects of curcumin on 6-OHDA-induced dopaminergic denervation, glial response, and SOD1 reduction in the striatum of hemiparkinsonian mice. *Eur Rev Med Pharmacol Sci.*17 (10); 1360-8.

Tripathi A, Srivastava UC (2008). Acetylcholinesterase: A Versatile Enzyme of Nervous System. *Annals Neurosci.* 15(4); 106-11.

Trujillo J, Chirino YI, Molina-Jijón E, et al (2013). Renoprotective effect of the antioxidant curcumin: Recent findings. *Redox Biol*.1; 448-456.

Tucci A, Nalls MA, Houlden H, et al (2010). Genetic variability at the PARK16 locus. Eur J Hum Genet. 18(12); 1356-1359.

Turmeric For Health (2018). 8-popular-curcumin-supplement-types-in-market-today-a-quick-review.

Turnbull S, Tabner BJ, El-Agnaf OM, *et al* (2001). alpha-Synuclein implicated in Parkinson's disease catalyses the formation of hydrogen peroxidein vitro. *Free Radic Biol Med.* 30(10); 1163-70.

Urbina-Cano P, Morales BL, Herrera MA (2006). DNA damage in mouse lymphocytes exposed to curcumin and copper. *J Appl Genet*. 7;377–382.

Valente EM, Abou-Sleiman PM, Caputo V, *et al* (2004). Hereditary early-onset Parkinson's disease caused by mutations in PINK1. *Science*. 304; 1158–1160.

Van der Merwe C, van Dyk HC, Engelbrecht L, *et al* (2017). Curcumin Rescues a PINK1 Knock Down SH-SY5Y Cellular Model of Parkinson's Disease from Mitochondrial Dysfunction and Cell Death.*MolNeurobiol.* 54(4); 2752-2762.

Van Vliet SA, Vanwersch RA, Jongsma MJ, *et al* (2006). Neuroprotective effect of modafinil in a marmoset Parkinson model: behavioural and neurochemical aspects. *Behav Pharmacol*. 17(5-6); 453-462.

Venken KJ, Bellen HJ (2007). Transgenesis upgrades for *Drosophila melanogaster*. *Development*. 134;3571–3584.

Vilarino-Guell C, Wider C, Ross OA, et al (2011). VPS35 mutations in Parkinson's disease. Am J Hum Genet. 89(1); 162-167.

Von Coelln R, Thomas B, Andrabi SA, *et al* (2006). Inclusion body formation and neurodegeneration are parkin independent in a mouse model of alpha-synucleinopathy. *J Neurosci*. 26(14); 3685-3696.

Walsh DM, Selkoe DJ (2007). Aβ Oligomers – a decade of discovery. J Neurochem. 101; 1172–1184.

Wang MC, Bohmann D, Jasper H (2003). JNK Signaling Confers Tolerance to Oxidative Stress and Extends Lifespan in *Drosophila*. *Dev Cell*. 5; 811–816.

Wang P, Su C, Li R, et al (2014). Mechanisms and effects of curcumin on spatial learning and memory improvement in APPswe/PS1dE9 mice. J Neurosci Res. 92(2); 218-31.

Wang X (2001). The expanding role of mitochondria in apoptosis. Genes Dev. 15(22); 2922-2933.

Wang YL, Ju B, Zhang YZ, *et al* (2017). Protective effect of curcumin against oxidative stress-induced injury in rats with Parkinson's disease through the Wnt/ β -catenin signaling pathway. *Cell Physiol Bioche*. 43(6); 2226-2241.

Warby SC, Visscher S and Collins JA, *et al* (2011). HTT haplotypes contribute to differences in Huntington disease prevalence betweenEurope and East Asia. *EurJ Hum Genet*. 19; 561–566.

Wheeler MR. 1986. Additions to the catalog of the world's Drosophilidae. In the Genetics and Biology of Drosophila. Vol 3e. Eds. Ashburner M, Carson HL and Thompson Jr. JN. Academic press. London 395-409.

White HL, Scates PW, Cooper BR (1996). Extracts of Ginkgo biloba leaves inhibit monoamine oxydase. *Life Sci.* 58;1315–21.

White KE, HumphreyDM, Hirth F (2010). The dopaminergic system in the aging brain of *Drosophila*. Front Neurosci. 4(205); 1-12.

Whitworth AJ, Lee JR, Ho VM, *et al* (2008). Rhomboid-7 and HtrA2/Omi act in a common pathway with the Parkinson's disease factors Pink1 and parkin. *Dis Model Mech.* 1;168-174.

Widdowson PS, Farnworth MJ, Upton R, *et al* (1996). No changes in behaviour, nigro-striatal system neurochemistry or neuronal cell death following toxic multiple oral paraquat administration to rats. *Hum Exp Toxicol.* 15(7);583-591.

Wilk JB, Tobin JE, Suchowersky O, *et al* (2006). Herbicide exposure modifies GSTP1 haplotype association to Parkinson onset age. *Neurology*. 67(12); 2206-2210.

Williams AJ, PaulsonHL (2008). Polyglutamine neurodegeneration: protein misfolding revisited. *Trends Neurosci*.31;521-528.

Witjas T, Kaphan E, Azulay JP, et al (2002). Nonmotor fluctuations in Parkinson's disease: frequent and diabling. Neurology. 59(3); 408-413.

Wolff PS (1994). Methods in enzymology. In: Packer L (ed). Academic, New York. 233; 182-189.

Wu J and Bauer CE (2008). RegB/RegA, a global redox-responding two-component system. *Adv Exp Med Biol.* 631; 131–148.

Yang C, Ma X, Wang Z, *et al* (2017). Curcumin induces apoptosis and protective autophagy in castration-resistant prostate cancer cells through iron chelation. *Drug Des DevelTher*. 11;431-439.

Yang F, Lim GP, Begum AN, et al (2005). Curcumin inhibits formation of amyloid beta oligomers and fibrils, binds plaques, and reduces amyloid in vivo. J Biol Chem. 280(7);5892-5901.

Yang Z, Zhao T, Zou Y, et al (2014). Curcumin inhibits microglia inflammation and confers neuroprotection in intracerebral hemorrhage. Immunol Lett. 160(1); 89-95.

Yoshino H, Tomiyama H, Tachibana N, *et al* (2010). Phenotypic spectrum of patients with PLA2G6 mutation and PARK14-linked parkinsonism. *Neurology*. 75;1356-1361.

Yuan Y, Tong Q, Zhang L, *et al* (2016). Plasma antioxidant status and motor features in de novo Chinese Parkinson's disease patients. *Int J Neurosci.* 126; 641–646.

Yun J, Cao JH, Dodson MW, *et al* (2008). Loss of function analysis suggests that Omi/HtrA2 is not an essential component of the PINK1/PARKIN pathway in vivo. *J Neurosci.* 283;14500-14510.

Zarse K, Schmeisser S, Groth M, et al (2012). Impaired insulin/IGF1 signaling extends life span by promoting mitochondrial L-proline catabolism to induce transient ROS signal. Cell Metab. 15(4); 451-465.

Zeevalk GD, Razmpour R, Bernard LP (2008). Glutathione and Parkinson's disease: is this the elephant in the room?. *Biomed Pharmacother*. 62(4); 236-49.

Zelko IN, Mariani TJ, Folz RJ (2002). Superoxide dismutase multigene family: a comparison of the CuZn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) gene structures, evolution, and expression. *Free Radic Biol Med.* 33; 337–349.

Zhang ZG, Niu XY, Lu AP, et al (2015). Effect of curcumin on aged Drosophila melanogaster: a pathway prediction analysis. Chin J Integr Med. 21(2); 115-22.

Zhao H, Song H, Chai OH (2012). Negative Effects of Curcumin on Liver Injury Induced by Alcohol. *Phytother Res.* 16(12);1857-1863.

Zhou C, Huang Y, Przedborski S (2008). Oxidative stress in Parkinson's disease: a mechanism of pathogenic and therapeutic significance. *Ann NY Acad Sci.* 1147; 93–104.

Zhu Y, Carvey PM, Ling Z (2007). Altered glutathione homeostasis in animals prenatally exposed to lipopolysaccharide. *Neurochem Int.* 50(4); 671-680.

Zimprich A, Benet-Pages A, Struhal W, *et al* (2011). A mutation in VPS35, encoding a subunit of the retromer complex, causes late onset of Parkinson's disease. *Am J Hum Genet*. 89(1); 168-175.

Zimprich A, Biskup S, Leitner P, *et al* (2004). Mutations in LRRK2 cause autosomal-dominant parkinsonism with pleomorphic pathology. *Neuron*. 44;601–607.

Zrinzo L, Foltynie T, Limousin P, *et al* (2012). Reducing hemorrhagic complications in functional neurosurgery: a large case series and systematic literature review. *J Neurosurg*. 116(1); 84-94.

CONFERENCE/SEMINAR/WORKSHOP ATTENDED

- Presented a paper on 'Understanding Neurodegeneration and Rescuing Pathology Associated with Parkinson's Disease in *Drosophila* Model' during Workshop on Introduction to Basic and Advanced Biomedical Approaches for Enhancing Quality Of Life in Aging Societies. Advanced Industrial Science and Technology Biomedical Research Institute (AIST), Tsukuba, Japan. 14-21st October, 2018.
- Presented a paper titled, 'Curcumin Improves Paraquat Induced Mobility Defects During Health Span and Modulates Brain Nitrite Levels in *Drosophila* Model of Parkinson's Disease' at National Seminar On Climate Change and Sustainable Development with Special Focus on North East India. Nagaland University, Nagaland, India. 17-18th May, 2017.
- Presented a poster titled, 'Curcumin's Neuroprotective Efficacy in *Drosophila* Model of Idiopahic Parkinson's Disease is Phase Specific: Implications of its Therapeutic Effectiveness' at International Conference on Parkinson's Disease and Movement Disorder. Frankfurt, Germany. 11 -13th August, 2015.
- Presented a poster titled, 'Curcumin Improves Paraquat Induced Mobility Defects During Health Span and Transition Phase in *Drosophila* Model of Parkinson's Disease' at National Seminar on Metabolomics: A New Frontier in Natural Products Research. NEHU, Meghalaya, India. 23-24th May, 2014.
- Abstract accepted titled, 'Understanding the Neuroprotective Properties of Curcumin During Health and Transition Stage: Insights from *Drosophila* Model of Parkinson's Disease' for National Conference on Contemporary Excitement in New Biology. Department of Zoology, Nagaland University, India. 30-31st October, 2018.

HONORS AND AWARDS

- Awarded with Department of Biotechnology (DBT), New Delhi Junior Research Fellowship and Senior Research Fellowship, 2012-2015.
- Awarded with Indian Council of Medical Research (ICMR), New Delhi Young Scientist International Travel Grant, 2015.
- Awarded with Indian Council of Medical Research (ICMR), New Delhi Senior Research Fellowship (not availed), 2018.
- Selected for Japan-Asia Youth Exchange Program in Science (Sakura Exchange Program in Science), 2018, through the Department of Biotechnology (DBT), New Delhi.

LIST OF PUBLICATIONS

- Phom L, Achumi B, Alone D, Muralidhara, and Yenisetti SC (2014). Curcumin's Neuroprotective Efficacy in *Drosophila* Model of Idiopathic Parkinson's Disease is Phase Specific: Implication of its Therapeutic Effectiveness. *Rejuvenation Research*. 17(6); 481-9.
- Modi P, Ayajuddin M, Phom L, Koza Z, Das A, Chaurasia R, Samadder S, Achumi B, Muralidhara, Pukhrambam RS, Yenisetti SC (2016). Understanding Pathophysiology of Sporadic Parkinson's Disease in *Drosophila* Model: Potential Opportunities and Notable Limitations. In Challenges in Parinson's Disease. Ed. Jolanta Dorszewska. *Intech Open.* 11; 217-44.
- Pukhrambam PR, Thepa A, Jamir N, Phom L, Yenisetti SC (2017). Parkinson's Disease and Therapeutic Strategies. *Int J of Neurology and Neurosurgery*. 9(2); 172-86.
- Pukhrambam PR, Koza Z, Phom L, Lal P, Yenisetti SC (2017). Parkinson's Disease
 An Overview. Fazl Ali College Journal. 7;1-17.
- Ayajuddin M, Das A, Phom L, Modi P, Chaurasia R, Koza Z, Thepa A, Jamir N, Singh PR, Longkumer S, Lal P, Yenisetti SC (2018). Parkinson's Disease: Insights from *Drosophila* Model. In *Drosophila melanogaster*: Model for Recent Advances in Genetics and Therapeutics. Ed. Farzana Khan Perveen. *Intech Open*. 8;157-92.
- Sub-Editor. Dopamine Health and Disease (In Press). *Intech Open*. ISBN 978-953-51-6729-7.

PUBLICATIONS THROUGH COLLABORATION WITH OTHER LAB MEMBERS

- Achumi B, Phom L, Zevelou, Ayajuddin M, Lal P, Yenisetti SC (2014). Ecogeopraphic Pattern of Genus *Drosophila* (Insecta, Diptera: Drosophilidae) in Nagaland State, India. *Nagaland University Research Journal*. 7; 320-37.
- Achumi B, Ayajuddin M, Phom L, Hegde SN, Lal P, Singh OP, Yenisetti SC (2016). Drosophilid (Insecta, Diptera: Drosophilidae) Biodiversity of North-East India. *Bioprospecting of Indigenous Bioresources of North East India*. 14; 231-51.